

Package ‘spicyR’

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Type Package

Title Spatial analysis of in situ cytometry data

Version 1.14.3

Description The spicyR package provides a framework for performing inference on changes in spatial relationships between pairs of cell types for cell-resolution spatial omics technologies. spicyR consists of three primary steps: (i) summarizing the degree of spatial localization between pairs of cell types for each image; (ii) modelling the variability in localization summary statistics as a function of cell counts and (iii) testing for changes in spatial localizations associated with a response variable.

License GPL (>=2)

LazyData true

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BugReports <https://github.com/SydneyBioX/spicyR/issues>

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<https://github.com/SydneyBioX/spicyR>

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R topics documented:

| | |
|--|-----------|
| Accessors | 2 |
| as.data.frame.SegmentedCells | 4 |
| bind | 5 |
| colTest | 6 |
| convPairs | 6 |
| diabetesData | 7 |
| diabetesData_SCE | 8 |
| getPairwise | 8 |
| getProp | 10 |
| plot,SegmentedCells,ANY-method | 10 |
| SegmentedCells-class | 11 |
| show-SegmentedCells | 13 |
| signifPlot | 14 |
| spicyBoxPlot | 15 |
| SpicyResults-class | 16 |
| spicyTest | 18 |
| topPairs | 18 |
| Index | 20 |

Accessors

Accessors for SegmentedCells

Description

Methods to access various components of the ‘SegmentedCells’ object.

Usage

```
cellSummary(x, imageID = NULL, bind = TRUE)
```

```
cellSummary(x, imageID = NULL) <- value
```

```
cellMarks(x, imageID = NULL, bind = TRUE)
```

```
cellMarks(x, imageID = NULL) <- value
```

```
cellMorph(x, imageID = NULL, bind = TRUE)
```

```
cellMorph(x, imageID = NULL) <- value  
imagePheno(x, imageID = NULL, bind = TRUE, expand = FALSE)  
imagePheno(x, imageID = NULL) <- value  
imageID(x, imageID = NULL)  
cellID(x, imageID = NULL)  
cellID(x) <- value  
imageCellID(x, imageID = NULL)  
imageCellID(x) <- value  
cellType(x, imageID = NULL)  
cellType(x, imageID = NULL) <- value  
filterCells(x, select)  
cellAnnotation(x, variable, imageID = NULL)  
cellAnnotation(x, variable, imageID = NULL) <- value
```

Arguments

| | |
|----------|--|
| x | A 'SegmentedCells' object. |
| imageID | A vector of imageIDs to specifically extract. |
| bind | When false outputs a list of DataFrames split by imageID |
| expand | Used to expand the phenotype information from per image to per cell. |
| value | The relevant information used to replace. |
| select | A logical vector of the cells to be kept. |
| variable | A variable to add or retrieve from cellSummary. |

Value

DataFrame or a list of DataFrames

Descriptions

'cellSummary': Retrieves the DataFrame containing 'x' and 'y' coordinates of each cell as well as 'cellID', 'imageID' and 'cellType'. imageID can be used to select specific images and bind=FALSE outputs the information as a list split by imageID.

'cellMorph': Retrieves the DataFrame containing morphology information.

‘cellMarks‘: Retrieves the DataFrame containing intensity of gene or protein markers.

‘imagePheno‘: Retrieves the DataFrame containing the phenotype information for each image. Using `expand = TRUE` will produce a DataFrame with the number of rows equal to the number of cells.

Examples

```
### Something that resembles cellProfiler data

set.seed(51773)

n = 10

cells <- data.frame(row.names = seq_len(n))
cells$ObjectNumber <- seq_len(n)
cells$ImageNumber <- rep(1:2,c(n/2,n/2))
cells$AreaShape_Center_X <- runif(n)
cells$AreaShape_Center_Y <- runif(n)
cells$AreaShape_round <- rexp(n)
cells$AreaShape_diameter <- rexp(n, 2)
cells$Intensity_Mean_CD8 <- rexp(n, 10)
cells$Intensity_Mean_CD4 <- rexp(n, 10)

cellExp <- SegmentedCells(cells, cellProfiler = TRUE)

### Cluster cell types
intensities <- cellMarks(cellExp)
kM <- kmeans(intensities,2)
cellType(cellExp) <- paste('cluster',kM$cluster, sep = '')

cellSummary(cellExp, imageID = 1)
```

```
as.data.frame.SegmentedCells
      as.data.frame
```

Description

Function to coerce a `SegmentedCells` object to a data frame.

