

Introduction to RBM package

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1 Overview

This document provides an introduction to the `RBM` package. The `RBM` package executes the resampling-based empirical Bayes approach using either permutation or bootstrap tests based on moderated t-statistics through the following steps.

- Firstly, the `RBM` package computes the moderated t-statistics based on the observed data set for each feature using the `lmFit` and `eBayes` function.
- Secondly, the original data are permuted or bootstrapped in a way that matches the null hypothesis to generate permuted or bootstrapped resamples, and the reference distribution is constructed using the resampled moderated t-statistics calculated from permutation or bootstrap resamples.
- Finally, the p-values from permutation or bootstrap tests are calculated based on the proportion of the permuted or bootstrapped moderated t-statistics that are as extreme as, or more extreme than, the observed moderated t-statistics.

Additional detailed information regarding resampling-based empirical Bayes approach can be found elsewhere (Li et al., 2013).

2 Getting started

The RBM package can be installed and loaded through the following R code.
Install the RBM package with:

```
> if (!requireNamespace("BiocManager", quietly=TRUE))
+   install.packages("BiocManager")
> BiocManager::install("RBM")
```

Load the RBM package with:

```
> library(RBM)
```

3 RBM_T and RBM_F functions

There are two functions in the RBM package: `RBM_T` and `RBM_F`. Both functions require input data in the matrix format with rows denoting features and columns denoting samples. `RBM_T` is used for two-group comparisons such as study designs with a treatment group and a control group. `RBM_F` can be used for more complex study designs such as more than two groups or time-course studies. Both functions need a vector for group notation, i.e., "1" denotes the treatment group and "0" denotes the control group. For the `RBM_F` function, a contrast vector need to be provided by users to perform pairwise comparisons between groups. For example, if the design has three groups (0, 1, 2), the `aContrast` parameter will be a vector such as ("X1-X0", "X2-X1", "X2-X0") to denote all pairwise comparisons. Users just need to add an extra "X" before the group labels to do the contrasts.

- Examples using the `RBM_T` function: `normdata` simulates a standardized gene expression data and `unifdata` simulates a methylation microarray data. The p -values from the `RBM_T` function could be further adjusted using the `p.adjust` function in the `stats` package through the Benjamini-Hochberg method.

```
> library(RBM)
> normdata <- matrix(rnorm(1000*6, 0, 1),1000,6)
> mydesign <- c(0,0,0,1,1,1)
> myresult <- RBM_T(normdata,mydesign,100,0.05)
> summary(myresult)
```

	Length	Class	Mode
ordfit_t	1000	-none-	numeric
ordfit_pvalue	1000	-none-	numeric
ordfit_beta0	1000	-none-	numeric
ordfit_beta1	1000	-none-	numeric
permutation_p	1000	-none-	numeric
bootstrap_p	1000	-none-	numeric

```
> sum(myresult$permutation_p<=0.05)
```

```

[1] 34

> which(myresult$permutation_p<=0.05)

[1] 2 40 106 148 189 200 209 280 284 316 348 413 489 500 560 569 625 644 670
[20] 699 709 736 749 760 764 795 826 831 834 866 911 939 987 992

> sum(myresult$bootstrap_p<=0.05)

[1] 20

> which(myresult$bootstrap_p<=0.05)

[1] 93 106 109 142 342 433 486 497 498 560 682 690 737 749 751 756 789 826 866
[20] 954

> permutation_adj_p <- p.adjust(myresult$permutation_p, "BH")
> sum(permutation_adj_p<=0.05)

[1] 6

> bootstrap_adj_p <- p.adjust(myresult$bootstrap_p, "BH")
> sum(bootstrap_adj_p<=0.05)

[1] 0

> unifdata <- matrix(runif(1000*7,0.10, 0.95), 1000, 7)
> mydesign2 <- c(0,0,0, 1,1,1,1)
> myresult2 <- RBM_T(unifdata,mydesign2,100,0.05)
> sum(myresult2$permutatioin_p<=0.05)

[1] 0

> sum(myresult2$bootstrap_p<=0.05)

[1] 41

> which(myresult2$bootstrap_p<=0.05)

[1] 40 53 120 136 165 169 189 195 242 254 259 312 333 364 388 417 459 466 472
[20] 515 571 583 617 621 649 680 684 685 712 714 728 811 835 844 851 903 933 975
[39] 976 986 991

> bootstrap2_adj_p <- p.adjust(myresult2$bootstrap_p, "BH")
> sum(bootstrap2_adj_p<=0.05)

[1] 2

