

Using the **SRADB** Package to Query the Sequence Read Archive

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1 Introduction

High throughput sequencing technologies have very rapidly become standard tools in biology. The data that these machines generate are large, extremely rich. As such, the Sequence Read Archives (SRA) have been set up at NCBI in the United States, EMBL in Europe, and DDBJ in Japan to capture these data in public repositories in much the same spirit as MIAME-compliant microarray databases like NCBI GEO and EBI ArrayExpress.

Accessing data in SRA requires finding it first. This R package provides a convenient and powerful framework to do just that. In addition, **SRADB** features functionality to determine availability of sequence files and to download files of interest.

SRA currently store aligned reads or other processed data that relies on alignment to a reference genome. Please refer to the SRA handbook (<http://www.ncbi.nlm.nih.gov/books/NBK47537/>) for details. NCBI GEO also often contain aligned reads for sequencing experiments and the **SRADB** package can help to provide links to these data as well. In combination with the **GEOmetadb** and **GEOquery** packages, these data are also, then, accessible.

2 Getting Started

Since SRA is a continuously growing repository, the **SRADB** SQLite file is updated regularly. The first step, then, is to get the **SRADB** SQLite file from the online location. The download and uncompress steps are done automatically with a single command, `getSRADBFile`.

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Figure 1: A graphical representation (sometimes called an *Entity-Relationship Diagram*) of the relationships between the main tables in the SRAdB package.

```
> library(SRAdb)
> sqlfile <- 'SRAmetadb.sqlite'
> if(!file.exists('SRAmetadb.sqlite')) sqlfile <- getSRADBFile()
```

The default storage location is in the current working directory and the default filename is “SRAmetadb.sqlite”; it is best to leave the name unchanged unless there is a pressing reason to change it. Note: the above downloading and uncompressing steps could take quite a few moments due to file size, depending on your network bandwidth. If interested, it can be timed using the following commands:

```
> timeStart <- proc.time()
> sqlfile <- getSRADBFile()
> proc.time() - timeStart
```

Since this SQLite file is of key importance in SRAdb, it is perhaps of some interest to know some details about the file itself.

```
> file.info('SRAmetadb.sqlite')

              size isdir mode
SRAmetadb.sqlite 18905319424 FALSE  644
              mtime
SRAmetadb.sqlite 2016-05-03 19:37:38
              ctime
SRAmetadb.sqlite 2016-05-03 19:37:38
              atime uid gid
SRAmetadb.sqlite 2016-05-03 19:37:39 501  20
              uname grname
SRAmetadb.sqlite biocbuild  staff
```

Then, create a connection for later queries. The standard DBI functionality as implemented in RSQLite function `dbConnect` makes the connection to the database. The `dbDisconnect` function disconnects the connection.

```
> sra_con <- dbConnect(RSQLite(), sqlfile)
```

For further details, at this time see `help('SRAdb-package')`.

3 Using the SRAdb package

3.1 Interacting with the database

The functionality covered in this section is covered in much more detail in the DBI and RSQLite package documentation. We cover enough here only to be useful. The `dbListTables` function lists all the tables in the SQLite database handled by the connection object `sra_con` created in the previous section. A simplified illustration of the relationship between the SRA main data types is shown in the Figure 1.

```
> sra_tables <- dbListTables(sra_con)
> sra_tables

[1] "col_desc"          "experiment"
[3] "fastq"             "metaInfo"
[5] "run"               "sample"
[7] "sra"               "sra_ft"
[9] "sra_ft_content"    "sra_ft_segdir"
[11] "sra_ft_segments"   "study"
[13] "submission"
```

There is also the `dbListFields` function that can list database fields associated with a table.

```
> dbListFields(sra_con, "study")

[1] "study_ID"           "study_alias"
[3] "study_accession"    "study_title"
[5] "study_type"         "study_abstract"
[7] "broker_name"        "center_name"
[9] "center_project_name" "study_description"
[11] "related_studies"    "primary_study"
[13] "sra_link"           "study_url_link"
[15] "xref_link"          "study_entrez_link"
[17] "ddbj_link"          "ena_link"
[19] "study_attribute"    "submission_accession"
[21] "sradb_updated"
```

