

# Package ‘pairedGSEA’

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**Title** Paired DGE and DGS analysis for gene set enrichment analysis

**Version** 1.5.0

**Description** pairedGSEA makes it simple to run a paired Differential Gene Expression (DGE) and Differential Gene Splicing (DGS) analysis. The package allows you to store intermediate results for further investigation, if desired. pairedGSEA comes with a wrapper function for running an Over-Representation Analysis (ORA) and functionalities for plotting the results.

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**Imports** DESeq2, DEXSeq, limma, fgsea, sva, SummarizedExperiment, S4Vectors, BiocParallel, ggplot2, aggregation, stats, utils, methods

**Suggests** writexl, readxl, readr, rhdf5, msigdb, plotly, testthat (>= 3.0.0), knitr, rmarkdown, covr, BiocStyle

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**URL** <https://github.com/shdam/pairedGSEA>

**BugReports** <https://github.com/shdam/pairedGSEA/issues>

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example\_diff\_result    *Output of running paired\_diff on example\_se.*

---

## Description

This example result is used primarily to do package tests and for function man pages

## Usage

```
data("example_diff_result")
```

## Format

A ‘DataFrame’ with 954 rows and 7 columns.

## Value

A ‘DataFrame’.

---

example_gene_sets	<i>MSigDB gene sets from humans, category C5 with ensemble gene IDs</i>
-------------------	---

---

**Description**

This example gene set list is used primarily to do package tests and for function man pages.

**Usage**

```
data("example_gene_sets")
```

**Format**

A list of 77 human gene sets

**Value**

A list of gene sets

---

example_ora_results	<i>Output of running paired_ora on example_diff_result and gene sets extracted from MSigDB</i>
---------------------	--

---

**Description**

This example result is used primarily to do package tests and for function man pages.

**Usage**

```
data("example_ora_results")
```

**Format**

A 'DataFrame' with 559 rows and 18 columns.

**Value**

A 'DataFrame'

---

`example_se`*A small subset of the GEO:GSE61220 data set.*

---

**Description**

The subset is used in the vignettes and function man pages. The subset was created by extracting genes belonging to Telomere-related gene sets and randomly selecting 900 other genes from the original dataset.

**Usage**

```
data("example_se")
```

**Format**

A ‘SummarizedExperiment’

**assay** Count matrix with 5611 transcripts and 6 samples

**colData** The metadata associated with the count matrix

**Value**

A ‘SummarizedExperiment’

**Source**

<https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE61220>

---

`paired_diff`*Run paired DESeq2 and DEXSeq analyses*

---

**Description**

With `paired_diff` you can run a paired differential gene expression and splicing analysis. The function expects a counts matrix or a `SummarizedExperiment` or `DESeqDataSet` object as input. A preliminary prefiltering step is performed to remove genes with a summed count lower than the provided threshold. Likewise, genes with counts in only one sample are removed. This step is mostly to speed up differential analyses, as `DESeq2` will do a stricter filtering. Surrogate Variable Analysis is recommended to allow the analyses to take batch effects etc. into account. After the two differential analyses, the transcript-level p-values will be aggregated to gene-level to allow subsequent Gene-Set Enrichment Analysis. Transcript-level results can be extracted by setting `store_results = TRUE`.

**Usage**

```
paired_diff(
  object,
  group_col,
  sample_col,
  baseline,
  case,
  metadata = NULL,
  covariates = NULL,
  experiment_title = NULL,
  store_results = FALSE,
  run_sva = TRUE,
  use_limma = FALSE,
  prefilter = 10,
  test = "LRT",
  fit_type = "local",
  quiet = FALSE,
  parallel = FALSE,
  BPPARAM = BiocParallel::bpparam(),
  expression_only = FALSE,
  custom_design = FALSE,
  ...
)
```

**Arguments**

object	A data object of the types matrix, <a href="#">SummarizedExperiment</a> , or <a href="#">DESeqDataSet</a> . If a matrix is used, please also provide metadata.
group_col	The metadata column specifying the what group each sample is associated with
sample_col	The column in the metadata that specifies the sample IDs (should correspond to column names in object). Set to "rownames" if the rownames should be used.
baseline	Group value of baseline samples
case	Group value of case samples
metadata	(Default: NULL) A metadata file or data.frame object
covariates	Name of column(s) in the metadata that indicate(s) covariates. E.g., c("gender", "tissue_type")
experiment_title	Title of your experiment. Your results will be stored in <code>paste0("results/", experiment_title, "_pairedGSEA.RDS")</code> .
store_results	(Default: FALSE) A logical indicating if results should be stored in the folder "results/".
run_sva	(Default: TRUE) A logical stating whether SVA should be run.
use_limma	(Default: FALSE) A logical determining if limma+voom or DESeq2 + DEXSeq should be used for the analysis