```

- Examples using the `RBM_F` function: `normdata_F` simulates a standardized gene expression data and `unifdata_F` simulates a methylation microarray data. In both examples, we were interested in pairwise comparisons.

```
> normdata_F <- matrix(rnorm(1000*9,0,2), 1000, 9)
> mydesign_F <- c(0, 0, 0, 1, 1, 1, 2, 2, 2)
> aContrast <- c("X1-X0", "X2-X1", "X2-X0")
> myresult_F <- RBM_F(normdata_F, mydesign_F, aContrast, 100, 0.05)
> summary(myresult_F)
```

	Length	Class	Mode
ordfit_t	3000	-none-	numeric
ordfit_pvalue	3000	-none-	numeric
ordfit_beta1	3000	-none-	numeric
permutation_p	3000	-none-	numeric
bootstrap_p	3000	-none-	numeric

```
> sum(myresult_F$permutation_p[, 1]<=0.05)

[1] 58

> sum(myresult_F$permutation_p[, 2]<=0.05)

[1] 54

> sum(myresult_F$permutation_p[, 3]<=0.05)

[1] 48

> which(myresult_F$permutation_p[, 1]<=0.05)

[1] 2 25 29 30 44 48 60 80 128 189 213 219 221 244 245 255 258 282 290
[20] 298 304 373 377 385 394 412 432 435 478 494 497 501 514 527 528 554 574 588
[39] 614 637 641 647 652 666 708 711 715 716 718 769 783 816 862 868 876 884 936
[58] 952

> which(myresult_F$permutation_p[, 2]<=0.05)

[1] 1 2 25 29 30 44 48 60 80 87 128 189 213 219 221 244 245 255 258
[20] 282 304 373 382 385 394 412 435 478 494 497 500 501 514 527 528 554 574 588
[39] 637 641 647 652 658 708 711 715 716 728 816 862 868 884 936 952

> which(myresult_F$permutation_p[, 3]<=0.05)

[1] 2 29 48 80 189 213 221 244 255 258 282 298 304 319 323 373 377 385 394
[20] 412 432 435 478 494 497 500 501 574 588 614 631 641 647 652 658 666 711 715
[39] 718 728 783 816 841 876 884 928 936 952
```

```

> con1_adjp <- p.adjust(myresult_F$permutation_p[, 1], "BH")
> sum(con1_adjp<=0.05/3)

[1] 11

> con2_adjp <- p.adjust(myresult_F$permutation_p[, 2], "BH")
> sum(con2_adjp<=0.05/3)

[1] 11

> con3_adjp <- p.adjust(myresult_F$permutation_p[, 3], "BH")
> sum(con3_adjp<=0.05/3)

[1] 2

> which(con2_adjp<=0.05/3)

[1] 30 244 373 385 478 574 647 652 708 816 884

> which(con3_adjp<=0.05/3)

[1] 501 816

> unifdata_F <- matrix(runif(1000*18, 0.15, 0.98), 1000, 18)
> mydesign2_F <- c(rep(0, 6), rep(1, 6), rep(2, 6))
> aContrast <- c("X1-X0", "X2-X1", "X2-X0")
> myresult2_F <- RBM_F(unifdata_F, mydesign2_F, aContrast, 100, 0.05)
> summary(myresult2_F)

              Length Class  Mode
ordfit_t      3000   -none-  numeric
ordfit_pvalue 3000   -none-  numeric
ordfit_beta1   3000   -none-  numeric
permutation_p 3000   -none-  numeric
bootstrap_p    3000   -none-  numeric

> sum(myresult2_F$bootstrap_p[, 1]<=0.05)

[1] 38

> sum(myresult2_F$bootstrap_p[, 2]<=0.05)

[1] 47

> sum(myresult2_F$bootstrap_p[, 3]<=0.05)

[1] 44