Sometimes it is useful to get the actual SQL schema associated with a table. Here, we get the table schema for the *study* table:

```
> dbGetQuery(sra_con, 'PRAGMA TABLE_INFO(study)')
```

	cid	name	type	notnull
1	0	study_ID	REAL	0
2	1	study_alias	TEXT	0
3	2	study_accession	TEXT	0
4	3	study_title	TEXT	0
5	4	study_type	TEXT	0
6	5	study_abstract	TEXT	0
7	6	broker_name	TEXT	0
8	7	center_name	TEXT	0
9	8	center_project_name	TEXT	0
10	9	study_description	TEXT	0

11	10	related_studies	TEXT	0
12	11	primary_study	TEXT	0
13	12	sra_link	TEXT	0
14	13	study_url_link	TEXT	0
15	14	xref_link	TEXT	0
16	15	study_entrez_link	TEXT	0
17	16	ddbj_link	TEXT	0
18	17	ena_link	TEXT	0
19	18	study_attribute	TEXT	0
20	19	submission_accession	TEXT	0
21	20	sradb_updated	TEXT	0
		dflt_value	pk	
1		<NA>	0	
2		<NA>	0	
3		<NA>	0	
4		<NA>	0	
5		<NA>	0	
6		<NA>	0	
7		<NA>	0	
8		<NA>	0	
9		<NA>	0	
10		<NA>	0	
11		<NA>	0	
12		<NA>	0	
13		<NA>	0	
14		<NA>	0	
15		<NA>	0	
16		<NA>	0	
17		<NA>	0	
18		<NA>	0	
19		<NA>	0	
20		<NA>	0	
21		<NA>	0	

The table "col_desc" contains information of filed name, type, description and default values:

```
> colDesc <- colDescriptions(sra_con=sra_con)[1:5,]
> colDesc[, 1:4]
```

	col_desc_ID	table_name	field_name
1	1	submission	ID
2	2	submission	accession
3	3	submission	alias

```

4          4 submission submission_comment
5          5 submission                files
      type
1      int
2 varchar
3 varchar
4      text
5      text

```

3.2 Writing SQL queries and getting results

Select 3 records from the *study* table and show the first 5 columns:

```

> rs <- dbGetQuery(sra_con,"select * from study limit 3")
> rs[, 1:3]

```

```

      study_ID study_alias study_accession
1          1    DRP000001    DRP000001
2          2    DRP000002    DRP000002
3          3    DRP000003    DRP000003

```

Get the SRA study accessions and titles from SRA study that study_type contains “Transcriptome”. The “%” sign is used in combination with the “like” operator to do a “wildcard” search for the term “Transcriptome” with any number of characters after it.

```

> rs <- dbGetQuery(sra_con, paste( "select study_accession,
+      study_title from study where",
+      "study_description like 'Transcriptome%'",sep=" "))
> rs[1:3,]

```

```

      study_accession
1      ERP000233
2      ERP000350
3      ERP000527

```

```

1 Identification of the expression profile of Staphylococcus aureus grown in the presence of
2
3

```

Transcriptome Analysis of the

Of course, we can combine programming and data access. A simple `sapply` example shows how to query each of the tables for number of records.

```

> getTableCounts <- function(tableName,conn) {
+   sql <- sprintf("select count(*) from %s",tableName)

```

```
+ return(dbGetQuery(conn,sql)[1,1])
+ }
> do.call(rbind,sapply(sra_tables[c(2,4,5,11,12)],
+      getTableCounts, sra_con, simplify=FALSE))
```

```
      [,1]
experiment 1832510
metaInfo    2
run         2264680
sra_ft_segments 584856
study       69466
```

Get some high-level statistics could be to helpful to get overall idea about what data are available in the SRA database. List all study types and number of studies contained for each of the type:

```
> rs <- dbGetQuery(sra_con, paste( "SELECT study_type AS StudyType,
+      count( * ) AS Number FROM `study` GROUP BY study_type order
+      by Number DESC ", sep=""))
> rs
```

	StudyType	Number
1	Whole Genome Sequencing	29292
2	Other	21796
3	Transcriptome Analysis	9887
4	Metagenomics	6346
5	Population Genomics	773
6	Epigenetics	649
7	<NA>	389
8	Exome Sequencing	185
9	Cancer Genomics	107
10	Pooled Clone Sequencing	31
11	Synthetic Genomics	9
12	Transcriptome Sequencing	1
13	Whole Genome Sequencing	1

