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MailingList microbiome <microbiome-devel@googlegroups.com>

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microbiome-package *R package for microbiome studies*

Description

Brief summary of the microbiome package

Details

Package:	microbiome
Type:	Package
Version:	See sessionInfo() or DESCRIPTION file
Date:	2014-2017
License:	FreeBSD
LazyLoad:	yes

R package for microbiome studies

Author(s)

Leo Lahti et al. <microbiome-admin@googlegroups.com>

References

See citation('microbiome') http://microbiome.github.io

Examples

```
citation('microbiome')
```

abundances *Abundance Matrix from Phyloseq*

Description

Retrieves the taxon abundance table from phyloseq-class object and ensures it is systematically returned as taxa x samples matrix.

Usage

```
abundances(x, transform = "identity")
```

Arguments

x	phyloseq-class object
transform	Transformation to apply. The options include: 'compositional' (ie relative abundance), 'Z', 'log10', 'log10p', 'hellinger', 'identity', 'clr', 'alr', or any method from the vegan::decostand function.

Value

Abundance matrix (OTU x samples).

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

References

See citation('microbiome')

Examples

```
data(dietswap)
a <- abundances(dietswap)
# b <- abundances(dietswap, transform='compositional')
```

---

add_besthit *Adds* best_hist *to a* phyloseq-class *Object*

Description

Add the lowest classification for an OTU or ASV.

Usage

```
add_besthit(x, sep = ":")
```

Arguments

x	phyloseq-class object
sep	separator e.g. ASV161:Roseburia

Details

Most commonly it is observed that taxa names are either OTU ids or ASV ids. In such cases it is useful to know the taxonomic identity. For this purpose, best_hist identifies the best available taxonomic identity and adds it to the OTU ids or ASV ids. If genus and species columns are present in input the function internally combines the names.

Value

phyloseq-class object phyloseq-class

Author(s)

Contact: Sudarshan A. Shetty <sudarshanshetty9@gmail.com>

Examples

```
## Not run:
# Example data
library(microbiome)
data(dietswap)
p0.f <- add_besthit(atlas1006, sep=":")

## End(Not run)
```

---

add_refseq *Add* refseq *Slot for* dada2 *based* phyloseq *Object*

Description

Utility to add refseq slot for dada2 based phyloseq Object. Here, the taxa_names which are unique sequences, are stored in refseq slot of phyloseq. Sequence ids are converted to ids using tag option.

Usage

```
add_refseq(x, tag = "ASV")
```

Arguments

x	phyloseq-class object with sequences as rownames.
tag	Provide name for Ids, Default="ASV".

Value

phyloseq-class object

Author(s)

Contact: Sudarshan A. Shetty <sudarshanshetty9@gmail.com>

Examples

```
# ps <- add_refseq(p0,tag="ASV")
# ps
```

aggregate_rare *Aggregate Rare Groups*

Description

Combining rare taxa.

Usage

```
aggregate_rare(x, level, detection, prevalence, include.lowest = FALSE, ...)
```

Arguments

x	phyloseq-class object
level	Summarization level (from rank_names(pseq))
detection	Detection threshold for absence/presence (strictly greater by default).
prevalence	Prevalence threshold (in [0, 1]). The required prevalence is strictly greater by default. To include the limit, set include.lowest to TRUE.
include.lowest	Include the lower boundary of the detection and prevalence cutoffs. FALSE by default.
...	Arguments to pass.

Value

phyloseq-class object

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

References

See citation('microbiome')

Examples

```
data(dietswap)
s <- aggregate_rare(dietswap, level = 'Phylum',
    detection = 0.1/100, prevalence = 5/100)
```

aggregate_taxa *Aggregate Taxa*

Description

Summarize phyloseq data into a higher phylogenetic level.

Usage

```
aggregate_taxa(x, level, verbose = FALSE)
```

Arguments

x	phyloseq-class object
level	Summarization level (from rank_names(pseq))
verbose	verbose

Details

This provides a convenient way to aggregate phyloseq OTUs (or other taxa) when the phylogenetic tree is missing. Calculates the sum of OTU abundances over all OTUs that map to the same higher-level group. Removes ambiguous levels from the taxonomy table. Returns a phyloseq object with the summarized abundances.

Value

Summarized phyloseq object

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

References

See citation('microbiome')

Examples

```
data(dietswap)
s <- aggregate_taxa(dietswap, 'Phylum')
```

alpha *Global Ecosystem State Variables*

Description

Global indicators of the ecosystem state, including richness, evenness, diversity, and other indicators

Usage

```
alpha(x, index = "all", zeroes = TRUE)
```

Arguments

x	A species abundance vector, or matrix (taxa/features x samples) with the absolute count data (no relative abundances), or phyloseq-class object
index	Default is 'NULL', meaning that all available indices will be included. For specific options, see details.
zeroes	Include zero counts in the diversity estimation.

Details

This function returns various indices of the ecosystem state. The function is named alpha (global in some previous versions of this package) as these indices can be viewed as measures of alpha diversity. The function uses default choices for detection, prevalence and other parameters for simplicity and standardization. See the individual functions for more options. All indicators from the richness, diversity, evenness, dominance, and rarity functions are available. Some additional measures, such as Chao1 and ACE are available via estimate_richness function in the **phyloseq** package but not included here. The index names are given the prefix richness_, evenness_, diversity_, dominance_, or rarity_ in the output table to avoid confusion between similarly named but different indices (e.g. Simpson diversity and Simpson dominance). All parameters are set to their default. To experiment with different parameterizations, see the more specific index functions (richness, diversity, evenness, dominance, rarity).

Value

A data.frame of samples x alpha diversity indicators

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

References

See citation('microbiome')

See Also

dominance, rarity, phyloseq::estimate_richness

Examples

```
data(dietswap)
d <- alpha(dietswap, index='shannon')
# d <- alpha(dietswap, index='all')
```

`associate` *Cross Correlation Wrapper*

Description

Cross-correlate columns of the input matrices.

Usage

```
associate(
  x,
  y = NULL,
  method = "spearman",
  p.adj.threshold = Inf,
  cth = NULL,
  order = FALSE,
  n.signif = 0,
  mode = "table",
  p.adj.method = "fdr",
  verbose = FALSE,
  filter.self.correlations = FALSE
)
```

Arguments

`x`	matrix (samples x features if annotation matrix)
`y`	matrix (samples x features if cross-correlated with annotations)
`method`	association method ('pearson', or 'spearman' for continuous)
`p.adj.threshold`	q-value threshold to include features
`cth`	correlation threshold to include features
`order`	order the results
`n.signif`	mininum number of significant correlations for each element
`mode`	Specify output format ('table' or 'matrix')
`p.adj.method`	p-value multiple testing correction method. One of the methods in p.adjust function ('BH' and others; see help(p.adjust)). Default: 'fdr'
`verbose`	verbose
`filter.self.correlations`	Filter out correlations between identical items.

Details

The p-values in the output table depend on the method. For the spearman and pearson correlation values, the p-values are provided by the default method in the cor.test function.

Value

List with cor, pval, pval.adjusted

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

References

See citation('microbiome')

Examples

```
data(peerj32)
d1 <- peerj32$microbes[1:20, 1:10]
d2 <- peerj32$lipids[1:20,1:10]
cc <- associate(d1, d2, method='pearson')
```

---

atlas1006 *HITChip Atlas with 1006 Western Adults*

Description

This data set contains genus-level microbiota profiling with HITChip for 1006 western adults with no reported health complications, reported in Lahti et al. (2014) https://doi.org/10.1038/ncomms5344.

Usage

```
data(atlas1006)
```

Format

The data set in phyloseq-class format.

Details

The data is also available for download from the Data Dryad http://doi.org/10.5061/dryad.pk75d.

Value

Loads the data set in R.

Author(s)

Leo Lahti <microbiome-admin@googlegroups.com>

References

Lahti et al. Tipping elements of the human intestinal ecosystem. Nature Communications 5:4344, 2014. To cite the microbiome R package, see citation('microbiome')

---

baseline *Pick Baseline Timepoint Samples*

Description

Identify and select the baseline timepoint samples in a phyloseq object.

Usage

```
baseline(x, na.omit = TRUE)
```

Arguments

x	phyloseq object. Assuming that the sample_data(x) has the fields 'time', 'sample' and 'subject'
na.omit	Logical. Ignore samples with no time point information. If this is FALSE, the first sample for each subject is selected even when there is no time information.

Details

Arranges the samples by time and picks the first sample for each subject. Compared to simple subsetting at time point zero, this checks NAs and possibility for multiple samples at the baseline, and guarantees that a single sample per subject is selected.

Value

Phyloseq object with only baseline time point samples selected.

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

References

See citation('microbiome')

Examples

```
data(peerj32)
a <- baseline(peerj32$phyloseq)
```

`bimodality` *Bimodality Analysis*

Description

Estimate bimodality scores.

Usage

```
bimodality(
  x,
  method = "potential_analysis",
  peak.threshold = 1,
  bw.adjust = 1,
  bs.iter = 100,
  min.density = 1,
  verbose = TRUE
)
```

Arguments

`x`	A vector, matrix, or a phyloseq object
`method`	bimodality quantification method ('potential_analysis', 'Sarle.finite.sample', or 'Sarle.asymptotic'). If method='all', then a data.frame with all scores is returned.
`peak.threshold`	Mode detection threshold
`bw.adjust`	Bandwidth adjustment
`bs.iter`	Bootstrap iterations
`min.density`	minimum accepted density for a maximum; as a multiple of kernel height
`verbose`	Verbose

Details

- Sarle.finite.sample Coefficient of bimodality for finite sample. See SAS 2012.
- Sarle.asymptotic Coefficient of bimodality, used and described in Shade et al. (2014) and Ellison AM (1987).
- potential_analysis Repeats potential analysis (Livina et al. 2010) multiple times with bootstrap sampling for each row of the input data (as in Lahti et al. 2014) and returns the bootstrap score.

The coefficient lies in (0, 1).

The 'Sarle.asymptotic' version is defined as

$$b = (g^2 + 1)/k$$

. This is coefficient of bimodality from Ellison AM Am. J. Bot. 1987, for microbiome analysis it has been used for instance in Shade et al. 2014. The formula for 'Sarle.finite.sample' (SAS 2012):

$$b = \frac{g^2 + 1}{k + (3(n-1)^2)/((n-2)(n-3))}$$

where n is sample size and In both formulas, g is sample skewness and k is the kth standardized moment (also called the sample kurtosis, or excess kurtosis).

Value

A list with following elements:

- scoreFraction of bootstrap samples where multiple modes are observed
- nmodesThe most frequently observed number of modes in bootstrap sampling results.
- resultsFull results of potential_analysis for each row of the input matrix.

Author(s)

Leo Lahti <leo.lahti@iki.fi>

References

- Livina et al. (2010). Potential analysis reveals changing number of climate states during the last 60 kyr. *Climate of the Past*, 6, 77-82.
- Lahti et al. (2014). Tipping elements of the human intestinal ecosystem. *Nature Communications* 5:4344.
- Shade et al. mBio 5(4):e01371-14, 2014.
- AM Ellison, Am. J. Bot 74:1280-8, 1987.
- SAS Institute Inc. (2012). SAS/STAT 12.1 user's guide. Cary, NC.
- To cite the microbiome R package, see citation('microbiome')

See Also

A classical test of multimodality is provided by `dip.test` in the **DIP** package.

Examples

```
# In practice, use more bootstrap iterations
b <- bimodality(c(rnorm(100, mean=0), rnorm(100, mean=5)),
    method = "Sarle.finite.sample", bs.iter=5)
# The classical DIP test:
# quantifies unimodality. Values range between 0 to 1.
# dip.test(x, simulate.p.value=TRUE, B=200)$statistic
# Values less than 0.05 indicate significant deviation from unimodality.
# Therefore, to obtain an increasing multimodality score, use
# library(diptest)
# multimodality.dip <- apply(abundances(pseq), 1,
# function (x) {1 - unname(dip.test(x)$p.value)})
```

bimodality_sarle *Sarle's Bimodality Coefficient*

Description

Sarle's bimodality coefficient.

Usage

```
bimodality_sarle(x, bs.iter = 1, type = "Sarle.finite.sample")
```

Arguments

x	Data vector for which bimodality will be quantified
bs.iter	Bootstrap iterations
type	Score type ('Sarle.finite.sample' or 'Sarle.asymptotic')

Details

The coefficient lies in (0, 1).