```

```

> which(myresult2_F$bootstrap_p[, 1]<=0.05)

[1] 20 42 50 66 155 177 224 254 272 293 355 386 403 422 469 487 563 565 580
[20] 591 610 626 629 667 679 699 710 726 782 792 802 804 815 826 856 871 873 943

> which(myresult2_F$bootstrap_p[, 2]<=0.05)

[1] 20 42 50 66 155 177 248 254 263 272 287 293 343 355 386 403 422 441 469
[20] 487 563 580 591 608 610 629 634 657 679 699 710 738 746 765 772 782 792 801
[39] 802 804 815 822 856 873 889 977 979

> which(myresult2_F$bootstrap_p[, 3]<=0.05)

[1] 20 42 66 155 177 207 248 263 272 287 293 345 355 376 403 412 422 469 487
[20] 563 580 590 591 610 629 634 667 679 699 710 726 738 772 782 792 802 804 815
[39] 856 858 861 873 977 979

> con21_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 1], "BH")
> sum(con21_adj_p<=0.05/3)

[1] 2

> con22_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 2], "BH")
> sum(con22_adj_p<=0.05/3)

[1] 5

> con23_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 3], "BH")
> sum(con23_adj_p<=0.05/3)

[1] 4

```

4 Ovarian cancer methylation example using the RBM_T function

Two-group comparisons are the most common contrast in biological and biomedical field. The ovarian cancer methylation example is used to illustrate the application of RBM_T in identifying differentially methylated loci. The ovarian cancer methylation example is taken from the genome-wide DNA methylation profiling of United Kingdom Ovarian Cancer Population Study (UKOPS). This study used Illumina Infinium 27k Human DNA methylation Beadchip v1.2 to obtain DNA methylation profiles on over 27,000 CpGs in whole blood cells from 266 ovarian cancer women and 274 age-matched healthy controls. The data are downloaded from the NCBI GEO website with access number GSE19711. For illustration purpose, we chose the first 1000 loci in 8 randomly selected women with 4 ovarian cancer cases (pre-treatment) and 4 healthy controls. The following codes show the process of generating significant differential DNA methylation loci using the RBM_T function and presenting the results for further validation and investigations.

```

> system.file("data", package = "RBM")

```

```
[1] "/private/tmp/RtmpqbKQAS/Rinstf90e18c7ded1/RBM/data"
```

```
> data(ovarian_cancer_methylation)
> summary(ovarian_cancer_methylation)
```

IlmnID	Beta	exmdata2[, 2]	exmdata3[, 2]
cg00000292: 1	Min. :0.01058	Min. :0.01187	Min. :0.009103
cg00002426: 1	1st Qu.:0.04111	1st Qu.:0.04407	1st Qu.:0.041543
cg00003994: 1	Median :0.08284	Median :0.09531	Median :0.087042
cg00005847: 1	Mean :0.27397	Mean :0.28872	Mean :0.283729
cg00006414: 1	3rd Qu.:0.52135	3rd Qu.:0.59032	3rd Qu.:0.558575
cg00007981: 1	Max. :0.97069	Max. :0.96937	Max. :0.970155
(Other) :994		NA's :4	

exmdata4[, 2]	exmdata5[, 2]	exmdata6[, 2]	exmdata7[, 2]
Min. :0.01019	Min. :0.01108	Min. :0.01937	Min. :0.01278
1st Qu.:0.04092	1st Qu.:0.04059	1st Qu.:0.05060	1st Qu.:0.04260
Median :0.09042	Median :0.08527	Median :0.09502	Median :0.09362
Mean :0.28508	Mean :0.28482	Mean :0.27348	Mean :0.27563
3rd Qu.:0.57502	3rd Qu.:0.57300	3rd Qu.:0.52099	3rd Qu.:0.52240
Max. :0.96658	Max. :0.97516	Max. :0.96681	Max. :0.95974
	NA's :1		

exmdata8[, 2]
Min. :0.01357
1st Qu.:0.04387
Median :0.09282
Mean :0.28679
3rd Qu.:0.57217
Max. :0.96268

```
> ovarian_cancer_data <- ovarian_cancer_methylation[, -1]
> label <- c(1, 1, 0, 0, 1, 1, 0, 0)
> diff_results <- RBM_T(aData=ovarian_cancer_data, vec_trt=label, repetition=100, alpha=0.05)
> summary(diff_results)
```