List all Instrument Models and number of experiments for each of the Instrument Models:

```
> rs <- dbGetQuery(sra_con, paste( "SELECT instrument_model AS
+      'Instrument Model', count( * ) AS Experiments FROM `experiment`
+      GROUP BY instrument_model order by Experiments DESC", sep=""))
> rs
```

	Instrument Model
1	Illumina HiSeq 2000
2	Illumina MiSeq
3	Illumina HiSeq 2500
4	454 GS FLX Titanium
5	Illumina Genome Analyzer II
6	<NA>
7	Illumina Genome Analyzer IIx
8	unspecified
9	454 GS FLX
10	Illumina Genome Analyzer
11	Ion Torrent PGM
12	454 GS Junior
13	NextSeq 500
14	AB SOLiD 4 System
15	Illumina HiSeq 1000
16	454 GS FLX+
17	PacBio RS II
18	HiSeq X Ten
19	PacBio RS
20	454 GS
21	Helicos HeliScope
22	Complete Genomics
23	Illumina HiSeq 1500
24	AB 5500xl Genetic Analyzer
25	AB SOLiD System 3.0
26	AB 5500 Genetic Analyzer
27	Illumina HiScanSQ
28	Ion Torrent Proton
29	454 GS 20
30	AB 3730xL Genetic Analyzer
31	AB SOLiD System 2.0
32	MinION
33	AB SOLiD System
34	Illumina HiSeq 4000
35	AB SOLiD 3 Plus System
36	AB SOLiD 4hq System
37	AB 5500xl-W Genetic Analysis System
38	AB 3130 Genetic Analyzer
39	Illumina HiSeq 3000
40	AB 3130xL Genetic Analyzer
41	AB 3730 Genetic Analyzer

42	Illumina NextSeq 500
43	NextSeq 550
44	454 GS FLX
45	AB 3500xL Genetic Analyzer
46	HiSeq X Five
47	AB SOLiD PI System
48	Illumina Genome Analyzer IIX
49	AB 310 Genetic Analyzer
50	AB 3500 Genetic Analyzer

Experiments

1	904632
2	220233
3	164222
4	111837
5	100599
6	78358
7	50931
8	36333
9	31440
10	17799
11	15007
12	14900
13	10502
14	10141
15	9377
16	8954
17	6634
18	5623
19	5525
20	5218
21	3831
22	3165
23	2800
24	2668
25	2511
26	1811
27	1774
28	1641
29	980
30	610
31	466
32	445

33	442
34	303
35	218
36	158
37	119
38	101
39	84
40	33
41	22
42	18
43	15
44	10
45	9
46	5
47	2
48	2
49	1
50	1

List all types of library strategies and number of runs for each of them:

```
> rs <- dbGetQuery(sra_con, paste( "SELECT library_strategy AS
+      'Library Strategy', count( * ) AS Runs FROM `experiment`
+      GROUP BY library_strategy order by Runs DESC", sep=""))
> rs
```

	Library Strategy	Runs
1	WGS	600917
2	AMPLICON	316851
3	RNA-Seq	271862
4	WXS	228705
5	OTHER	148922
6	<NA>	78358
7	ChIP-Seq	50506
8	POOLCLONE	50143
9	SELEX	21657
10	Bisulfite-Seq	13457
11	miRNA-Seq	10052
12	CLONE	9299
13	WGA	8212
14	EST	3475
15	VALIDATION	3284
16	RAD-Seq	2031
17	ncRNA-Seq	1961

18	MNase-Seq	1582
19	MeDIP-Seq	1574
20	DNase-Hypersensitivity	1545
21	FL-cDNA	1489
22	Tn-Seq	1370
23	RIP-Seq	1159
24	MRE-Seq	1111
25	MBD-Seq	1090
26	Targeted-Capture	554
27	WCS	514
28	CLONEEND	327
29	CTS	186
30	FAIRE-seq	174
31	Hi-C	78
32	ChIA-PET	29
33	FINISHING	24
34	Synthetic-Long-Read	12

3.3 Conversion of SRA entity types

Large-scale consumers of SRA data might want to convert SRA entity type from one to others, e.g. finding all experiment accessions (SRX, ERX or DRX) and run accessions (SRR, ERR or DRR) associated with "SRP001007" and "SRP000931". Function `sraConvert` does the conversion with a very fast mapping between entity types.