The 'Sarle.asymptotic' version is defined as

$$b = (g^2 + 1)/k$$

. This is coefficient of bimodality from Ellison AM Am. J. Bot. 1987, for microbiome analysis it has been used for instance in Shade et al. 2014.

The formula for 'Sarle.finite.sample' (SAS 2012):

$$b = \frac{g^2 + 1}{k + (3(n-1)^2)/((n-2)(n-3))}$$

where n is sample size and

In both formulas, g is sample skewness and k is the kth standardized moment (also called the sample kurtosis, or excess kurtosis).

Value

Bimodality score

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

References

- Shade et al. mBio 5(4):e01371-14, 2014.
- Ellison AM (1987) Am J Botany 74(8):1280-1288.
- SAS Institute Inc. (2012). SAS/STAT 12.1 user's guide. Cary, NC.
- To cite the microbiome R package, see citation('microbiome')

See Also

Check the dip.test from the **DIP** package for a classical test of multimodality.

Examples

```
# b <- bimodality_sarle(rnorm(50), type='Sarle.finite.sample')
```

`boxplot_abundance` *Abundance Boxplot*

Description

Plot phyloseq abundances.

Usage

```
boxplot_abundance(
  d,
  x,
  y,
  line = NULL,
  violin = FALSE,
  na.rm = FALSE,
  show.points = TRUE
)
```

Arguments

`d`	`phyloseq-class` object
`x`	Metadata variable to map to the horizontal axis.
`y`	OTU to map on the vertical axis
`line`	The variable to map on lines
`violin`	Use violin version of the boxplot
`na.rm`	Remove NAs
`show.points`	Include data points in the figure

Details

The directionality of change in paired boxplot is indicated by the colors of the connecting lines.

Value

A ggplot plot object

Examples

```
data(peerj32)
p <- boxplot_abundance(peerj32$phyloseq, x='time', y='Akkermansia',
   line='subject')
```

---

boxplot_alpha *Alpha Boxplot*

Description

Plot alpha index.

Usage

```
boxplot_alpha(
  x,
  x_var = NULL,
  index = NULL,
  violin = FALSE,
  na.rm = FALSE,
  show.points = TRUE,
  zeroes = TRUE,
  element.alpha = 0.5,
  element.width = 0.2,
  fill.colors = NA,
  outlier.fill = "grey50"
)
```

Arguments

x	phyloseq-class object
x_var	Metadata variable to map to the horizontal axis.
index	Alpha index to plot. See function alpha.
violin	Use violin version of the boxplot
na.rm	Remove NAs
show.points	Include data points in the figure
zeroes	Include zero counts in diversity estimation. Default is TRUE

element.alpha	Alpha value for plot elements. Controls the transparency of plots elements.
element.width	Width value for plot elements. Controls the transparency of plots elements.
fill.colors	Specify a list of colors passed on to ggplot2 scale_fill_manual
outlier.fill	If using boxplot and and points together how to deal with outliers. See ggplot2 outlier.fill argument in geom_ elements.

Details

A simple wrapper to visualize alpha diversity index.

Value

A ggplot plot object

Examples

```
data("dietswap")
p <- boxplot_alpha(dietswap, x_var = "sex", index="observed", violin=FALSE,
                   na.rm=FALSE, show.points=TRUE, zeroes=TRUE,
                   element.alpha=0.5, element.width=0.2,
                   fill.colors= c("steelblue", "firebrick"),
                   outlier.fill="white")
p
```

---

chunk_reorder *Chunk Reorder*

Description

Chunk re-order a vector so that specified newstart is first. Different than relevel.

Usage

```
chunk_reorder(x, newstart = x[[1]])
```

Details

Borrowed from **phyloseq** package as needed here and not exported there. Rewritten.

Value

Reordered x

Examples

```
# Typical use-case
# chunk_reorder(1:10, 5)
# # Default is to not modify the vector
# chunk_reorder(1:10)
# # Another example not starting at 1
# chunk_reorder(10:25, 22)
# # Should silently ignore the second element of `newstart`
# chunk_reorder(10:25, c(22, 11))
# # Should be able to handle `newstart` being the first argument already
# # without duplicating the first element at the end of `x`
# chunk_reorder(10:25, 10)
# all(chunk_reorder(10:25, 10) == 10:25)
# # This is also the default
# all(chunk_reorder(10:25) == 10:25)
# # An example with characters
# chunk_reorder(LETTERS, 'G')
# chunk_reorder(LETTERS, 'B')
# chunk_reorder(LETTERS, 'Z')
# # What about when `newstart` is not in `x`? Return x as-is, throw warning.
# chunk_reorder(LETTERS, 'g')
```

`cmat2table` *Convert Correlation Matrix into a Table*

Description

Arrange correlation matrices from associate into a table format.

Usage

```
cmat2table(res, verbose = FALSE)
```

Arguments

`res`	Output from associate
`verbose`	verbose

Value

Correlation table

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

References

See citation('microbiome')

Examples

```
data(peerj32)
d1 <- peerj32$microbes[1:20, 1:10]
d2 <- peerj32$lipids[1:20,1:10]
cc <- associate(d1, d2, mode='matrix', method='pearson')
cmat <- associate(d1, d2, mode='table', method='spearman')
```

---

collapse_replicates *Collapse Replicate Samples*

Description

Collapse samples, mostly meant for technical replicates.

Usage

```
collapse_replicates(
  x,
  method = "sample",
  replicate_id = NULL,
  replicate_fields = NULL
)
```

Arguments

x	phyloseq-class object
method	Collapsing method. Only random sampling ("sample") implemented.
replicate_id	Replicate identifier. A character vector.
replicate_fields	Metadata fields used to determine replicates.

Value

Collapsed phyloseq object.

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

References

To cite the microbiome R package, see citation('microbiome')

Examples

```
data(atlas1006)
pseq <- collapse_replicates(atlas1006,
        method = "sample",
        replicate_fields = c("subject", "time"))
```

core *Core Microbiota*

Description

Filter the phyloseq object to include only prevalent taxa.

Usage

```
core(x, detection, prevalence, include.lowest = FALSE, ...)
```

Arguments

x	phyloseq-class object
detection	Detection threshold for absence/presence (strictly greater by default).
prevalence	Prevalence threshold (in [0, 1]). The required prevalence is strictly greater by default. To include the limit, set include.lowest to TRUE.
include.lowest	Include the lower boundary of the detection and prevalence cutoffs. FALSE by default.
...	Arguments to pass.

Value

Filtered phyloseq object including only prevalent taxa

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

References

Salonen A, Salojarvi J, Lahti L, de Vos WM. The adult intestinal core microbiota is determined by analysis depth and health status. Clinical Microbiology and Infection 18(S4):16-20, 2012 To cite the microbiome R package, see citation('microbiome')

See Also

core_members, rare_members

Examples

```
data(dietswap)
# Detection threshold 0 (strictly greater by default);
# Prevalence threshold 50 percent (strictly greater by default)
pseq <- core(dietswap, 0, 50/100)
# Detection threshold 0 (strictly greater by default);
# Prevalence threshold exactly 100 percent; for this set
```

```
# include.lowest=TRUE, otherwise the required prevalence is
# strictly greater than 100
pseq <- core(dietswap, 0, 100/100, include.lowest = TRUE)
```

---

core_abundance *Core Abundance*

Description

Calculates the community core abundance index.

Usage

```
core_abundance(
  x,
  detection = 0.1/100,
  prevalence = 50/100,
  include.lowest = FALSE
)
```

Arguments

x	phyloseq-class object
detection	Detection threshold for absence/presence (strictly greater by default).
prevalence	Prevalence threshold (in [0, 1]). The required prevalence is strictly greater by default. To include the limit, set include.lowest to TRUE.
include.lowest	Include the lower boundary of the detection and prevalence cutoffs. FALSE by default.

Details

The core abundance index gives the relative proportion of the core species (in [0,1]). The core taxa are defined as those that exceed the given population prevalence threshold at the given detection level.

Value

A vector of core abundance indices

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

See Also

rarity

Examples

```
data(dietswap)
d <- core_abundance(dietswap, detection=0.1/100, prevalence=50/100)
```

`core_heatmap` *Core Heatmap*

Description

Core heatmap.

Usage

```
core_heatmap(x, dets, cols, min.prev, taxa.order)
```

Arguments

`x`	OTU matrix
`dets`	A vector or a scalar indicating the number of intervals in (0, log10(max(data))). The dets are calculated for relative abundancies.
`cols`	colours for the heatmap
`min.prev`	If minimum prevalence is set, then filter out those rows (taxa) and columns (dets) that never exceed this prevalence. This helps to zoom in on the actual core region of the heatmap.
`taxa.order`	Ordering of the taxa.

Value

Used for its side effects

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

References

A Salonen et al. The adult intestinal core microbiota is determined by analysis depth and health status. Clinical Microbiology and Infection 18(S4):16 20, 2012. To cite the microbiome R package, see citation('microbiome')

core_matrix *Core Matrix*

Description

Creates the core matrix.

Usage

```
core_matrix(x, prevalences = seq(0.1, 1, , 1), detections = NULL)
```

Arguments

x	phyloseq object or a taxa x samples abundance matrix
prevalences	a vector of prevalence percentages in [0,1]
detections	a vector of intensities around the data range

Value

Estimated core microbiota

Author(s)

Contact: Jarkko Salojarvi <microbiome-admin@googlegroups.com>

References

A Salonen et al. The adult intestinal core microbiota is determined by analysis depth and health status. Clinical Microbiology and Infection 18(S4):16 20, 2012. To cite the microbiome R package, see citation('microbiome')

Examples

```
# Not exported
#data(peerj32)
#core <- core_matrix(peerj32$phyloseq)
```

core_members *Core Taxa*

Description

Determine members of the core microbiota with given abundance and prevalences

Usage

```
core_members(x, detection = 1/100, prevalence = 50/100, include.lowest = FALSE)
```

Arguments

x	phyloseq-class object
detection	Detection threshold for absence/presence (strictly greater by default).
prevalence	Prevalence threshold (in [0, 1]). The required prevalence is strictly greater by default. To include the limit, set include.lowest to TRUE.
include.lowest	Include the lower boundary of the detection and prevalence cutoffs. FALSE by default.

Details

For phyloseq object, lists taxa that are more prevalent with the given detection threshold. For matrix, lists columns that satisfy these criteria.

Value

Vector of core members

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

References

A Salonen et al. The adult intestinal core microbiota is determined by analysis depth and health status. Clinical Microbiology and Infection 18(S4):16 20, 2012. To cite the microbiome R package, see citation('microbiome')

Examples

```
data(dietswap)
# Detection threshold 1 (strictly greater by default);
# Note that the data (dietswap) is here in absolute counts
# (and not compositional, relative abundances)
# Prevalence threshold 50 percent (strictly greater by default)
a <- core_members(dietswap, 1, 50/100)
```

coverage *Coverage Index*

Description

Community coverage index.

Usage

```
coverage(x, threshold = 0.5)
```

Arguments

x	A species abundance vector, or matrix (taxa/features x samples) with the absolute count data (no relative abundances), or phyloseq-class object
threshold	Indicates the fraction of the ecosystem to be occupied by the N most abundant species (N is returned by this function). If the detection argument is a vector, then a data.frame is returned, one column for each detection threshold.

Details

The coverage index gives the number of groups needed to have a given proportion of the ecosystem occupied (by default 0.5 ie 50

Value

A vector of coverage indices

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

See Also

dominance, alpha

Examples

```
data(dietswap)
d <- coverage(dietswap, threshold=0.5)
```

default_colors *Default Colors*

Description

Default colors for different variables.

Usage

```
default_colors(x, v = NULL)
```

Arguments

x	Name of the variable type ("Phylum")
v	Optional. Vector of elements to color.

Value

Named character vector of default colors

Author(s)

Leo Lahti <leo.lahti@iki.fi>

References

See citation("microbiome")

Examples

```
col <- default_colors("Phylum")
```

densityplot *Density Plot*

Description

Density visualization for data points overlaid on cross-plot.

