

# Package ‘mitology’

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**Type** Package

**Title** Study of mitochondrial activity from RNA-seq data

**Version** 1.3.0

**Description** mitology allows to study the mitochondrial activity through high-throughput RNA-seq data. It is based on a collection of genes whose proteins localize in to the mitochondria. From these, mitology provides a reorganization of the pathways related to mitochondria activity from Reactome and Gene Ontology. Further a ready-to-use implementation of MitoCarta3.0 pathways is included.

**License** AGPL-3

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mitology-package      *mitology: Study of mitochondrial activity from RNA-seq data*

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## Description

mitology allows to study the mitochondrial activity through high-throughput RNA-seq data. It is based on a collection of genes whose proteins localize in to the mitochondria. From these, mitology provides a reorganization of the pathways related to mitochondria activity from Reactome and Gene Ontology. Further a ready-to-use implementation of MitoCarta3.0 pathways is included.

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## See Also

Useful links:

- <https://github.com/CaluraLab/mitology>
- Report bugs at <https://github.com/CaluraLab/mitology/issues>

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**enrichMito***Mitochondrial Enrichment Analysis of a gene list.*

---

**Description**

Given a vector of genes, this function will return the enrichment analysis for the mitochondrial gene sets after FDR control. For the Reactome, GO-CC and GO-BP databases it returns also the enrichment results for the corresponding original pathways.

**Usage**

```
enrichMito(genes, database)
```

**Arguments**

genes	a vector of gene ENSEMBL id.
database	character string saying the database to use for the analysis. Either one of "MitoCarta", "Reactome", "GO-CC" and "GO-BP".

**Value**

enrichment analysis for the mitochondrial gene sets.

**Examples**

```
data(ovse)
```

---

**getGeneSets***Get the mitochondrial gene sets*

---

**Description**

It returns the mitochondrial gene sets (in form of list or data frame) of the four possible databases: "MitoCarta", "Reactome", "GO-CC" and "GO-BP".

**Usage**

```
getGeneSets(  
  database = "MitoCarta",  
  nametype = "ENSEMBL",  
  objectType = "list",  
  sections = FALSE  
)
```

**Arguments**

database	character string saying the database to use for the analysis. Either one of "Mito-Carta", "Reactome", "GO-CC" and "GO-BP".
nametype	character string saying the type of gene name ID. Either one of "SYMBOL", "ENTREZID" or "ENSEMBL".
objectType	character string saying the type of needed object. Either one of "list" or "dataframe".
sections	logical. Either to keep the aggregated gene set categories or the specific gene sets. Default is FALSE.

**Value**

the mitochondrial gene sets.

**Examples**

```
Mclist <- getGeneSets()
```

---

gseaMito

*Mitochondrial GSEA of a gene list.*

---

**Description**

Gene set enrichment analysis for the mitochondrial gene sets. For the Reactome, GO-CC and GO-BP databases it returns also the GSEA results for the corresponding original pathways.

**Usage**

```
gseaMito(genes, database)
```

**Arguments**

genes	order ranked gene vector named by ENSEMBL id.
database	character string saying the database to use for the analysis. Either one of "Mito-Carta", "Reactome", "GO-CC" and "GO-BP".

**Value**

GSEA results for the mitochondrial gene sets.

**Examples**

```
data(ovse)
```

---

MitoGenesDB

*Mitochondrial genes*

---

### Description

Here are listed all the final mitochondrial genes in ENSEMBL id, the corresponding SYMBOL id and the database from where they were collected.

### Usage

```
data(MitoGenesDB)
```

### Format

An object of class `data.frame` with 2996 rows and 3 columns.

---

mitoHeatmap

*Heatmap of mitochondrial gene sets.*

---

### Description

Given a matrix of scores, it returns a heatmap of the mitochondrial gene sets.

### Usage

```
mitoHeatmap(  
  data,  
  database = "MitoCarta",  
  sampleAnnot = NULL,  
  splitSamples = FALSE,  
  splitSections = FALSE,  
  ...  
)
```

### Arguments

<code>data</code>	matrix or <code>data.frame</code> with samples in columns and mitochondrial gene sets in rows.
<code>database</code>	character string saying the database used for the analysis. Either one of "MitoCarta", "Reactome", "GO-CC" and "GO-BP".
<code>sampleAnnot</code>	character vector with samples' annotation.
<code>splitSamples</code>	logical. If TRUE it splits samples by annotation. <code>sampleAnnot</code> must be provided.
<code>splitSections</code>	logical. If TRUE it splits gene sets by main section.
<code>...</code>	other parameters specific of the function <a href="#">Heatmap</a> .