Covert "SRP001007" and "SRP000931" to other possible types in the `SRAMetadb.sqlite`:

```
> conversion <- sraConvert( c('SRP001007', 'SRP000931'), sra_con = sra_con )
> conversion[1:3,]
```

```
      study submission      sample experiment
1 SRP000931  SRA009053  SRS003453  SRX006122
2 SRP000931  SRA009053  SRS003458  SRX006127
3 SRP000931  SRA009053  SRS003462  SRX006133

      run
1 SRR018256
2 SRR018261
3 SRR018267
```

Check what SRA types and how many entities for each type:

```
> apply(conversion, 2, unique)
```

```
$study
[1] "SRP000931" "SRP001007"
```

```

$submission
[1] "SRA009053" "SRA009276"

$sample
[1] "SRS003453" "SRS003458" "SRS003462"
[4] "SRS003457" "SRS003456" "SRS003460"
[7] "SRS003461" "SRS003454" "SRS003455"
[10] "SRS003464" "SRS003459" "SRS003463"
[13] "SRS004650"

$experiment
[1] "SRX006122" "SRX006127" "SRX006133"
[4] "SRX006126" "SRX006125" "SRX006131"
[7] "SRX006132" "SRX006123" "SRX006130"
[10] "SRX006124" "SRX006135" "SRX006129"
[13] "SRX006128" "SRX006134" "SRX007396"

$run
[1] "SRR018256" "SRR018261" "SRR018267"
[4] "SRR018260" "SRR018259" "SRR018265"
[7] "SRR018266" "SRR018257" "SRR018264"
[10] "SRR018258" "SRR018269" "SRR018263"
[13] "SRR018262" "SRR018268" "SRR020739"
[16] "SRR020740"

```

3.4 Full text search

Searching by regular table and field specific SQL commands can be very powerful and if you are familiar with SQL language and the table structure. If not, SQLite has a very handy module called Full text search (fts3), which allow users to do Google like search with terms and operators. The function `getSRA` does Full text search against all fields in a fts3 table with terms constructed with the Standard Query Syntax and Enhanced Query Syntax. Please see <http://www.sqlite.org/fts3.html> for detail.

Find all run and study combined records in which any given fields has "breast" and "cancer" words, including "breast" and "cancer" are not next to each other:

```

> rs <- getSRA( search_terms = "breast cancer",
+               out_types = c('run', 'study'), sra_con )
> dim(rs)

[1] 19311    23

```

```
> rs <- getSRA( search_terms = "breast cancer",
+               out_types = c("submission", "study", "sample",
+                             "experiment", "run"), sra_con )
> # get counts for some information interested
> apply( rs[, c('run','sample','study_type','platform',
+               'instrument_model')], 2, function(x)
+       {length(unique(x))} )
```

```

           run           sample
       19311         12962
study_type      platform
         9             6
instrument_model
       28
```

```
>
```

If you only want SRA records containing exact phrase of "breast cancer", in which "breast" and "cancer" do not have other characters between other than a space:

```
> rs <- getSRA (search_terms = '"breast cancer"',
+               out_types=c('run','study'), sra_con)
> dim(rs)
```

```
[1] 15400    23
```

Find all sample records containing words of either "MCF7" or "MCF-7":

```
> rs <- getSRA( search_terms = 'MCF7 OR "MCF-7"',
+               out_types = c('sample'), sra_con )
> dim(rs)
```

```
[1] 2599    10
```

Find all submissions by GEO:

```
> rs <- getSRA( search_terms = 'submission_center: GEO',
+               out_types = c('submission'), sra_con )
> dim(rs)
```

```
[1] 11951     6
```

Find study records containing a word beginning with 'Carcino':

```
> rs <- getSRA( search_terms = 'Carcino*',
+               out_types = c('study'), sra_con=sra_con )
> dim(rs)
```

```
[1] 695    12
```

3.5 Download SRA data files

List ftp addresses of the fastq files associated with "SRX000122":

```
> rs = listSRAfile( c("SRX000122"), sra_con, fileType = 'sra' )
```