Usage

```
densityplot(
  x,
  main = NULL,
  x.ticks = 10,
  rounding = 0,
  add.points = TRUE,
  col = "black",
  adjust = 1,
  size = 1,
  legend = FALSE,
  shading = TRUE,
  shading.low = "white",
  shading.high = "black",
  point.opacity = 0.75
)
```

Arguments

x	Data matrix to plot. The first two columns will be visualized as a cross-plot.
main	title text
x.ticks	Number of ticks on the X axis
rounding	Rounding for X axis tick values
add.points	Plot the data points as well
col	Color of the data points. NAs are marked with darkgray.
adjust	Kernel width adjustment
size	point size
legend	plot legend TRUE/FALSE
shading	Shading
shading.low	Color for shading low density regions
shading.high	Color for shading high density regions
point.opacity	Transparency-level for points

Value

ggplot2 object

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

References

See citation('microbiome')

Examples

```
# p <- densityplot(cbind(rnorm(100), rnorm(100)))
```

---

dietswap *Diet Swap Data*

Description

The diet swap data set represents a study with African and African American groups undergoing a two-week diet swap. For details, see dx.doi.org/10.1038/ncomms7342.

Usage

```
data(dietswap)
```

Format

The data set in phyloseq-class format.

Details

The data is also available for download from the Data Dryad repository http://datadryad.org/resource/doi:10.5061/dryad.1mn1n.

Value

Loads the data set in R.

Author(s)

Leo Lahti <microbiome-admin@googlegroups.com>

References

O'Keefe et al. Nature Communications 6:6342, 2015. dx.doi.org/10.1038/ncomms7342 To cite the microbiome R package, see citation('microbiome')

`divergence` *Divergence within a Sample Group*

Description

Quantify microbiota divergence (heterogeneity) within a given sample set with respect to a reference.

Usage

```
divergence(x, y, method = "bray")
```

Arguments

`x`	phyloseq object or a vector
`y`	Reference sample. A vector.
`method`	dissimilarity method: any method available via phyloseq::distance function. Note that some methods ("jsd" and 'unifrac' for instance) do not work with the group divergence.

Details

Microbiota divergence (heterogeneity / spread) within a given sample set can be quantified by the average sample dissimilarity or beta diversity with respect to a given reference sample.

This measure is sensitive to sample size. Subsampling or bootstrapping can be applied to equalize sample sizes between comparisons.

Value

Vector with dissimilarities; one for each sample, quantifying the dissimilarity of the sample from the reference sample.

Author(s)

Leo Lahti <microbiome-admin@googlegroups.com>

References

To cite this R package, see citation('microbiome')

See Also

the vegdist function from the **vegan** package provides many standard beta diversity measures

Examples

```
# Assess beta diversity among the African samples
# in a diet swap study (see \code{help(dietswap)} for references)
data(dietswap)
pseq <- subset_samples(dietswap, nationality == 'AFR')
reference <- apply(abundances(pseq), 1, median)
b <- divergence(pseq, reference, method = "bray")
```

`diversity` *Diversity Index*

Description

Various community diversity indices.

Usage

```
diversity(x, index = "all", zeroes = TRUE)
```

Arguments

`x`	A species abundance vector, or matrix (taxa/features x samples) with the absolute count data (no relative abundances), or `phyloseq-class` object
`index`	Diversity index. See details for options.
`zeroes`	Include zero counts in the diversity estimation.

Details

By default, returns all diversity indices. The available diversity indices include the following:

- inverse_simpson Inverse Simpson diversity: $1/lambda$ where $lambda=sum(p^2)$ and p are relative abundances.
- gini_simpson Gini-Simpson diversity $1 - lambda$. This is also called Gibbs–Martin, or Blau index in sociology, psychology and management studies.
- shannon Shannon diversity ie entropy
- fisher Fisher alpha; as implemented in the **vegan** package
- coverage Number of species needed to cover 50% of the ecosystem. For other quantiles, apply the function coverage directly.

Value

A vector of diversity indices

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

References

Beisel J-N. et al. A Comparative Analysis of Diversity Index Sensitivity. Internal Rev. Hydrobiol. 88(1):3-15, 2003. URL: https://portais.ufg.br/up/202/o/2003-comparative_evennes_index.pdf

Bulla L. An index of diversity and its associated diversity measure. Oikos 70:167–171, 1994

Magurran AE, McGill BJ, eds (2011) Biological Diversity: Frontiers in Measurement and Assessment (Oxford Univ Press, Oxford), Vol 12.

Smith B and Wilson JB. A Consumer's Guide to Diversity Indices. Oikos 76(1):70-82, 1996.

See Also

dominance, richness, evenness, rarity, alpha

Examples

```
data(dietswap)
d <- alpha(dietswap, 'shannon')
```

`dominance` *Dominance Index*

Description

Calculates the community dominance index.

Usage

```
dominance(x, index = "all", rank = 1, relative = TRUE, aggregate = TRUE)
```

Arguments

`x`	A species abundance vector, or matrix (taxa/features x samples) with the absolute count data (no relative abundances), or `phyloseq-class` object
`index`	If the index is given, it will override the other parameters. See the details below for description and references of the standard dominance indices. By default, this function returns the Berger-Parker index, ie relative dominance at rank 1.
`rank`	Optional. The rank of the dominant taxa to consider.
`relative`	Use relative abundances (default: TRUE)
`aggregate`	Aggregate (TRUE; default) the top members or not. If aggregate=TRUE, then the sum of relative abundances is returned. Otherwise the relative abundance is returned for the single taxa with the indicated rank.

Details

The dominance index gives the abundance of the most abundant species. This has been used also in microbiomics context (Locey & Lennon (2016)). The following indices are provided:

- 'absolute' This is the most simple variant, giving the absolute abundance of the most abundant species (Magurran & McGill 2011). By default, this refers to the single most dominant species (rank=1) but it is possible to calculate the absolute dominance with rank n based on the abundances of top-n species by tuning the rank argument.
- 'relative' Relative abundance of the most abundant species. This is with rank=1 by default but can be calculated for other ranks.
- 'DBP' Berger–Parker index, a special case of relative dominance with rank 1; This also equals the inverse of true diversity of the infinite order.
- 'DMN' McNaughton's dominance. This is the sum of the relative abundance of the two most abundant taxa, or a special case of relative dominance with rank 2
- 'simpson' Simpson's index ($sum(p^2)$) where p are relative abundances has an interpretation as a dominance measure. Also the version ($sum(q * (q-1)) / S(S-1)$) based on absolute abundances q has been proposed by Simpson (1949) but not included here as it is not within [0,1] range, and it is highly correlated with the simpler Simpson dominance. Finally, it is also possible to calculated dominances up to an arbitrary rank by setting the rank argument
- 'core_abundance' Relative proportion of the core species that exceed detection level 0.2% in over 50% of the samples
- 'gini' Gini index is calculated with the function `inequality`.

By setting aggregate=FALSE, the abundance for the single n'th most dominant taxa (n=rank) is returned instead the sum of abundances up to that rank (the default).

Value

A vector of dominance indices

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

References

Kenneth J. Locey and Jay T. Lennon. Scaling laws predict global microbial diversity. PNAS 2016 113 (21) 5970-5975; doi:10.1073/pnas.1521291113.

Magurran AE, McGill BJ, eds (2011) Biological Diversity: Frontiers in Measurement and Assessment (Oxford Univ Press, Oxford), Vol 12

See Also

coverage, core_abundance, rarity, alpha

Examples

```
data(dietswap)
# vector
d <- dominance(abundances(dietswap)[,1], rank=1, relative=TRUE)
# matrix
# d <- dominance(abundances(dietswap), rank=1, relative=TRUE)
# Phyloseq object
# d <- dominance(dietswap, rank=1, relative=TRUE)
```

`dominant` *Dominant taxa*

Description

Returns the dominant taxonomic group for each sample.

Usage

```
dominant(x, level = NULL)
```

Arguments

`x`	A `phyloseq-class` object
`level`	Optional. Taxonomic level.

Value

A vector of dominance indices

Author(s)

Leo Lahti <microbiome-admin@googlegroups.com>

Examples

```
data(dietswap)
# vector
d <- dominant(dietswap)
```

`estimate_stability` *Estimate Stability*

Description

Quantify intermediate stability with respect to a given reference point.

Usage

```
estimate_stability(df, reference.point = NULL, method = "lm", spl.list)
```

Arguments

`df`	Combined input data vector (samples x variables) and metadata data.frame (samples x features) with the 'data', 'subject' and 'time' field for each sample
`reference.point`	Optional. Calculate stability of the data w.r.t. this point. By default the intermediate range is used (min + (max - min)/2)
`method`	'lm' (linear model) or 'correlation'; the linear model takes time into account as a covariate
`spl`	split object to speed up

Details

Decomposes each column in x into differences between consecutive time points. For each variable and time point we calculate for the data values: (i) the distance from reference point; (ii) distance from the data value at the consecutive time point. The 'correlation' method calculates correlation between these two variables. Negative correlations indicate that values closer to reference point tend to have larger shifts in the consecutive time point. The 'lm' method takes the time lag between the consecutive time points into account as this may affect the comparison and is not taken into account by the straightforward correlation. Here the coefficients of the following linear model are used to assess stability: abs(change) ~ time + abs(start.reference.distance). Samples with missing data, and subjects with less than two time point are excluded.

Value

A list with following elements: stability: estimated stability data: processed data set used in calculations

Author(s)

Leo Lahti <leo.lahti@iki.fi>

Examples

```
# df <- data.frame(list(
#           subject=rep(paste('subject', 1:50, sep='-'), each=2),
#           time=rep(1:2, 50),
#           data=rnorm(100)))
#s <- estimate_stability_single(df, reference.point=NULL, method='lm')
```

`evenness` *Evenness Index*

Description

Various community evenness indices.

Usage

```
evenness(x, index = "all", zeroes = TRUE, detection = 0)
```

Arguments

`x`	A species abundance vector, or matrix (taxa/features x samples) with the absolute count data (no relative abundances), or `phyloseq-class` object
`index`	Evenness index. See details for options.
`zeroes`	Include zero counts in the evenness estimation.
`detection`	Detection threshold

Details

By default, Pielou's evenness is returned.

The available evenness indices include the following: 1) 'camargo': Camargo's evenness (Camargo 1992) 2) 'simpson': Simpson's evenness (inverse Simpson diversity / S) 3) 'pielou': Pielou's evenness (Pielou, 1966), also known as Shannon or Shannon-Weaver/Wiener/Weiner evenness; H/ln(S). The Shannon-Weaver is the preferred term; see A tribute to Claude Shannon (1916 –2001) and a plea for more rigorous use of species richness, species diversity and the 'Shannon–Wiener' Index. Spellerberg and Fedor. Alpha Ecology & Biogeography (2003) 12, 177–197 4) 'evar': Smith and Wilson's Evar index (Smith & Wilson 1996) 5) 'bulla': Bulla's index (O) (Bulla 1994)

Desirable statistical evenness metrics avoid strong bias towards very large or very small abundances; are independent of richness; and range within [0,1] with increasing evenness (Smith & Wilson 1996). Evenness metrics that fulfill these criteria include at least camargo, simpson, smith-wilson, and bulla. Also see Magurran & McGill (2011) and Beisel et al. (2003) for further details.

Value

A vector of evenness indices

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

References

Beisel J-N. et al. A Comparative Analysis of Evenness Index Sensitivity. Internal Rev. Hydrobiol. 88(1):3-15, 2003. URL: https://portais.ufg.br/up/202/o/2003-comparative_evennes_index.pdf

Bulla L. An index of evenness and its associated diversity measure. Oikos 70:167–171, 1994

Camargo, JA. New diversity index for assessing structural alterations in aquatic communities. Bull. Environ. Contam. Toxicol. 48:428–434, 1992.

Locey KJ and Lennon JT. Scaling laws predict global microbial diversity. PNAS 113(21):5970-5975, 2016; doi:10.1073/pnas.1521291113.

Magurran AE, McGill BJ, eds (2011) Biological Diversity: Frontiers in Measurement and Assessment (Oxford Univ Press, Oxford), Vol 12.

Pielou, EC. The measurement of diversity in different types of biological collections. Journal of Theoretical Biology 13:131–144, 1966.

Smith B and Wilson JB. A Consumer's Guide to Evenness Indices. Oikos 76(1):70-82, 1996.

See Also

coverage, core_abundance, rarity, alpha

Examples

```
data(dietswap)
# phyloseq object
#d <- evenness(dietswap, 'pielou')
# matrix
#d <- evenness(abundances(dietswap), 'pielou')
# vector
d <- evenness(abundances(dietswap)[,1], 'pielou')
```

---

find_optima *Find Optima*

Description

Detect optima, excluding local optima below peak.threshold.