<code>prefilter</code>	(Default: 10) The prefilter threshold, where rowSums lower than the prefilter threshold will be removed from the count matrix. Set to 0 or FALSE to prevent prefiltering
<code>test</code>	either "Wald" or "LRT", which will then use either Wald significance tests (defined by <code>nbinomWaldTest</code> ), or the likelihood ratio test on the difference in deviance between a full and reduced model formula (defined by <code>nbinomLRT</code> )
<code>fit_type</code>	(Default: "local") Either "parametric", "local", "mean", or "glmGamPoi" for the type of fitting of dispersions to the mean intensity.
<code>quiet</code>	(Default: FALSE) Whether to print messages
<code>parallel</code>	(Default: FALSE) If FALSE, no parallelization. If TRUE, parallel execution using <code>BiocParallel</code> , see next argument BPPARAM.
<code>BPPARAM</code>	(Default: <code>bpparam()</code> ) An optional parameter object passed internally to <code>bplapply</code> when <code>parallel = TRUE</code> . If not specified, the parameters last registered with <code>register</code> will be used.
<code>expression_only</code>	(Default: FALSE) A logical that indicates whether to only run <code>DESeq2</code> analysis. Not generally recommended. The setting was implemented to make the SVA impact analysis easier
<code>custom_design</code>	(Default: FALSE) A logical or formula. Can be used to apply a custom design formula for the analysis. Generally not recommended, as <code>pairedGSEA</code> will make its own design formula from the <code>group</code> and <code>covariate</code> columns
<code>...</code>	Additional parameters passed to <code>DESeq()</code>

**Value**

A DFrame of aggregated pvalues

**See Also**

Other paired: `paired_ora()`

**Examples**

```
# Run analysis on included example data
data("example_se")

diff_results <- paired_diff(
  object = example_se[1:15, ],
  group_col = "group_nr",
  sample_col = "id",
  baseline = 1,
  case = 2,
  experiment_title = "Example",
  store_results = FALSE
)
```

---

paired_ora	<i>Paired Over-Representation Analysis</i>
------------	--

---

## Description

paired\_ora uses [fora](#) to run the over-representation analysis. First the aggregated pvalues are adjusted using the Benjamini & Hochberg method. The analysis is run on all significant genes found by [DESeq2](#) and [DEXSeq](#) individually. I.e., two runs of [fora](#) are executed and subsequently joined into a single object. You can use [prepare\\_msigdb](#) to create a list of gene\_sets.

## Usage

```
paired_ora(  
  paired_diff_result,  
  gene_sets,  
  cutoff = 0.05,  
  min_size = 25,  
  experiment_title = NULL,  
  expression_only = FALSE,  
  quiet = FALSE  
)
```

## Arguments

paired_diff_result	The output of <a href="#">paired_diff</a>
gene_sets	List of gene sets to analyse
cutoff	(Default: 0.05) Adjusted p-value cutoff for selecting significant genes
min_size	(Default: 25) Minimal size of a gene set to test. All pathways below the threshold are excluded.
experiment_title	Title of your experiment. Your results will be stored in <code>paste0("results/", experiment_title, "_fora.RDS")</code> .
expression_only	(Default: FALSE) A logical that indicates whether to only run <a href="#">DESeq2</a> analysis. Not generally recommended.
quiet	(Default: FALSE) Whether to print messages

## Value

A data.table of merged ORA results

## See Also

Other paired: [paired\\_diff\(\)](#)

**Examples**

```
data("example_diff_result")
data("example_gene_sets")

ora <- paired_ora(
  example_diff_result,
  example_gene_sets)
```

---

plot\_ora

*Scatter plot of Over-Representation Analysis results*


---

**Description**

Scatter plot of Over-Representation Analysis results

**Usage**

```
plot_ora(
  ora,
  pattern = NULL,
  paired = TRUE,
  plotly = FALSE,
  cutoff = 0.05,
  lines = TRUE,
  colors = c("darkgray", "purple", "lightblue", "maroon")
)
```

**Arguments**

ora	Output of <a href="#">paired_ora</a>
pattern	Highlight pathways containing a specific regex pattern
paired	(Default: TRUE) New plotting mode for paired ora analysis
plotly	(Default: FALSE) Logical on whether to return plot as an interactive <a href="#">plotly</a> plot or a simple ggplot.
cutoff	(Default: 0.2) Adjusted p-value cutoff for pathways to include
lines	(Default: TRUE) Whether to show dashed lines
colors	(Default: c("darkgray", "purple", "navy")) Colors to use in plot. The colors are ordered as "Both", "DGS", and "DGE"

**Value**

A ggplot

**Note**

Suggested: `importFrom plotly ggplotly`

**Examples**

```
data(example_ora_results)

plot_ora(example_ora_results, pattern = "Telomer")
```

---

<code>prepare_msigdb</code>	<i>Load MSigDB and convert to names list of gene sets</i>
-----------------------------	---

---

**Description**

This function is wrapper around `msigdb()`. Please see their manual for details on its use. The function extracts the gene set name and a user-defined gene id type (Default: "ensembl\_gene"). Please make sure the gene IDs match those from your DE analysis. This function will format the gene sets such that they can be directly used with `paired_ora()`.

**Usage**

```
prepare_msigdb(
  gene_id_type = "ensembl_gene",
  species = "Homo sapiens",
  category = "C5",
  subcategory = NULL
)
```

**Arguments**

<code>gene_id_type</code>	(Default: "ensembl_gene") The gene ID type to extract. The IDs should match the gene IDs from your DE analysis.
<code>species</code>	Species name, such as Homo sapiens or Mus musculus.
<code>category</code>	MSigDB collection abbreviation, such as H or C1.
<code>subcategory</code>	MSigDB sub-collection abbreviation, such as CGP or BP.

**Value**

A list of gene sets

**Note**

Suggested: `importFrom msigdb msigdb`

**Examples**

```
gene_sets <- prepare_msigdb(species = "Homo sapiens")
```

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