The above function does not check file availability, size and date of the sra data files on the server, but the function getSRAinfo does this, which is good to know if you are preparing to download them:

```
> rs = getSRAinfo ( c("SRX000122"), sra_con, sraType = "sra" )
> rs[1:3,]
```

```
1 ftp://ftp-trace.ncbi.nlm.nih.gov/sra/sra-instant/reads/ByExp/sra/SRX/SRX000/SRX000122/
2 ftp://ftp-trace.ncbi.nlm.nih.gov/sra/sra-instant/reads/ByExp/sra/SRX/SRX000/SRX000122/
3 ftp://ftp-trace.ncbi.nlm.nih.gov/sra/sra-instant/reads/ByExp/sra/SRX/SRX000/SRX000122/
  experiment      study      sample      run
1 SRX000122 SRP000098 SRS000290 SRR000648
2 SRX000122 SRP000098 SRS000290 SRR000649
3 SRX000122 SRP000098 SRS000290 SRR000650
  size(KB)      date
1      281 Jan 19 2012
2    130940 Jan 19 2012
3      844 Jan 19 2012
```

Next you might want to download sra data files from the ftp site. The getSRAfile function will download all available sra data files associated with "SRR000648" and "SRR000657" from the NCBI SRA ftp site to the current directory:

```
> getSRAfile( c("SRR000648","SRR000657"), sra_con, fileType = 'sra' )
```

```
      run      study      sample experiment
1 SRR000648 SRP000098 SRS000290  SRX000122
2 SRR000657 SRP000098 SRS000290  SRX000122
```

```
1 ftp://ftp-trace.ncbi.nlm.nih.gov/sra/sra-instant/reads/ByExp/sra/SRX/SRX000/SRX000122/
2 ftp://ftp-trace.ncbi.nlm.nih.gov/sra/sra-instant/reads/ByExp/sra/SRX/SRX000/SRX000122/
```

Then downloaded sra data files can be easily converted into fastq files using fastq-dump in SRA Toolkit (<http://trace.ncbi.nlm.nih.gov/Traces/sra/sra.cgi?view=software>):

```
> ## system ("fastq-dump SRR000648.lite.sra")
```

Or directly download fastq files from EBI using ftp protocol:

```
> getFASTQinfo( c("SRR000648","SRR000657"), sra_con, srcType = 'ftp' )
> getSRAfile( c("SRR000648","SRR000657"), sra_con, fileType = 'fastq' )
```

3.6 Download SRA data files using fasp protocol

Currently both NCBI and EBI supports fasp protocol for downloading SRA data files, which has several advantages over ftp protocol, including high-speed transferring large files over long distance. Please check EBI or NCBI web site or Aspera (<http://www.asperasoft.com/>) for details. SRADB has included two wrapper functions for using ascp command line program (fasp protocol) to download SRA data files from either the NCBI or EBI, which is included in Aspera Connect software. But, due to complexity of installation of the software and options within it, the functions developed here ask users to supply main ascp commands.

Download fastq files from EBI ftp site using fasp protocol:

```
> ## List fasp addresses for associated fastq files:
> listSRAfile( c("SRX000122"), sra_con, fileType = 'fastq', srcType='fasp')
> ## get fasp addresses for associated fastq files:
> getFASTQinfo( c("SRX000122"), sra_con, srcType = 'fasp' )
> ## download fastq files using fasp protocol:
> # the following ascpCMD needs to be constructed according custom
> # system configuration
> # common ascp installation in a Linux system:
> ascpCMD <- 'ascp -QT -l 300m -i
+ /usr/local/aspera/connect/etc/asperaweb_id_dsa.putty'
> ## common ascpCMD for a Mac OS X system:
> # ascpCMD <- "'/Applications/Aspera Connect.app/Contents/
> # Resources/ascp' -QT -l 300m -i '/Applications/
> # Aspera Connect.app/Contents/Resources/asperaweb_id_dsa.putty'"
>
> getSRAfile( c("SRX000122"), sra_con, fileType = 'fastq',
+           srcType = 'fasp', ascpCMD = ascpCMD )
```

Download sra files from NCBI using fasp protocol:

```
> ## List fasp addresses of sra files associated with "SRX000122"
> listSRAfile( c("SRX000122"), sra_con, fileType = 'sra', srcType='fasp')
> ## download sra files using fasp protocol
> getSRAfile( c("SRX000122"), sra_con, fileType = 'sra',
+           srcType = 'fasp', ascpCMD = ascpCMD )
```

The downloading message will show significant faster downloading speed than the ftp protocol:

```
' SRR000658.sra 100Completed: 159492K bytes transferred in 5 seconds (249247K bits/sec),
in 1 file. ... '
```

4 Interactive views of sequence data

Working with sequence data is often best done interactively in a genome browser, a task not easily done from R itself. We have found the Integrative Genomics Viewer (IGV) a high-performance visualization tool for interactive exploration of large, integrated datasets, increasing usefully for visualizing sequence alignments. In **SRADB**, functions **startIGV**, **load2IGV** and **load2newIGV** provide convenient functionality for R to interact with IGV. Note that for some OS, these functions might not work or work well.