Usage

```
find_optima(f, peak.threshold = 0, bw = 1, min.density = 1)
```

Arguments

f	density
peak.threshold	Mode detection threshold
bw	bandwidth
min.density	Minimun accepted density for a maximum; as a multiple of kernel height

Value

A list with min (minima), max (maxima), and peak.threshold (minimum detection density)

Author(s)

Leo Lahti <leo.lahti@iki.fi>

References

See citation('microbiome')

Examples

```
# Not exported
# o <- find_optima(rnorm(100), bw=1)
```

---

gktau *gktau*

Description

Measure association between nominal (no order for levels) variables

Usage

```
gktau(x, y)
```

Arguments

x	first variable
y	second variable

Details

Measure association between nominal (no order for levels) variables using Goodman and Kruskal tau. Code modified from the original source: r-bloggers.com/measuring-associations-between-non-numeric-variables/ An important feature of this procedure is that it allows missing values in either of the variables x or y, treating 'missing' as an additional level. In practice, this is sometimes very important since missing values in one variable may be strongly associated with either missing values in another variable or specific non-missing levels of that variable. An important characteristic of Goodman and Kruskal's tau measure is its asymmetry: because the variables x and y enter this expression differently, the value of a(y,x) is not the same as the value of a(x, y), in general. This stands in marked contrast to either the product-moment correlation coefficient or the Spearman rank correlation coefficient, which are both symmetric, giving the same association between x and y as that between y and x. The fundamental reason for the asymmetry of the general class of measures defined above is that they quantify the extent to which the variable x is useful in predicting y, which may be very different than the extent to which the variable y is useful in predicting x.

Value

Dependency measure

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

References

Code modified from the original source: http://r-bloggers.com/measuring-associations-between-non-numeric-va
To cite the microbiome R package, see citation('microbiome')

Examples

```
data(peerj32)
v1 <- factor(peerj32$microbes[,1])
v2 <- factor(peerj32$meta$gender)
tc <- gktau(v1, v2)
```

---

group_age *Age Classes*

Description

Cut age information to discrete factors.

Usage

```
group_age(
  x,
  breaks = "decades",
  n = 10,
  labels = NULL,
  include.lowest = TRUE,
  right = FALSE,
  dig.lab = 3,
  ordered_result = FALSE
)
```

Arguments

x	Numeric vector (age in years)
breaks	Class break points. Either a vector of breakpoints, or one of the predefined options ("years", "decades", "even").
n	Number of groups for the breaks = "even" option.
labels	labels for the levels of the resulting category. By default, labels are constructed using "(a,b]" interval notation. If labels = FALSE, simple integer codes are returned instead of a factor.
include.lowest	logical, indicating if an 'x[i]' equal to the lowest (or highest, for right = FALSE) 'breaks' value should be included.
right	logical, indicating if the intervals should be closed on the right (and open on the left) or vice versa.
dig.lab	integer which is used when labels are not given. It determines the number of digits used in formatting the break numbers.
ordered_result	logical: should the result be an ordered factor?

Details

Regarding the breaks arguments, the "even" option aims to cut the samples in groups with approximately the same size (by quantiles). The "years" and "decades" options are self-explanatory.

Value

Factor of age groups.

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

References

See citation('microbiome')

See Also

base::cut

Examples

```
data(atlas1006)
age.numeric <- meta(atlas1006)$age
age.factor <- group_age(age.numeric)
```

---

group_bmi *Body-Mass Index (BMI) Classes*

Description

Cut BMI information to standard discrete factors.

Usage

```
group_bmi(
  x,
  breaks = "standard",
  n = 10,
  labels = NULL,
  include.lowest = TRUE,
  right = FALSE,
  dig.lab = 3,
  ordered_result = FALSE
)
```

Arguments

x	Numeric vector (BMI)
breaks	Class break points. Either a vector of breakpoints, or one of the predefined options ("standard", "standard_truncated", "even").
n	Number of groups for the breaks = "even" option.
labels	labels for the levels of the resulting category. By default, labels are constructed using "(a,b]" interval notation. If labels = FALSE, simple integer codes are returned instead of a factor.
include.lowest	logical, indicating if an 'x[i]' equal to the lowest (or highest, for right = FALSE) 'breaks' value should be included.
right	logical, indicating if the intervals should be closed on the right (and open on the left) or vice versa.
dig.lab	integer which is used when labels are not given. It determines the number of digits used in formatting the break numbers.
ordered_result	logical: should the result be an ordered factor?

Details

Regarding the breaks arguments, the "even" option aims to cut the samples in groups with approximately the same size (by quantiles). The "standard" option corresponds to standard obesity categories defined by the cutoffs <18.5 (underweight); <25 (lean); <30 (obese); <35 (severe obese); <40 (morbid obese); <45 (super obese). The standard_truncated combines the severe, morbid and super obese into a single group.

Value

Factor of BMI groups.

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

References

See citation('microbiome')

See Also

base::cut

Examples

```
bmi.numeric <- range(rnorm(100, mean = 25, sd = 3))
bmi.factor <- group_bmi(bmi.numeric)
```

---

heat *Association Heatmap*

Description

Visualizes n x m association table as heatmap.

Usage

```
heat(
  df,
  Xvar = names(df)[[1]],
  Yvar = names(df)[[2]],
  fill = names(df)[[3]],
  star = NULL,
  p.adj.threshold = 1,
  association.threshold = 0,
  step = 0.2,
  colours = c("darkblue", "blue", "white", "red", "darkred"),
  limits = NULL,
```

```
  legend.text = "",
  order.rows = TRUE,
  order.cols = TRUE,
  filter.significant = TRUE,
  star.size = NULL,
  plot.values = FALSE
)
```

Arguments

df	Data frame. Each row corresponds to a pair of associated variables. The columns give variable names, association scores and significance estimates.
Xvar	X axis variable column name. For instance 'X'.
Yvar	Y axis variable column name. For instance 'Y'.
fill	Column to be used for heatmap coloring. For instance 'association'.
star	Column to be used for cell highlighting. For instance 'p.adj'.
p.adj.threshold	Significance threshold for the stars.
association.threshold	Include only elements that have absolute association higher than this value
step	color interval
colours	heatmap colours
limits	colour scale limits
legend.text	legend text
order.rows	Order rows to enhance visualization interpretability. If this is logical, then hclust is applied. If this is a vector then the rows are ordered using this index.
order.cols	Order columns to enhance visualization interpretability. If this is logical, then hclust is applied. If this is a vector then the rows are ordered using this index.
filter.significant	Keep only the elements with at least one significant entry
star.size	NULL Determine size of the highlight symbols
plot.values	Show values as text

Value

ggplot2 object

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

References

See citation('microbiome')

Examples

```
data(peerj32)
d1 <- peerj32$lipids[, 1:10]
d2 <- peerj32$microbes[, 1:10]
cc <- associate(d1, d2, method='pearson')
p <- heat(cc, 'X1', 'X2', 'Correlation', star='p.adj')
```

`hitchip.taxonomy` *HITChip Taxonomy*

Description

HITChip taxonomy table.

Usage

```
data(hitchip.taxonomy)
```

Format

List with the element 'filtered', including a simplified version of the HITChip taxonomy.

Value

Loads the data set in R.

Author(s)

Leo Lahti <microbiome-admin@googlegroups.com>

References

Lahti et al. Tipping elements of the human intestinal ecosystem. Nature Communications 5:4344, 2014. To cite the microbiome R package, see citation('microbiome')

hotplot *Univariate Bimodality Plot*

Description

Coloured bimodality plot.

Usage

```
hotplot(
  x,
  taxon,
  tipping.point = NULL,
  lims = NULL,
  shift = 0.001,
  log10 = TRUE
)
```

Arguments

x	phyloseq-class object
taxon	Taxonomic group to visualize.
tipping.point	Indicate critical point for abundance variations to be highlighted.
lims	Optional. Figure X axis limits.
shift	Small constant to avoid problems with zeroes in log10
log10	Use log10 abundances for the OTU table and tipping point

Value

ggplot object

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

References

See citation('microbiome')

Examples

```
data(atlas1006)
pseq <- subset_samples(atlas1006, DNA_extraction_method == 'r')
pseq <- transform(pseq, 'compositional')
# Set a tipping point manually
tipp <- .3/100 # .3 percent relative abundance
# Bimodality is often best visible at log10 relative abundances
p <- hotplot(pseq, 'Dialister', tipping.point=tipp, log10=TRUE)
```

`inequality` *Gini Index*

Description

Calculate Gini indices for a phyloseq object.

Usage

```
inequality(x)
```

Arguments

`x`	`phyloseq-class` object

Details

Gini index is a common measure for relative inequality in economical income, but can also be used as a community diversity measure. Gini index is between [0,1], and increasing gini index implies increasing inequality.

Value

A vector of Gini indices

Author(s)

Contact: Leo Lahti `<microbiome-admin@googlegroups.com>`

References

Relative Distribution Methods in the Social Sciences. Mark S. Handcock and Martina Morris, Springer-Verlag, Inc., New York, 1999. ISBN 0387987789.

See Also

diversity, reldist::gini (inspired by that implementation but independently written here to avoid external depedencies)

Examples

```
data(dietswap)
d <- inequality(dietswap)
```

`intermediate_stability` *Intermediate Stability*

Description

Quantify intermediate stability with respect to a given reference point.

Usage

```
intermediate_stability(
  x,
  reference.point = NULL,
  method = "correlation",
  output = "scores"
)
```

Arguments

`x`	**phyloseq** object. Includes abundances (variables x samples) and sample_data data.frame (samples x features) with 'subject' and 'time' field for each sample.
`reference.point`	Calculate stability of the data w.r.t. this point. By default the intermediate range is used (min + (max - min)/2). If a vector of points is provided, then the scores will be calculated for every point and a data.frame is returned.
`method`	'lm' (linear model) or 'correlation'; the linear model takes time into account as a covariate
`output`	Specify the return mode. Either the 'full' set of stability analysis outputs, or the 'scores' of intermediate stability.

Details

Decomposes each column in x into differences between consecutive time points. For each variable and time point we calculate for the data values: (i) the distance from reference point; (ii) distance from the data value at the consecutive time point. The 'correlation' method calculates correlation between these two variables. Negative correlations indicate that values closer to reference point tend to have larger shifts in the consecutive time point. The 'lm' method takes the time lag between the consecutive time points into account as this may affect the comparison and is not taken into account by the straightforward correlation. Here the coefficients of the following linear model are used to assess stability: abs(change) ~ time + abs(start.reference.distance). Samples with missing data, and subjects with less than two time point are excluded. The absolute count data x is logarithmized before the analysis with the log10(1 + x) trick to circumvent logarithmization of zeroes.

Value

A list with following elements: stability: estimated stability data: processed data set used in calculations

Author(s)

Leo Lahti <leo.lahti@iki.fi>

Examples

```
data(atlas1006)
x <- subset_samples(atlas1006, DNA_extraction_method == 'r')
x <- prune_taxa(c('Akkermansia', 'Dialister'), x)
res <- intermediate_stability(x, reference.point=NULL)
```

---

is_compositional *Test Compositionality*

Description

Test if phyloseq object is compositional.

Usage

```
is_compositional(x, tolerance = 1e-06)
```

Arguments

x	phyloseq-class object
tolerance	Tolerance for detecting compositionality.

Details

This function tests that the sum of abundances within each sample is almost zero, within the tolerance of 1e-6 by default.

Value

Logical TRUE/FALSE

See Also

transform

Examples

```
data(dietswap)
a <- is_compositional(dietswap)
b <- is_compositional(transform(dietswap, "identity"))
c <- is_compositional(transform(dietswap, "compositional"))
```

log_modulo_skewness *Log-Modulo Skewness Rarity Index*

Description

Calculates the community rarity index by log-modulo skewness.

Usage

```
log_modulo_skewness(x, q = 0.5, n = 50)
```

Arguments

x	Abundance matrix (taxa x samples) with counts
q	Arithmetic abundance classes are evenly cut up to to this quantile of the data. The assumption is that abundances higher than this are not common, and they are classified in their own group.
n	The number of arithmetic abundance classes from zero to the quantile cutoff indicated by q.