## Value

A [Heatmap-class](#) object.

## Examples

```
MClust <- getGeneSets()
n <- length(names(MClust)) * 5
rmatrix <- matrix(rnorm(n, 0), ncol = 5)
rownames(rmatrix) <- names(MClust)
colnames(rmatrix) <- paste0("Sample_", seq_len(5))
mitoHeatmap(data = rmatrix, database = "MitoCarta")
```

---

mitoTreeHeatmap

*Circular heatmap on mitochondrial gene set tree.*

---

## Description

Given a matrix of scores, it returns a circular heatmap of the mitochondrial gene sets (leaf of the database tree) or gene set groups (section of the database tree).

## Usage

```
mitoTreeHeatmap(
  data,
  database = "MitoCarta",
  sections = FALSE,
  samples = NULL,
  labelNames = "sections",
  ...
)
```

## Arguments

<code>data</code>	matrix or data.frame with samples in columns and mitochondrial gene sets in rows.
<code>database</code>	character string saying the database used for the analysis. Either one of "MitoCarta", "Reactome", "GO-CC" and "GO-BP".
<code>sections</code>	logical. Either to keep the aggregated gene set categories or the specific gene sets. Default is FALSE.
<code>samples</code>	character vector with the names of samples to be plotted. Otherwise all samples are plotted.
<code>labelNames</code>	character string that says to plot either the names of "sections" or "leaves".
...	other arguments passed on to the <a href="#">gheatmap</a> function.

**Value**

A [ggplot](#) object.

**Examples**

```
MClust <- getGeneSets()
n <- length(names(MClust)) * 5
rmatrix <- matrix(rnorm(n, 0), ncol = 5)
rownames(rmatrix) <- names(MClust)
colnames(rmatrix) <- paste0("Sample_", seq_len(5))
mitoTreeHeatmap(data = rmatrix, database = "MitoCarta")
```

---

**mitoTreePoint***Circular dotplot on mitochondrial gene set tree.*

---

**Description**

A circular dotplot of the mitochondrial enrichment results.

**Usage**

```
mitoTreePoint(
  data,
  database = "MitoCarta",
  pvalCutoff = 0.05,
  labsizes = 3,
  max_point_size = 4,
  color = "p.adjust"
)
```

**Arguments**

<code>data</code>	named list of the result from <code>enrichMito</code> or <code>gseaMito</code> .
<code>database</code>	character string saying the database to use for the analysis. Either one of "MitoCarta", "Reactome", "GO-CC" and "GO-BP".
<code>pvalCutoff</code>	pvalue cutoff to select enriched gene sets
<code>labsizes</code>	label size
<code>max_point_size</code>	max point size
<code>color</code>	variable used to color enriched terms, e.g. 'pvalue', 'p.adjust' or 'NES'.

**Value**

A [ggplot](#) object.

## Examples

```
data(ovse)
```

---

ovse

*Example expression data.*

---

## Description

This is an example dataset containing gene expression values (in normalized counts) of 40 ovarian cancer (OVC) patients extracted from the Cancer Genome Atlas (TCGA) database. This dataset should be used only with example purpose. RNA sequencing OVC data were retrieved using [curatedTCGAData](#) package. Data were then normalized with the [betweenLaneNormalization](#) function. To lighten the dataset, the [consensusOVSign](#) function was computed, which return 4 different scores, one for each OVC subtype (Chen et al, 2018, Clinical Cancer Research) and the 10 samples with the highest scores were selected for each subgroup. Further, only the mitochondrial genes included in mitology were kept. Finally, the log fold change of the IMR versus the PRO samples were computed. Further details in mitology/inst/scripts/howToGenerateOvse.Rmd.

## Usage

```
data(ovse)
```

## Format

An object of class `SummarizedExperiment` with 2388 rows and 40 columns.

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