	Length	Class	Mode
ordfit_t	1000	-none-	numeric
ordfit_pvalue	1000	-none-	numeric
ordfit_beta0	1000	-none-	numeric
ordfit_beta1	1000	-none-	numeric
permutation_p	1000	-none-	numeric
bootstrap_p	1000	-none-	numeric

```
> sum(diff_results$ordfit_pvalue<=0.05)
```

```
[1] 47
```

```
> sum(diff_results$permutation_p<=0.05)
```

```
[1] 76
```

```
> sum(diff_results$bootstrap_p<=0.05)
```

```
[1] 39
```

```
> ordfit_adj_p <- p.adjust(diff_results$ordfit_pvalue, "BH")
```

```
> sum(ordfit_adj_p<=0.05)
```

```
[1] 0
```

```
> perm_adj_p <- p.adjust(diff_results$permutation_p, "BH")
```

```
> sum(perm_adj_p<=0.05)
```

```
[1] 33
```

```
> boot_adj_p <- p.adjust(diff_results$bootstrap_p, "BH")
```

```
> sum(boot_adj_p<=0.05)
```

```
[1] 3
```

```
> diff_list_perm <- which(perm_adj_p<=0.05)
```

```
> diff_list_boot <- which(boot_adj_p<=0.05)
```

```
> sig_results_perm <- cbind(ovarian_cancer_methylation[diff_list_perm, ], diff_results$ordfit_t[diff_list_perm, ])
```

```
> print(sig_results_perm)
```

	IlmnID	Beta	exmdata2[, 2]	exmdata3[, 2]	exmdata4[, 2]
16	cg00014085	0.05906804	0.04518973	0.04211710	0.03665208
19	cg00016968	0.80628480	NA	0.81440820	0.83623180
83	cg00072216	0.04505377	0.04598964	0.04000674	0.03231534
95	cg00081975	0.03633894	0.04975194	0.06024723	0.05598723
103	cg00094319	0.73784280	0.73532960	0.75574900	0.73830220
106	cg00095674	0.07076291	0.05045181	0.03861991	0.03337576
131	cg00121904	0.15449580	0.17949750	0.23608110	0.24354150
146	cg00134539	0.61101320	0.53321780	0.45999340	0.46787420
219	cg00202702	0.04104248	0.04685628	0.03793627	0.03529329
237	cg00215066	0.94926640	0.95311870	0.94634910	0.94561120
245	cg00224508	0.04479948	0.04972043	0.04152814	0.04189373
259	cg00234961	0.04192170	0.04321576	0.05707140	0.05327565
280	cg00260778	0.64319890	0.60488960	0.56735060	0.53150910
283	cg00262415	0.03850601	0.04621248	0.03579758	0.03765227
285	cg00263760	0.09050395	0.10197760	0.14801710	0.12242400
349	cg00332745	0.04703361	0.04634372	0.03676908	0.04518837
437	cg00424946	0.04122172	0.04325330	0.03339863	0.02876798
520	cg00502442	0.03163993	0.03581662	0.02785063	0.02549502
627	cg00612467	0.04777553	0.03783457	0.05380982	0.05582291
632	cg00615377	0.11265030	0.16140570	0.19404450	0.17468600