Details

The rarity index characterizes the concentration of species at low abundance. Here, we use the skewness of the frequency distribution of arithmetic abundance classes (see Magurran & McGill 2011). These are typically right-skewed; to avoid taking log of occasional negative skews, we follow Locey & Lennon (2016) and use the log-modulo transformation that adds a value of one to each measure of skewness to allow logarithmization.

Value

A vector of rarity indices

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

References

Kenneth J. Locey and Jay T. Lennon. Scaling laws predict global microbial diversity. PNAS 2016 113 (21) 5970-5975; doi:10.1073/pnas.1521291113.

Magurran AE, McGill BJ, eds (2011) Biological Diversity: Frontiers in Measurement and Assessment (Oxford Univ Press, Oxford), Vol 12

See Also

core_abundance, low_abundance, alpha

Examples

```
data(dietswap)
d <- log_modulo_skewness(dietswap)
```

`low_abundance` *Low Abundance Index*

Description

Calculates the concentration of low-abundance taxa below the indicated detection threshold.

Usage

```
low_abundance(x, detection = 0.2/100)
```

Arguments

`x`	`phyloseq-class` object
`detection`	Detection threshold for absence/presence (strictly greater by default).

Details

The low_abundance index gives the concentration of species at low abundance, or the relative proportion of rare species in [0,1]. The species that are below the indicated detection threshold are considered rare. Note that population prevalence is not considered. If the detection argument is a vector, then a data.frame is returned, one column for each detection threshold.

Value

A vector of indicators.

Author(s)

Contact: Leo Lahti `<microbiome-admin@googlegroups.com>`

See Also

core_abundance, rarity, global

Examples

```
data(dietswap)
d <- low_abundance(dietswap, detection=0.2/100)
```

map_levels *Map Taxonomic Levels*

Description

Map taxa between hierarchy levels.

Usage

```
map_levels(taxa = NULL, from, to, data)
```

Arguments

taxa	taxa to convert; if NULL then considering all taxa in the tax.table
from	convert from taxonomic level
to	convert to taxonomic level
data	Either a phyloseq object or its taxonomyTable-class , see the **phyloseq** package.

Value

mappings

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

References

See citation('microbiome')

Examples

```
data(dietswap)
m <- map_levels('Akkermansia', from='Genus', to='Phylum',
tax_table(dietswap))
m <- map_levels('Verrucomicrobia', from='Phylum', to='Genus',
tax_table(dietswap))
```

merge_taxa2 *Merge Taxa*

Description

Merge taxonomic groups into a single group.

Usage

```
merge_taxa2(x, taxa = NULL, pattern = NULL, name = "Merged")
```

Arguments

x	phyloseq-class object
taxa	A vector of taxa names to merge.
pattern	Taxa that match this pattern will be merged.
name	Name of the merged group.

Details

In some cases it is necessary to place certain OTUs or other groups into an "other" category. For instance, unclassified groups. This wrapper makes this easy. This function differs from phyloseq::merge_taxa by the last two arguments. Here, in merge_taxa2 the user can specify the name of the new merged group. And the merging can be done based on common pattern in the name.

Value

Modified phyloseq object

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

References

See citation('microbiome')

Examples

```
data(dietswap)
s <- merge_taxa(dietswap, c())
```

meta *Retrieve Phyloseq Metadata as Data Frame*

Description

The output of the phyloseq::sample_data() function does not return data.frame, which is needed for many applications. This function retrieves the sample data as a data.frame

Usage

```
meta(x)
```

Arguments

x	a phyloseq object

Value

Sample metadata as a data.frame

Author(s)

Leo Lahti <leo.lahti@iki.fi>

See Also

sample_data in the **phyloseq** package

Examples

```
data(dietswap); df <- meta(dietswap)
```

multimodality *Multimodality Score*

Description

Multimodality score based on bootstrapped potential analysis.

Usage

```
multimodality(
  x,
  peak.threshold = 1,
  bw.adjust = 1,
  bs.iter = 100,
  min.density = 1,
  verbose = TRUE
)
```

Arguments

x	A vector, or data matrix (variables x samples)
peak.threshold	Mode detection threshold
bw.adjust	Bandwidth adjustment
bs.iter	Bootstrap iterations
min.density	minimum accepted density for a maximum; as a multiple of kernel height
verbose	Verbose

Details

Repeats potential analysis (Livina et al. 2010) multiple times with bootstrap sampling for each row of the input data (as in Lahti et al. 2014) and returns the specified results.

Value

A list with following elements:

- scoreFraction of bootstrap samples with multiple observed modes
- nmodesThe most frequently observed number of modes in bootstrap
- resultsFull results of potential_analysis for each row of the input matrix.

Author(s)

Leo Lahti <leo.lahti@iki.fi>

References

- Livina et al. (2010). Potential analysis reveals changing number of climate states during the last 60 kyr. *Climate of the Past*, 6, 77-82.
- Lahti et al. (2014). Tipping elements of the human intestinal ecosystem. *Nature Communications* 5:4344.

Examples

```
#data(peerj32)
#s <- multimodality(t(peerj32$microbes[, c('Akkermansia', 'Dialister')]))
```

`neat` *Neatmap Sorting*

Description

Order matrix or phyloseq OTU table based on the neatmap approach.

Usage

```
neat(
  x,
  arrange = "both",
  method = "NMDS",
  distance = "bray",
  first.feature = NULL,
  first.sample = NULL,
  ...
)
```

Arguments

`x`	A matrix or phyloseq object.
`arrange`	Order 'features', 'samples' or 'both' (for matrices). For matrices, it is assumed that the samples are on the columns and features are on the rows. For phyloseq objects, features are the taxa of the OTU table.
`method`	Ordination method. Only NMDS implemented for now.
`distance`	Distance method. See `vegdist` function from the **vegan** package.
`first.feature`	Optionally provide the name of the first feature to start the ordering
`first.sample`	Optionally provide the name of the first sample to start the ordering
`...`	Arguments to pass.

Details

Borrows elements from the heatmap implementation in the **phyloseq** package. The row/column sorting is not available there as a separate function. Therefore I implemented this function to provide an independent method for easy sample/taxon reordering for phyloseq objects. The ordering is cyclic so we can start at any point. The choice of the first sample may somewhat affect the overall ordering

Value

Sorted matrix

References

This function is partially based on code derived from the **phyloseq** package. However for the original neatmap approach for heatmap sorting, see (and cite): Rajaram, S., & Oono, Y. (2010). NeatMap–non-clustering heat map alternatives in R. BMC Bioinformatics, 11, 45.

Examples

```
data(peerj32)
# Take subset to speed up example
x <- peerj32$microbes[1:10,1:10]
xo <- neat(x, 'both', method='NMDS', distance='bray')
```

`neatsort` *Neatmap Sorting*

Description

Sort samples or features based on the neatmap approach.

Usage

```
neatsort(x, target, method = "NMDS", distance = "bray", first = NULL, ...)
```

Arguments

`x`	`phyloseq-class` object or a matrix
`target`	For `phyloseq-class` input, the target is either 'sites' (samples) or 'species' (features) (taxa/OTUs); for matrices, the target is 'rows' or 'cols'.
`method`	Ordination method. See `ordinate` from **phyloseq** package. For matrices, only the NMDS method is available.
`distance`	Distance method. See `ordinate` from **phyloseq** package.
`first`	Optionally provide the name of the first sample/taxon to start the ordering (the ordering is cyclic so we can start at any point). The choice of the first sample may somewhat affect the overall ordering.
`...`	Arguments to be passed.

Details

This function borrows elements from the heatmap implementation in the **phyloseq** package. The row/column sorting is there not available as a separate function at present, however, hindering reuse in other tools. Implemented in the microbiome package to provide an independent method for easy sample/taxon reordering for phyloseq objects.

Value

Vector of ordered elements

References

This function is partially based on code derived from the **phyloseq** package. For the original neatmap approach for heatmap sorting, see (and cite): Rajaram, S., & Oono, Y. (2010). NeatMap–non-clustering heat map alternatives in R. BMC Bioinformatics, 11, 45.

Examples

```
data(peerj32)
pseq <- peerj32$phyloseq
# For Phyloseq
sort.otu <- neatsort(pseq, target='species')
# For matrix
# sort.rows <- neatsort(abundances(pseq), target='rows')
```

`overlap` *Overlap Measure*

Description

Quantify microbiota 'overlap' between samples.

Usage

```
overlap(x, detection = 0)
```

Arguments

`x`	`phyloseq-class` object
`detection`	Detection threshold.

Value

Overlap matrix

Author(s)

Contact: Leo Lahti `<microbiome-admin@googlegroups.com>`

References

Bashan, A., Gibson, T., Friedman, J. et al. Universality of human microbial dynamics. Nature 534, 259–262 (2016). https://doi.org/10.1038/nature18301

Examples

```
data(atlas1006)
o <- overlap(atlas1006, detection = 0.1/100)
```

peerj32 *Probiotics Intervention Data*

Description

The peerj32 data set contains high-through profiling data from 389 human blood serum lipids and 130 intestinal genus-level bacteria from 44 samples (22 subjects from 2 time points; before and after probiotic/placebo intervention). The data set can be used to investigate associations between intestinal bacteria and host lipid metabolism. For details, see http://dx.doi.org/10.7717/peerj.32.

Usage

```
data(peerj32)
```

Format

List of the following data matrices as described in detail in Lahti et al. (2013):

- lipids: Quantification of 389 blood serum lipids across 44 samples
- microbes: Quantification of 130 genus-like taxa across 44 samples
- meta: Sample metadata including time point, sex, subjectID, sampleID and treatment group (probiotic LGG / Placebo)
- phyloseq The microbiome data set converted into a phyloseq-class object.

Value

Loads the data set in R.

Author(s)

Leo Lahti <microbiome-admin@googlegroups.com>

References

Lahti et al. (2013) PeerJ 1:e32 http://dx.doi.org/10.7717/peerj.32

plot_atlas *Visualize Samples of a Microbiota Atlas*

Description

Show all samples of a microbiota collection, colored by specific factor levels (x axis) and signal (y axis).

Usage

```
plot_atlas(pseq, x, y, ncol = 2)
```

Arguments

pseq	phyloseq object
x	Sorting variable for X axis and sample coloring
y	Signal variable for Y axis
ncol	Number of legend columns.

Details

Arranges the samples based on the given grouping factor (x), and plots the signal (y) on the Y axis. The samples are randomly ordered within each factor level. The factor levels are ordered by standard deviation of the signal (y axis).

Value

ggplot object

Author(s)

Leo Lahti <leo.lahti@iki.fi>

References

See citation('microbiome'); Visualization inspired by Kilpinen et al. 2008, Genome Biology 9:R139. DOI: 10.1186/gb-2008-9-9-r139

Examples

```
data(atlas1006)
p <- plot_atlas(atlas1006, 'DNA_extraction_method', 'diversity')
p <- plot_atlas(atlas1006, 'DNA_extraction_method', 'Bifidobacterium')
```

plot_composition *Taxonomic Composition Plot*

Description

Plot taxon abundance for samples.

Usage

```
plot_composition(
  x,
  sample.sort = NULL,
  otu.sort = NULL,
  x.label = "sample",
  plot.type = "barplot",
  verbose = FALSE,
  average_by = NULL,
  group_by = NULL,
  ...
)
```

Arguments

x	phyloseq-class object
sample.sort	Order samples. Various criteria are available: • NULL or 'none': No sorting • A single character string: indicate the metadata field to be used for ordering. Or: if this string is found from the tax_table, then sort by the corresponding taxonomic group. • A character vector: sample IDs indicating the sample ordering. • 'neatmap' Order samples based on the neatmap approach. See neatsort. By default, 'NMDS' method with 'bray' distance is used. For other options, arrange the samples manually with the function.
otu.sort	Order taxa. Same options as for the sample.sort argument but instead of metadata, taxonomic table is used. Also possible to sort by 'abundance'.
x.label	Specify how to label the x axis. This should be one of the variables in sample_variables(x).
plot.type	Plot type: 'barplot' or 'heatmap'
verbose	verbose (but not in sample/taxon ordering). The options are 'Z-OTU', 'Z-Sample', 'log10' and 'compositional'. See the transform function.
average_by	Average the samples by the average_by variable
group_by	Group by this variable (in plot.type "barplot")
...	Arguments to be passed (for neatsort function)

Value

A ggplot plot object.