Launch IGV with 2 GB maximum usable memory support:

```
> startIGV("mm")
```

IGV offers a remort control port that allows R to communicate with IGV. The current command set is fairly limited, but it does allow for some IGV operations to be performed in the R console. To utilize this functionality, be sure that IGV is set to allow communication via the “enable port” option in IGV preferences. To load BAM files to IGV and then manipulate the window:

```
> exampleBams = file.path(system.file('extdata',package='SRADB'),
+   dir(system.file('extdata',package='SRADB'),pattern='bam$'))
> sock <- IGVsocket()
> IGVgenome(sock, 'hg18')
> IGVload(sock, exampleBams)
> IGVgoto(sock, 'chr1:1-1000')
> IGVsnapshot(sock)
```

5 Graphic view of SRA entities

Due to the nature of SRA data and its design, sometimes it is hard to get a whole picture of the relationship between a set of SRA entities. Functions of **entityGraph** and **sraGraph** in this package generate graphNEL objects with `edgemode='directed'` from input `data.frame` or directly from search terms, and then the **plot** function can easily draw a diagram.

Create a graphNEL object directly from full text search results of terms 'primary thyroid cell line'

```
> library(SRADb)
> library(Rgraphviz)
> g <- sraGraph('primary thyroid cell line', sra_con)
> attrs <- getDefaultAttrs(list(node=list(
+   fillcolor='lightblue', shape='ellipse'))))
> plot(g, attrs=attrs)
> ## similiar search as the above, returned much larger data.frame and graph is too cl
> g <- sraGraph('Ewing Sarcoma', sra_con)
> plot(g)
>
```