743	cg00717862	0.07999436	0.07873347	0.06089359	0.06171374
764	cg00730260	0.90471270	0.90542290	0.91002680	0.91258610
772	cg00743372	0.03922780	0.02919634	0.02187972	0.02568053
804	cg00777121	0.04540701	0.05430304	0.04154242	0.04221162
848	cg00826384	0.05721674	0.05612171	0.06644259	0.06358381
851	cg00830029	0.58362500	0.59397870	0.64739610	0.67269640
887	cg00862290	0.43640520	0.54047160	0.60786800	0.56325950
911	cg00888479	0.07388961	0.07361080	0.10149800	0.09985076
928	cg00901493	0.03737166	0.03903724	0.04684618	0.04981432
931	cg00901704	0.05734342	0.04812868	0.04478214	0.03878488
939	cg00906183	0.03949030	0.04365079	0.03720015	0.03575748
957	cg00925229	0.04101549	0.04241425	0.04042823	0.03285262
979	cg00945507	0.13432250	0.23854600	0.34749760	0.28903340
	exmdata5[, 2]	exmdata6[, 2]	exmdata7[, 2]	exmdata8[, 2]	
16	0.04222944	0.05324246	0.03728026	0.04062589	
19	0.80831380	0.73306440	0.82968340	0.84917800	
83	0.04965089	0.04833366	0.03466159	0.04390894	
95	0.04561792	0.05115624	0.06068253	0.06168212	
103	0.67349260	0.73510200	0.75715920	0.78981220	
106	0.04693030	0.06837343	0.04534005	0.03709488	
131	0.17352980	0.12564280	0.18193170	0.20847670	
146	0.67191510	0.63137380	0.47929610	0.45428300	
219	0.04074652	0.05125000	0.03908795	0.04075583	
237	0.94837410	0.94665570	0.94089070	0.94600090	
245	0.04208405	0.05284988	0.03775905	0.03955271	
259	0.04030003	0.03996053	0.05086962	0.05445672	
280	0.61920530	0.61925200	0.46753250	0.55632410	
283	0.03746915	0.04200230	0.03014699	0.02903290	
285	0.11693600	0.10650430	0.12281160	0.12310430	
349	0.04975075	0.05253778	0.04444665	0.03717721	
437	0.03353116	0.03719167	0.03096761	0.03234779	
520	0.03111720	0.03189393	0.02415307	0.02941176	
627	0.04740551	0.05332965	0.05775211	0.05579710	
632	0.12573100	0.14483660	0.16338240	0.20130510	
743	0.07594936	0.09062161	0.06475791	0.07271878	
764	0.90575890	0.88760470	0.90756300	0.90946790	
772	0.02796053	0.03512214	0.02575992	0.02093909	
804	0.04911277	0.04872797	0.04261405	0.04474881	
848	0.05230160	0.06119713	0.06542751	0.06240686	
851	0.50820240	0.34657470	0.66276570	0.64634510	
887	0.50259740	0.40111730	0.56646700	0.54552980	
911	0.08633986	0.06765189	0.09070268	0.12417730	
928	0.04490690	0.04204062	0.05050039	0.05268215	
931	0.04497277	0.05751033	0.03089829	0.04423603	
939	0.03856975	0.06024309	0.03594439	0.03502819	

957	0.03950725	0.05103829	0.03825804	0.03826061
979	0.11848510	0.16653850	0.30718420	0.26624740

diff_results\$ordfit_t[diff_list_perm]

16	1.954876
19	-2.547097
83	1.947226
95	-2.654324
103	-2.343784
106	2.887876
131	-3.562745
146	5.636263
219	1.375603
237	1.021426
245	1.494678
259	-2.833203
280	4.337628
283	1.601804
285	-2.993292
349	1.659117
437	1.574598
520	1.319602
627	-1.797392
632	-3.722206
743	2.918806
764	-1.560713
772	1.885560
804	1.445572
848	-1.687144
851	-2.986319
887	-3.368752
911	-3.490240
928	-1.982308
931	2.127264
939	1.558215
957	1.209487
979	-4.968792

diff_results\$permutation_p[diff_list_perm]

16	0
19	0
83	0
95	0
103	0
106	0
131	0
146	0

219	0
237	0
245	0
259	0
280	0
283	0
285	0
349	0
437	0
520	0
627	0
632	0
743	0
764	0
772	0
804	0
848	0
851	0
887	0
911	0
928	0
931	0
939	0
957	0
979	0

```
> sig_results_boot <- cbind(ovarian_cancer_methylation[diff_list_boot, ], diff_results$ordfit_t[diff_list_boot, ])
> print(sig_results_boot)
```

	IlmnID	Beta	exmdata2[, 2]	exmdata3[, 2]	exmdata4[, 2]
146	cg00134539	0.6110132	0.5332178	0.4599934	0.4678742
189	cg00176210	0.2875652	0.3916187	0.4427252	0.4472533
979	cg00945507	0.1343225	0.2385460	0.3474976	0.2890334
	exmdata5[, 2]	exmdata6[, 2]	exmdata7[, 2]	exmdata8[, 2]	
146	0.6719151	0.6313738	0.4792961	0.4542830	
189	0.3410608	0.3376593	0.4125211	0.3702489	
979	0.1184851	0.1665385	0.3071842	0.2662474	
	diff_results\$ordfit_t[diff_list_boot]				
146	5.636263				
189	-3.232921				
979	-4.968792				
	diff_results\$bootstrap_p[diff_list_boot]				
146	0				
189	0				
979	0				