Examples

```
library(dplyr)
data(atlas1006)
pseq <- atlas1006 %>%
   subset_samples(DNA_extraction_method == "r") %>%
   aggregate_taxa(level = "Phylum") %>%
   transform(transform = "compositional")
p <- plot_composition(pseq, sample.sort = "Firmicutes",
         otu.sort = "abundance", verbose = TRUE) +
     scale_fill_manual(values = default_colors("Phylum")[taxa(pseq)])
```

---

plot_core *Visualize OTU Core*

Description

Core visualization (2D).

Usage

```
plot_core(
  x,
  prevalences = seq(0.1, 1, 0.1),
  detections = 20,
  plot.type = "lineplot",
  colours = NULL,
  min.prevalence = NULL,
  taxa.order = NULL,
  horizontal = FALSE
)
```

Arguments

x	A phyloseq object or a core matrix
prevalences	a vector of prevalence percentages in [0,1]
detections	a vector of intensities around the data range, or a scalar indicating the number of intervals in the data range.
plot.type	Plot type ('lineplot' or 'heatmap')
colours	colours for the heatmap
min.prevalence	If minimum prevalence is set, then filter out those rows (taxa) and columns (detections) that never exceed this prevalence. This helps to zoom in on the actual core region of the heatmap. Only affects the plot.type='heatmap'.
taxa.order	Ordering of the taxa: a vector of names.
horizontal	Logical. Horizontal figure.

Value

A list with three elements: the ggplot object and the data. The data has a different form for the lineplot and heatmap. Finally, the applied parameters are returned.

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

References

A Salonen et al. The adult intestinal core microbiota is determined by analysis depth and health status. Clinical Microbiology and Infection 18(S4):16 20, 2012. To cite the microbiome R package, see citation('microbiome')

Examples

```
data(dietswap)
p <- plot_core(transform(dietswap, "compositional"),
  prevalences=seq(0.1, 1, .1), detections=seq(0.01, 1, length = 10))
```

`plot_density` *Plot Density*

Description

Plot abundance density across samples for a given taxon.

Usage

```
plot_density(
  x,
  variable = NULL,
  log10 = FALSE,
  adjust = 1,
  kernel = "gaussian",
  trim = FALSE,
  na.rm = FALSE,
  fill = "gray",
  tipping.point = NULL,
  xlim = NULL
)
```

Arguments

x	phyloseq-class object or an OTU matrix (samples x phylotypes)
variable	OTU or metadata variable to visualize
log10	Logical. Show log10 abundances or not.
adjust	see stat_density
kernel	see stat_density
trim	see stat_density
na.rm	see stat_density
fill	Fill color
tipping.point	Optional. Indicate critical point for abundance variations to be highlighted.
xlim	X axis limits

Value

A ggplot plot object.

Examples

```
# Load gut microbiota data on 1006 western adults
# (see help(atlas1006) for references and details)
data(dietswap)
# Use compositional abundances instead of absolute signal
pseq.rel <- transform(dietswap, 'compositional')
# Population density for Dialister spp.; with log10 on the abundance (X)
# axis
library(ggplot2)
p <- plot_density(pseq.rel, variable='Dialister') + scale_x_log10()
```

---

plot_frequencies *Plot Frequencies*

Description

Plot relative frequencies within each Group for the levels of the given factor.

Usage

```
plot_frequencies(x, Groups, Factor)
```

Arguments

x	data.frame
Groups	Name of the grouping variable
Factor	Name of the frequency variable

Details

For table with the indicated frequencies, see the returned phyloseq object.

Value

ggplot plot object.

Examples

```
data(dietswap)
p <- plot_frequencies(meta(dietswap), 'group', 'sex')
```

---

plot_landscape *Landscape Plot*

Description

Wrapper for visualizing sample similarity landscape ie. sample density in various 2D projections.

Usage

```
plot_landscape(
  x,
  method = "PCoA",
  distance = "bray",
  transformation = "identity",
  col = NULL,
  main = NULL,
  x.ticks = 10,
  rounding = 0,
  add.points = TRUE,
  adjust = 1,
  size = 1,
  legend = FALSE,
  shading = TRUE,
  shading.low = "#ebf4f5",
  shading.high = "#e9b7ce",
  point.opacity = 0.75
)
```

Arguments

x	phyloseq-class object or a data matrix (samples x features; eg. samples vs. OTUs). If the input x is a 2D matrix then it is plotted as is.
method	Ordination method, see phyloseq::plot_ordination; or "PCA", or "t-SNE" (from the **Rtsne** package)

distance	Ordination distance, see phyloseq::plot_ordination; for method = "PCA", only euclidean distance is implemented now.
transformation	Transformation applied on the input object x
col	Variable name to highlight samples (points) with colors
main	title text
x.ticks	Number of ticks on the X axis
rounding	Rounding for X axis tick values
add.points	Plot the data points as well
adjust	Kernel width adjustment
size	point size
legend	plot legend TRUE/FALSE
shading	Add shading in the background.
shading.low	Color for shading low density regions
shading.high	Color for shading high density regions
point.opacity	Transparency-level for points

Details

For consistent results, set random seet (set.seed) before function call. Note that the distance and transformation arguments may have a drastic effect on the outputs.

Value

A ggplot plot object.

Examples

```
data(dietswap)

# PCoA
p <- plot_landscape(transform(dietswap, "compositional"),
   distance = "bray", method = "PCoA")

p <- plot_landscape(dietswap, method = "t-SNE", distance = "bray",
       transformation = "compositional")

# PCA
p <- plot_landscape(dietswap, method = "PCA", transformation = "clr")
```

plot_regression *Visually Weighted Regression Plot*

Description

Draw regression curve with smoothed error bars with Visually-Weighted Regression by Solomon M. Hsiang; see http://www.fight-entropy.com/2012/07/visually-weighted-regression.html The R is modified from Felix Schonbrodt's original code http://www.nicebread.de/ visually-weighted-watercolor-plots-new-variants-please-vote

Usage

```
plot_regression(
  formula,
  data,
  B = 1000,
  shade = TRUE,
  shade.alpha = 0.1,
  spag = FALSE,
  mweight = TRUE,
  show.lm = FALSE,
  show.median = TRUE,
  median.col = "white",
  show.CI = FALSE,
  method = loess,
  slices = 200,
  ylim = NULL,
  quantize = "continuous",
  show.points = TRUE,
  color = NULL,
  pointsize = NULL,
  ...
)
```

Arguments

formula	formula
data	data
B	number bootstrapped smoothers
shade	plot the shaded confidence region?
shade.alpha	shade.alpha: should the CI shading fade out at the edges? (by reducing alpha; 0=no alpha decrease, 0.1=medium alpha decrease, 0.5=strong alpha decrease)
spag	plot spaghetti lines?
mweight	visually weight the median smoother
show.lm	plot the linear regression line

show.median	show median smoother
median.col	median color
show.CI	should the 95% CI limits be plotted?
method	the fitting function for the spaghettis; default: loess
slices	number of slices in x and y direction for the shaded region. Higher numbers make a smoother plot, but takes longer to draw. I wouldn'T go beyond 500
ylim	restrict range of the watercoloring
quantize	either 'continuous', or 'SD'. In the latter case, we get three color regions for 1, 2, and 3 SD (an idea of John Mashey)
show.points	Show points.
color	Point colors
pointsize	Point sizes
...	further parameters passed to the fitting function, in the case of loess, for example, 'span=.9', or 'family='symmetric"

Value

ggplot2 object

Author(s)

Based on the original version from F. Schonbrodt. Modified by Leo Lahti <microbiome-admin@googlegroups.com>

References

See citation('microbiome')

Examples

```
data(atlas1006)
pseq <- subset_samples(atlas1006,
   DNA_extraction_method == 'r' &
   sex == "female" &
   nationality == "UKIE",
   B=10, slices=10 # non-default used here to speed up examples
   )
p <- plot_regression(diversity ~ age, meta(pseq)[1:20,], slices=10, B=10)
```

plot_taxa_prevalence *Visualize Prevalence Distributions for Taxa*

Description

Create taxa prevalence plots at various taxonomic levels.

Usage

```
plot_taxa_prevalence(x, level, detection = 0)
```

Arguments

x	phyloseq-class object, OTU data must be counts and not relative abundance or other transformed data.
level	Phylum/Order/Class/Family
detection	Detection threshold for presence (prevalance)

Details

This helps to obtain first insights into how is the taxa distribution in the data. It also gives an idea about the taxonomic affiliation of rare and abundant taxa in the data. This may be helpful for data filtering or other downstream analysis.

Value

A ggplot plot object.

Author(s)

Sudarshan A. Shetty <sudarshanshetty9@gmail.com>

Examples

```
data(atlas1006)
# Pick data subset just to speed up example
p0 <- subset_samples(atlas1006, DNA_extraction_method == "r")
p0 <- prune_taxa(taxa(p0)[grep("Bacteroides", taxa(p0))], p0)
# Detection threshold (0 by default; higher especially with HITChip)
p <- plot_taxa_prevalence(p0, 'Phylum', detection = 1)
print(p)
```

plot_tipping *Variation Line Plot*

Description

Plot variation in taxon abundance for many subjects.

Usage

```
plot_tipping(
  x,
  taxon,
  tipping.point = NULL,
  lims = NULL,
  shift = 0.001,
  xlim = NULL
)
```

Arguments

x	phyloseq-class object
taxon	Taxonomic group to visualize.
tipping.point	Optional. Indicate critical point for abundance variations to be highlighted.
lims	Optional. Figure X axis limits.
shift	Small constant to avoid problems with zeroes in log10
xlim	Horizontal axis limits

Details

Assuming the sample_data(x) has 'subject' field and some subjects have multiple time points.

Value

ggplot object

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

References

See citation('microbiome')

Examples

```
data(atlas1006)
pseq <- subset_samples(atlas1006, DNA_extraction_method == 'r')
pseq <- transform(pseq, 'compositional')
p <- plot_tipping(pseq, 'Dialister', tipping.point=1)
```

`potential_analysis` *Bootstrapped Potential Analysis*

Description

Analysis of multimodality based on bootstrapped potential analysis of Livina et al. (2010) as described in Lahti et al. (2014).

Usage

```
potential_analysis(
  x,
  peak.threshold = 0,
  bw.adjust = 1,
  bs.iter = 100,
  min.density = 1
)
```

Arguments

`x`	Input data vector
`peak.threshold`	Mode detection threshold
`bw.adjust`	Bandwidth adjustment
`bs.iter`	Bootstrap iterations
`min.density`	minimum accepted density for a maximum; as a multiple of kernel height

Value

List with following elements:

- modesNumber of modes for the input data vector (the most frequent number of modes from bootstrap)
- minimaAverage of potential minima across the bootstrap samples (for the most frequent number of modes)
- maximaAverage of potential maxima across the bootstrap samples (for the most frequent number of modes)
- unimodality.supportFraction of bootstrap samples exhibiting unimodality
- bwsBandwidths

References

- Livina et al. (2010). Potential analysis reveals changing number of climate states during the last 60 kyr. *Climate of the Past*, 6, 77-82.
- Lahti et al. (2014). Tipping elements of the human intestinal ecosystem. *Nature Communications* 5:4344.

See Also

plot_potential

Examples

```
# Example data; see help(peerj32) for details
data(peerj32)

# Log10 abundance of Dialister
x <- abundances(transform(peerj32$phyloseq, "clr"))['Dialister',]

# Bootstrapped potential analysis
# In practice, use more bootstrap iterations
# res <- potential_analysis(x, peak.threshold=0, bw.adjust=1,
#    bs.iter=9, min.density=1)
```

---

potential_univariate *Potential Analysis for Univariate Data*

Description

One-dimensional potential estimation for univariate timeseries.