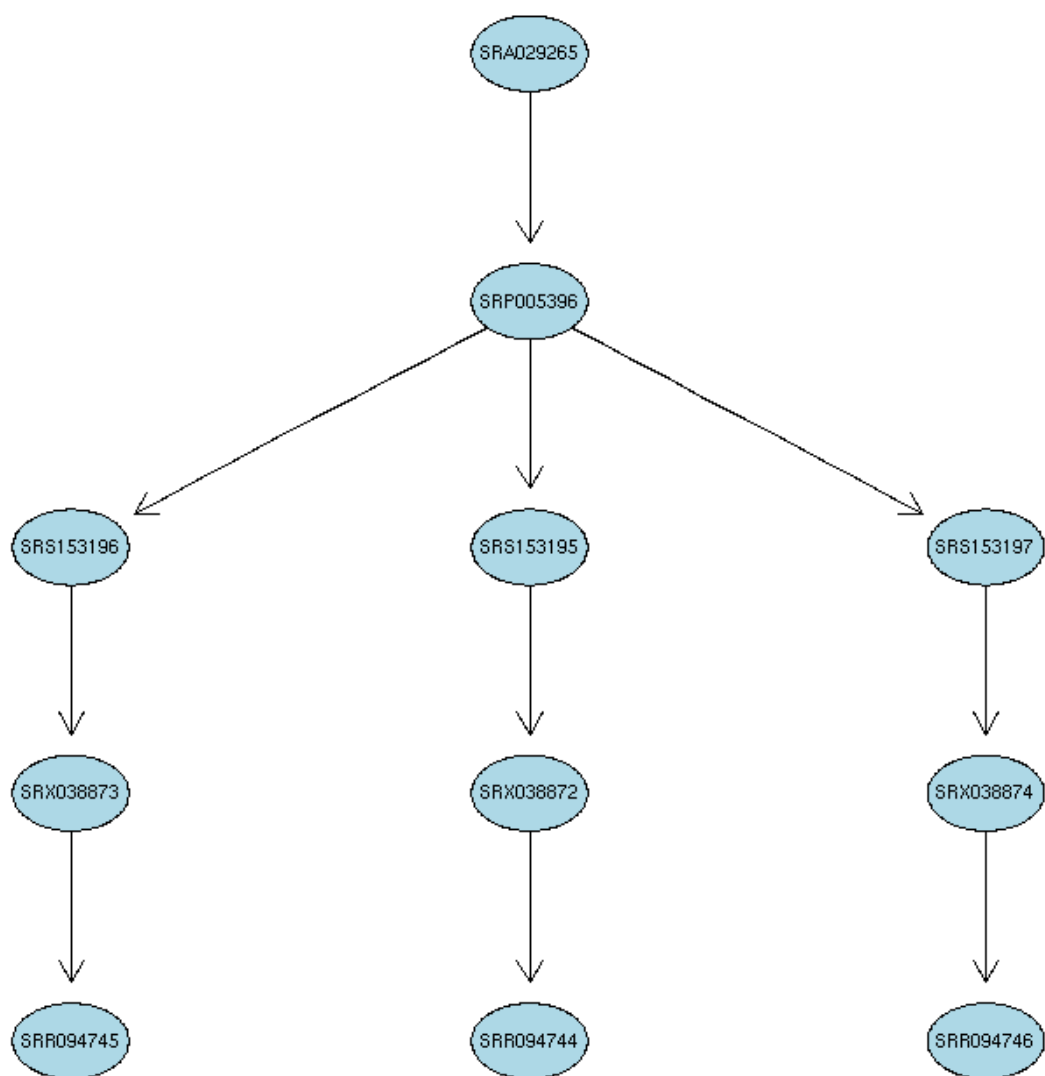



Figure 2: A graphical representation of the relationships between the SRA entities.

Please see the Figure 2 for an example diagram.

It's considered good practise to explicitly disconnect from the database once we are done with it:

```
> dbDisconnect(sra_con)
```

```
[1] TRUE
```

6 Example use case

This section will use the functionalities in the **SRADB** package to explore data from the 1000 genomes project. Mainly,

1. Get some statistics of meta data and data files from the 1000 genomes project using the **SRADB**
2. Download data files
3. Load bam files into the IGV from R
4. Create some snapshots programmatically from R

```
> library(SRADb)
> setwd('1000g')
> if( ! file.exists('SRAMetadb.sqlite') ) {
+   sqlfile <- getSRADBFile()
+ } else {
+   sqlfile <- 'SRAMetadb.sqlite'
+ }
> sra_con <- dbConnect(SQLite(),sqlfile)
> ## get all related accessions
> rs <- getSRA( search_terms = '"1000 Genomes Project"',
+   sra_con=sra_con, acc_only=TRUE)
> dim(rs)
> head(rs)
> ## get counts for each data types
> apply( rs, 2, function(x) {length(unique(x))} )
```

After you decided what data from the 1000 Genomes, you would like to download data files from the SRA. But, it might be helpful to know file size before downloading them:

```
> runs <- tail(rs$run)
> fs <- getSRAinfo( runs, sra_con, sraType = "sra" )
```

Now you can download the files through ftp protocol:

```
> getSRAfile( runs, sra_con, fileType='sra', srcType = "ftp" )
```

Or, you can download them through fasp protocol:

```
> ascpCMD <- "'/Applications/Aspera Connect.app/Contents/Resources/ascp' -QT -l 300m -"
> sra_files = getSRAfile( runs, sra_con, fileType = 'sra', srcType = "fasp", ascpCMD = .
```

Next you might want to convert the downloaded sra files into fastq files:

```
> for( fq in basename(sra_files$fasp) ) {
+     system ("fastq-dump SRR000648.lite.sra")
+ }
```

... to be completed.

7 sessionInfo

- R version 3.3.0 RC (2016-04-26 r70550), x86_64-apple-darwin13.4.0
- Locale: C/en_US.UTF-8/en_US.UTF-8/C/en_US.UTF-8/en_US.UTF-8
- Base packages: base, datasets, grDevices, graphics, methods, parallel, stats, utils
- Other packages: BiocGenerics 0.18.0, DBI 0.4, RCurl 1.95-4.8, RSQLite 1.0.0, SRADB 1.30.0, bitops 1.0-6, graph 1.50.0
- Loaded via a namespace (and not attached): Biobase 2.32.0, GEOquery 2.38.0, XML 3.98-1.4, stats4 3.3.0, tools 3.3.0