Usage

```
potential_univariate(
  x,
  std = 1,
  bw = "nrd",
  weights = c(),
  grid.size = NULL,
  peak.threshold = 1,
  bw.adjust = 1,
  density.smoothing = 0,
  min.density = 1
)
```

Arguments

x	Univariate data (vector) for which the potentials shall be estimated
std	Standard deviation of the noise (defaults to 1; this will set scaled potentials)
bw	kernel bandwidth estimation method
weights	optional weights in ksdensity (used by potential_slidingaverages).
grid.size	Grid size for potential estimation. of density kernel height dnorm(0, sd=bandwidth)/N
peak.threshold	Mode detection threshold
bw.adjust	The real bandwidth will be bw.adjust*bw; defaults to 1
density.smoothing	Add a small constant density across the whole observation range to regularize density estimation (and to avoid zero probabilities within the observation range). This parameter adds uniform density across the observation range, scaled by density.smoothing.
min.density	minimum accepted density for a maximum; as a multiple of kernel height

Value

potential_univariate returns a list with the following elements:

- xi the grid of points on which the potential is estimated
- pot The estimated potential: -log(f)*std^2/2, where f is the density.
- density Density estimate corresponding to the potential.
- min.inds indices of the grid points at which the density has minimum values; (-potentials; neglecting local optima)
- max.inds indices the grid points at which the density has maximum values; (-potentials; neglecting local optima)
- bw bandwidth of kernel used
- min.points grid point values at which the density has minimum values; (-potentials; neglecting local optima)
- max.points grid point values at which the density has maximum values; (-potentials; neglecting local optima)

Author(s)

Based on Matlab code from Egbert van Nes modified by Leo Lahti. Extended from the initial version in the **earlywarnings** R package.

References

- Livina et al. (2010). Potential analysis reveals changing number of climate states during the last 60 kyr. *Climate of the Past*, 6, 77-82.
- Lahti et al. (2014). Tipping elements of the human intestinal ecosystem. *Nature Communications* 5:4344.

Examples

```
# res <- potential_univariate(x)
```

`prevalence` *OTU Prevalence*

Description

Simple prevalence measure.

Usage

```
prevalence(
  x,
  detection = 0,
  sort = FALSE,
  count = FALSE,
  include.lowest = FALSE
)
```

Arguments

`x`	A vector, data matrix or phyloseq object
`detection`	Detection threshold for absence/presence (strictly greater by default).
`sort`	Sort the groups by prevalence
`count`	Logical. Indicate prevalence as fraction of samples (in percentage [0, 1]; default); or in absolute counts indicating the number of samples where the OTU is detected (strictly) above the given abundance threshold.
`include.lowest`	Include the lower boundary of the detection and prevalence cutoffs. FALSE by default.

Details

For vectors, calculates the fraction (count=FALSE) or number (count=TRUE) of samples that exceed the detection. For matrices, calculates this for each matrix column. For phyloseq objects, calculates this for each OTU. The relative prevalence (count=FALSE) is simply the absolute prevalence (count=TRUE) divided by the number of samples.

Value

For each OTU, the fraction of samples where a given OTU is detected. The output is readily given as a percentage.

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

References

A Salonen et al. The adult intestinal core microbiota is determined by analysis depth and health status. Clinical Microbiology and Infection 18(S4):16 20, 2012. To cite the microbiome R package, see citation('microbiome')

Examples

```
data(peerj32)
pr <- prevalence(peerj32$phyloseq, detection=0, sort=TRUE, count=TRUE)
```

---

psmelt2 *Convert* phyloseq-class *object to long data format*

Description

An alternative to psmelt function from phyloseq-class object.

Usage

```
psmelt2(x, sample.column = NULL, feature.column = NULL)
```

Arguments

x	phyloseq-class object
sample.column	A single character string specifying name of the column to hold sample names.
feature.column	A single character string specifying name of the column to hold OTU or ASV names.

Value

A tibble in long format

Author(s)

Contact: Sudarshan A. Shetty <sudarshanshetty9@gmail.com>

Examples

```
data("dietswap")
ps.melt <- psmelt2(dietswap, sample.column="SampleID",
                   feature.column="Feature")
head(ps.melt)
```

quiet *Quiet Output*

Description

Suppress all output from an expression. Works cross-platform.

Usage

```
quiet(expr, all = TRUE)
```

Arguments

expr	Expression to run.
all	If TRUE then suppress warnings and messages as well; otherwise, only suppress printed output (such as from print or cat).

Value

Used for its side effects.

Author(s)

Adapted from https://gist.github.com/daattali/6ab55aee6b50e8929d89

Examples

```
quiet(1 + 1)
```

radial_theta *Radial Theta Function*

Description

Adapted from **NeatMap** and **phyloseq** packages but not exported and hence not available via phyloseq. Completely rewritten to avoid license conflicts. Vectorized to gain efficiency; only calculates theta and omits r.

Usage

```
radial_theta(x)
```

Arguments

x	position parameter

Value

theta

`rare` *Rare Microbiota*

Description

Filter the phyloseq object to include only rare (non-core) taxa.

Usage

```
rare(x, detection, prevalence, include.lowest = FALSE, ...)
```

Arguments

`x`	`phyloseq-class` object
`detection`	Detection threshold for absence/presence (strictly greater by default).
`prevalence`	Prevalence threshold (in [0, 1]; strictly greater by default)
`include.lowest`	Include the lower boundary of the detection and prevalence cutoffs in core calculation. FALSE by default.
`...`	Arguments to pass.

Value

Filtered phyloseq object including only rare taxa

Author(s)

Contact: Leo Lahti `<microbiome-admin@googlegroups.com>`

References

Salonen A, Salojarvi J, Lahti L, de Vos WM. The adult intestinal core microbiota is determined by analysis depth and health status. Clinical Microbiology and Infection 18(S4):16-20, 2012 To cite the microbiome R package, see citation('microbiome')

See Also

core_members

Examples

```
data(dietswap)
# Detection threshold 0 (strictly greater by default);
# Prevalence threshold 50 percent (strictly greater by default)
pseq <- rare(dietswap, 0, 50/100)
```

rare_abundance *Rare (Non-Core) Abundance Index*

Description

Calculates the rare abundance community index.

Usage

```
rare_abundance(
  x,
  detection = 0.1/100,
  prevalence = 50/100,
  include.lowest = FALSE
)
```

Arguments

x	phyloseq-class object
detection	Detection threshold for absence/presence (strictly greater by default).
prevalence	Prevalence threshold (in [0, 1]). The required prevalence is strictly greater by default. To include the limit, set include.lowest to TRUE.
include.lowest	Include the lower boundary of the detection and prevalence cutoffs. FALSE by default.

Details

This index gives the relative proportion of rare species (ie. those that are not part of the core microbiota) in the interval [0,1]. This is the complement (1-x) of the core abundance. The rarity function provides the abundance of the least abundant taxa within each sample, regardless of the population prevalence.

Value

A vector of indices

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

See Also

core_abundance, rarity, diversity

Examples

```
data(dietswap)
d <- rare_abundance(dietswap, detection=0.1/100, prevalence=50/100)
```

rare_members *Rare Taxa*

Description

Determine members of the rare microbiota with given abundance and prevalence threshold.

Usage

```
rare_members(x, detection = 1/100, prevalence = 50/100, include.lowest = FALSE)
```

Arguments

x	phyloseq-class object
detection	Detection threshold for absence/presence (strictly greater by default).
prevalence	Prevalence threshold (in [0, 1]). The required prevalence is strictly greater by default. To include the limit, set include.lowest to TRUE.
include.lowest	Include the lower boundary of the detection and prevalence cutoffs. FALSE by default.

Details

For phyloseq object, lists taxa that are less prevalent than the given prevalence threshold. Optionally, never exceeds the given abundance threshold (by default, all abundanecs accepted). For matrix, lists columns that satisfy these criteria.

Value

Vector of rare taxa

Author(s)

Leo Lahti <microbiome-admin@googlegroups.com>

References

To cite the microbiome R package, see citation('microbiome')

See Also

core_members

Examples

```
data(dietswap)
# Detection threshold: the taxa never exceed the given detection threshold
# Prevalence threshold 20 percent (strictly greater by default)
a <- rare_members(dietswap, detection=100/100, prevalence=20/100)
```

`rarity` *Rarity Index*

Description

Calculates the community rarity index.

Usage

```
rarity(x, index = "all", detection = 0.2/100, prevalence = 20/100)
```

Arguments

`x`	`phyloseq-class` object
`index`	If the index is given, it will override the other parameters. See the details below for description and references of the standard rarity indices.
`detection`	Detection threshold for absence/presence (strictly greater by default).
`prevalence`	Prevalence threshold (in [0, 1]). The required prevalence is strictly greater by default. To include the limit, set include.lowest to TRUE.

Details

The rarity index characterizes the concentration of species at low abundance.

The following rarity indices are provided:

- log_modulo_skewness Quantifies the concentration of the least abundant species by the log-modulo skewness of the arithmetic abundance classes (see Magurran & McGill 2011). These are typically right-skewed; to avoid taking log of occasional negative skews, we follow Locey & Lennon (2016) and use the log-modulo transformation that adds a value of one to each measure of skewness to allow logarithmization. The values q=0.5 and n=50 are used here.
- low_abundance Relative proportion of the least abundant species, below the detection level of 0.2%. The least abundant species are determined separately for each sample regardless of their prevalence.
- rare_abundance Relative proportion of the non-core species, exceed the given detection level (default 20 at the given prevalence (default 20 This is complement of the core with the same thresholds.

Value

A vector of rarity indices

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

References

Kenneth J. Locey and Jay T. Lennon. Scaling laws predict global microbial diversity. PNAS 2016 113 (21) 5970-5975; doi:10.1073/pnas.1521291113.

Magurran AE, McGill BJ, eds (2011) Biological Diversity: Frontiers in Measurement and Assessment (Oxford Univ Press, Oxford), Vol 12

See Also

alpha, log_modulo_skewness, rare_abundance, low_abundance

Examples

```
data(dietswap)
d <- rarity(dietswap, index='low_abundance')
# d <- rarity(dietswap, index='all')
```

`readcount` *Total Read Count*

Description

Total Read Count

Usage

```
readcount(x)
```

Arguments

x	phyloseq-class object

Value

Vector of read counts.

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

References

See citation('microbiome')

Examples

```
data(dietswap)
d <- readcount(dietswap)
```

`read_biom2phyloseq` *Read BIOM File into a Phyloseq Object*

Description

Read biom and mapping files into a `phyloseq-class` object.

Usage

```
read_biom2phyloseq(
  biom.file = NULL,
  taxonomy.file = NULL,
  metadata.file = NULL,
  ...
)
```

Arguments

`biom.file`	A biom file with '.biom' extension
`taxonomy.file`	NULL the latest version has taxonomic information within the biom
`metadata.file`	A simple metadata/mapping file with .csv extension
`...`	Arguments to pass for import_biom

Details

Biom file and mapping files will be converted to `phyloseq-class`.

Value

`phyloseq-class` object.

Author(s)

Sudarshan A. Shetty <sudarshanshetty9@gmail.com>

Examples

```
p0 <- read_biom2phyloseq()
#biom.file <- qiita1629.biom"
#meta.file <- qiita1629_mapping.csv"
#p0 <- read_biom2phyloseq(biom.file = biom.file,
#                        metadata.file = meta.file,
#                        taxonomy.file = NULL)
```

read_csv2phyloseq *Read Simple OTU Tables into a Phyloseq Object*

Description

Read simple OTU tables, mapping and taxonomy files into a phyloseq-class object.

Usage

```
read_csv2phyloseq(
  otu.file = NULL,
  taxonomy.file = NULL,
  metadata.file = NULL,
  sep = ","
)
```

Arguments

otu.file	A simple otu_table with '.csv' extension
taxonomy.file	A simple taxonomy file with '.csv' extension
metadata.file	A simple metadata/mapping file with .csv extension
sep	CSV file separator

Details

Simple OTU tables, mapping and taxonomy files will be converted to phyloseq-class.

Value

phyloseq-class object.

Author(s)

Sudarshan A. Shetty <sudarshanshetty9@gmail.com>

Examples

```
# NOTE: the system.file command reads these example files from the
# microbiome R package. To use your own local files, simply write
# otu.file <- "/path/to/my/file.csv" etc.

#otu.file <-
#   system.file("extdata/qiita1629_otu_table.csv",
#   package='microbiome')

#tax.file <- system.file("extdata/qiita1629_taxonomy_table.csv",
#        package='microbiome')
```

```
#meta.file <- system.file("extdata/qiita1629_mapping_subset.csv",
#     package='microbiome')

#p0 <- read_csv2phyloseq(
#        otu.file=otu.file,
#        taxonomy.file=tax.file,
#        metadata.file=meta.file)
```

`read_mothur2phyloseq` *Read Mothur Output into a Phyloseq Object*

Description

Read mothur shared and consensus taxonomy files into a phyloseq-class object.

Usage

```
read_mothur2phyloseq(shared.file, consensus.taxonomy.file, mapping.file = NULL)
```

Arguments

`shared.file`	A shared file produced by *mothur*. Identified from the .shared extension
`consensus.taxonomy.file`	Consensus taxonomy file produced by *mothur*. Identified from with the .taxonomy extension. See http://www.mothur.org/wiki/ConTaxonomy_file.
`mapping.file`	Metadata/mapping file with .csv extension

Details

Mothur shared and consensus taxonomy files will be converted to phyloseq-class.

Value

phyloseq-class object.

Author(s)

Sudarshan A. Shetty <sudarshanshetty9@gmail.com>

Examples

```
#otu.file <- system.file(
#"extdata/Baxter_FITs_Microbiome_2016_fit.final.tx.1.subsample.shared",
#   package='microbiome')

#tax.file <- system.file(
#"extdata/Baxter_FITs_Microbiome_2016_fit.final.tx.1.cons.taxonomy",
#   package='microbiome')
```

```
#meta.file <- system.file(
#"extdata/Baxter_FITs_Microbiome_2016_mapping.csv",
#   package='microbiome')

#p0 <- read_mothur2phyloseq(
#       shared.file=otu.file,
#       consensus.taxonomy.file=tax.file,
#       mapping.file=meta.file)
```

`read_phyloseq` *Import phyloseq Data*

Description

Read the otu, taxonomy and metadata from various formats.

Usage

```
read_phyloseq(
  otu.file = NULL,
  taxonomy.file = NULL,
  metadata.file = NULL,
  type = c("simple", "mothur", "biom"),
  sep = ","
)
```

Arguments

`otu.file`	File containing the OTU table (for mothur this is the file with the .shared extension)
`taxonomy.file`	(for mothur this is typically the consensus taxonomy file with the .taxonomy extension)
`metadata.file`	File containing samples x variables metadata.
`type`	Input data type: 'mothur' or 'simple' or 'biom' type.
`sep`	CSV file separator

Details

See help(read_mothur2phyloseq) for details on the Mothur input format; and help(read_biom2phyloseq) for details on the biom format. The simple format refers to the set of CSV files.

Value

`phyloseq-class` object

Author(s)

Sudarshan A. Shetty <sudarshanshetty9@gmail.com>

Examples

```
#  pseq <- read_phyloseq(otu.file,
#                     taxonomy.file,
#                     metadata.file,
#                     type=c('mothur', 'simple', 'biom'))
```

---

remove_samples *Exclude Samples*

Description

Filter out selected samples from a phyloseq object.

Usage

```
remove_samples(samples = NULL, x)
```

Arguments

samples	Names of samples to be removed.
x	phyloseq-class object

Details

This complements the phyloseq function prune_samples by providing a way to exclude given groups from a phyloseq object.

Value

Filtered phyloseq object.

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

References

To cite the microbiome R package, see citation('microbiome')

See Also

phyloseq::prune_samples, phyloseq::subset_samples

Examples

```
data(dietswap)
pseq <- remove_samples(c("Sample-182", "Sample-222"), dietswap)
```

`remove_taxa` *Exclude Taxa*

Description

Filter out selected taxa from a phyloseq object.

Usage

```
remove_taxa(taxa = NULL, x)
```

Arguments

`taxa`	Names of taxa to be removed.
`x`	`phyloseq-class` object

Details

This complements the phyloseq function prune_taxa by providing a way to exclude given groups from a phyloseq object.

Value

Filtered phyloseq object.

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

References

To cite the microbiome R package, see citation('microbiome')

See Also

phyloseq::prune_taxa, phyloseq::subset_taxa

Examples

```
data(dietswap)
pseq <- remove_taxa(c("Akkermansia", "Dialister"), dietswap)
```

richness *Richness Index*

Description

Community richness index.

Usage

```
richness(x, index = c("observed", "chao1"), detection = 0)
```

Arguments

x	A species abundance vector, or matrix (taxa/features x samples) with the absolute count data (no relative abundances), or phyloseq-class object
index	"observed" or "chao1"
detection	Detection threshold. Used for the "observed" index.

Details

By default, returns the richness for multiple detection thresholds defined by the data quantiles. If the detection argument is provided, returns richness with that detection threshold. The "observed" richness corresponds to index="observed", detection=0.

Value

A vector of richness indices

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

See Also

alpha

Examples

```
data(dietswap)
d <- richness(dietswap, detection=0)
```

spreadplot *Abundance Spread Plot*

Description

Visualize abundance spread for OTUs

Usage

```
spreadplot(x, trunc = 0.001/100, alpha = 0.15, width = 0.35)
```

Arguments

x	phyloseq-class object; or a data.frame with fields "otu" (otu name); "sample" (sample name); and "abundance" (otu abundance in the given sample)
trunc	Truncate abundances lower than this to zero
alpha	Alpha level for point transparency
width	Width for point spread

Value

ggplot2 object

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

References

See citation('microbiome')

Examples

```
data(dietswap)
p <- spreadplot(transform(dietswap, "compositional"))
```

summarize_phyloseq *Summarize phyloseq object*

Description

Prints basic information of data.

Usage

```
summarize_phyloseq(x)
```

Arguments

x	Input is a phyloseq-class object.

Details

The summarize_phyloseq function will give information on weather data is compositional or not, reads (min. max, median, average), sparsity, presence of singletons and sample variables.

Value

Prints basic information of phyloseq-class object.

Author(s)

Contact: Sudarshan A. Shetty <sudarshanshetty9@gmail.com>

Examples

```
data(dietswap)
summarize_phyloseq(dietswap)
```

taxa *Taxa Names*

Description

List the names of taxonomic groups in a phyloseq object.

Usage

```
taxa(x)
```

Arguments

x	phyloseq-class object

Details

A handy shortcut for phyloseq::taxa_names, with a potential to add to add some extra tweaks later.

Value

A vector with taxon names.

Author(s)

Contact: Leo Lahti <microbiome-admin@googlegroups.com>

References

To cite the microbiome R package, see citation('microbiome')

Examples

```
data(dietswap)
x <- taxa(dietswap)
```

TibbleUtilites *Utilities For* phyloseq-class *Slots to Tibbles*

Description

Utility to convert phyloseq slots to tibbles.

Usage

```
otu_tibble(x, column.id = "FeatureID")

tax_tibble(x, column.id = "FeatureID")

sample_tibble(x, column.id = "SampleID")

combine_otu_tax(x, column.id = "FeatureID")
```

Arguments

x	phyloseq-class object.
column.id	Provide name for the column which will hold the rownames. of slot.

Details

Convert different phyloseq slots into tibbles. otu_tibble gets the otu_table in tibble format. tax_tibble gets the taxa_table in tibble format. combine_otu_tax combines otu_table and taxa_table into one tibble.

Value

A tibble

Author(s)

Contact: Sudarshan A. Shetty <sudarshanshetty9@gmail.com>

Examples

```
library(microbiome)
data("dietswap")
otu_tib <- otu_tibble(dietswap,column.id="FeatureID")
tax_tib <- tax_tibble(dietswap,column.id="FeatureID")
sample_tib <- sample_tibble(dietswap,column.id="SampleID")
otu_tax <- combine_otu_tax(dietswap,column.id = "FeatureID")
head(otu_tax)
```

---

timesplit *Time Split*

Description

For each subject, return temporally consecutive sample pairs together with the corresponding time difference.

Usage

```
timesplit(x)
```

Arguments

x	**phyloseq** object.

Value

data.frame

Author(s)

Leo Lahti <leo.lahti@iki.fi>

Examples

```
data(atlas1006)
x <- timesplit(subset_samples(atlas1006,
  DNA_extraction_method == 'r' & sex == "male"))
```

time_normalize *Normalize Phyloseq Metadata Time Field*

Description

Shift the time field in phyloseq sample_data such that the first time point of each subject is always 0.

Usage

```
time_normalize(x)
```

Arguments

x	phyloseq object. The sample_data(x) should contain the following fields: subject, time

Value

Phyloseq object with a normalized time field

Examples

```
data(peerj32)
pseq <- time_normalize(peerj32$phyloseq)
```

time_sort *Temporal Sorting Within Subjects*

Description

Within each subject, sort samples by time and calculate distance from the baseline point (minimum time).

Usage

```
time_sort(x)
```

Arguments

x	A metadata data.frame including the following columns: time, subject, sample, signal. Or a phyloseq object.

Value

A list with sorted metadata (data.frame) for each subject.

Author(s)

Leo Lahti <leo.lahti@iki.fi>

References

See citation('microbiome')

Examples

```
data(atlas1006)
pseq <- subset_samples(atlas1006, DNA_extraction_method == "r")
ts <- time_sort(meta(pseq))
```

---

top *Identify Top Entries*

Description

Identify top entries in a vector or given field in data frame.

Usage

```
top(
  x,
  field = NULL,
  n = NULL,
  output = "vector",
  round = NULL,
  na.rm = FALSE,
  include.rank = FALSE
)
```

Arguments

x	data.frame, matrix, or vector
field	Field or column to check for a data.frame or matrix
n	Number of top entries to show
output	Output format: vector or data.frame
round	Optional rounding
na.rm	Logical. Remove NA before calculating the statistics.
include.rank	Include ranking if the output is data.frame. Logical.

Value

Vector of top counts, named by the corresponding entries

Author(s)

Leo Lahti <leo.lahti@iki.fi>

References

See citation("bibliographica")

Examples

```
data(dietswap)
p <- top(meta(dietswap), "group", 10)
```

---

top_taxa *Top Taxa*

Description

Return n most abundant taxa (based on total abundance over all samples), sorted by abundance

Usage

```
top_taxa(x, n = ntaxa(x))
```

Arguments

x	phyloseq object
n	Number of top taxa to return (default: all)

Value

Character vector listing the top taxa

Examples

```
data(dietswap)
topx <- top_taxa(dietswap, n=10)
```

transform *Data Transformations for phyloseq Objects*

Description

Standard transforms for phyloseq-class.

Usage

```
transform(
  x,
  transform = "identity",
  target = "OTU",
  shift = 0,
  scale = 1,
  log10 = TRUE,
  reference = 1,
  ...
)
```

Arguments

x	phyloseq-class object
transform	Transformation to apply. The options include: 'compositional' (ie relative abundance), 'Z', 'log10', 'log10p', 'hellinger', 'identity', 'clr', 'alr', or any method from the vegan::decostand function.
target	Apply the transform for 'sample' or 'OTU'. Does not affect the log transform.
shift	A constant indicating how much to shift the baseline abundance (in transform='shift')
scale	Scaling constant for the abundance values when transform = "scale".
log10	Used only for Z transformation. Apply log10 before Z.
reference	Reference feature for the alr transformation.
...	arguments to be passed

Details

In transformation typ, the 'compositional' abundances are returned as relative abundances in [0, 1] (convert to percentages by multiplying with a factor of 100). The Hellinger transform is square root of the relative abundance but instead given at the scale [0,1]. The log10p transformation refers to log10(1 + x). The log10 transformation is applied as log10(1 + x) if the data contains zeroes. CLR transform applies a pseudocount of min(relative abundance)/2 to exact zero relative abundance entries in OTU table before taking logs.

Value

Transformed phyloseq object

Examples

```
data(dietswap)
x <- dietswap

# No transformation
xt <- transform(x, 'identity')

# OTU relative abundances
# xt <- transform(x, 'compositional')

# Z-transform for OTUs
# xt <- transform(x, 'Z', 'OTU')

# Z-transform for samples
# xt <- transform(x, 'Z', 'sample')

# Log10 transform (log10(1+x) if the data contains zeroes)
# xt <- transform(x, 'log10')

# Log10p transform (log10(1+x) always)
# xt <- transform(x, 'log10p')

# CLR transform
# Note that small pseudocount is added if data contains zeroes
xt <- microbiome::transform(x, 'clr')

# ALR transform
# The pseudocount must be specified explicitly
# The reference feature is 1 by default
xt <- microbiome::transform(x, 'alr', shift=1, reference=1)

# Shift the baseline
# xt <- transform(x, 'shift', shift=1)

# Scale
# xt <- transform(x, 'scale', scale=1)
```

`ztransform` *Z Transformation*

Description

Z transform for matrices

Usage

```
ztransform(x, which, log10 = TRUE)
```

Arguments

x	a matrix
which	margin
log10	apply log10 transformation before Z

Details

Performs centering (to zero) and scaling (to unit variance) across samples for each taxa.

Value

Z-transformed matrix

Author(s)

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References

See citation('microbiome')

Examples

```
#data(peerj32)
#pseqz <- ztransform(abundances(peerj32$phyloseq))
```

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