Usage

```
## S3 method for class 'SegmentedCells'
as.data.frame(x, ...)
```

Arguments

```
x          A SegmentedCells object.
...        Other arguments.
```

Value

A data.frame

```
## Generate toy data set.seed(51773) x <- round(c(runif(200),runif(200)+1,runif(200)+2,runif(200)+3,
runif(200)+3,runif(200)+2,runif(200)+1,runif(200)),4) y <- round(c(runif(200),runif(200)+1,runif(200)+2,runif(200)+3,
runif(200),runif(200)+1,runif(200)+2,runif(200)+3),4) cellType <- factor(paste('c',rep(rep(c(1:2),rep(200,2)),4),sep
= '')) imageID <- rep(c('s1', 's2'),c(800,800)) cells <- data.frame(x, y, cellType, imageID)

## Store data in SegmentedCells object cellExp <- SegmentedCells(cells, cellTypeString = 'cell-
Type')

## Generate LISA cellsDF <- as.data.frame(cellExp)

NULL
```

| | |
|------|---|
| bind | <i>Produces a dataframe showing L-function metric for each imageID entry.</i> |
|------|---|

Description

Produces a dataframe showing L-function metric for each imageID entry.

Usage

```
bind(results, pairName = NULL)
```

Arguments

| | |
|----------|---|
| results | Spicy test result obtained from spicy. |
| pairName | A string specifying the pairwise interaction of interest. If NULL, all pairwise interactions are shown. |

Value

A data.frame containing the colData related to the results.

Examples

```
data(spicyTest)
df <- bind(spicyTest)
```

| | |
|---------|---|
| colTest | <i>Perform a simple wilcoxon-rank-sum test or t-test on the columns of a data frame</i> |
|---------|---|

Description

Perform a simple wilcoxon-rank-sum test or t-test on the columns of a data frame

Usage

```
colTest(df, condition, type = NULL, feature = NULL, imageID = "imageID")
```

Arguments

| | |
|-----------|---|
| df | A data.frame or SingleCellExperiment, SpatialExperiment |
| condition | The condition of interest |
| type | The type of test, "wilcox", "ttest" or "survival". |
| feature | Can be used to calculate the proportions of this feature for each image |
| imageID | The imageID's if presenting a SingleCellExperiment |

Value

Proportions

Examples

```
# Test for an association with long-duration diabetes
# This is clearly ignoring the repeated measures...
data("diabetesData")
props <- getProp(diabetesData)
condition <- imagePheno(diabetesData)$stage
names(condition) <- imagePheno(diabetesData)$imageID
condition <- condition[condition %in% c("Long-duration", "Onset")]
test <- colTest(props[names(condition), ], condition)
```

| | |
|-----------|--|
| convPairs | <i>Converts colPairs object into an abundance matrix based on number of nearby interactions for every cell type.</i> |
|-----------|--|

Description

Converts colPairs object into an abundance matrix based on number of nearby interactions for every cell type.

Usage

```
convPairs(cells, colPair, cellType = "cellType", imageID = "imageID")
```

Arguments

| | |
|----------|--|
| cells | A SingleCellExperiment that contains objects in the colPairs slot. |
| colPair | The name of the object in the colPairs slot for which the dataframe is constructed from. |
| cellType | The cell type if using SingleCellExperiment. |
| imageID | The image ID if using SingleCellExperiment. |

Value

Matrix of abundances

Examples

```
data("diabetesData_SCE")

diabetesData_SPE <- SpatialExperiment::SpatialExperiment(diabetesData_SCE,
  colData = SingleCellExperiment::colData(diabetesData_SCE))
SpatialExperiment::spatialCoords(diabetesData_SPE) <- data.frame(
  SingleCellExperiment::colData(diabetesData_SPE)$x,
  SingleCellExperiment::colData(diabetesData_SPE)$y) |>
  as.matrix()

SpatialExperiment::spatialCoordsNames(diabetesData_SPE) <- c("x", "y")

diabetesData_SPE <- imcRtools::buildSpatialGraph(diabetesData_SPE,
  img_id = "imageID",
  type = "knn",
  k = 20,
  coords = c("x", "y"))

pairAbundances <- convPairs(diabetesData_SPE,
  colPair = "knn_interaction_graph")
```

diabetesData

Diabetes IMC data

Description

This is a subset of the Damond et al 2019 imaging mass cytometry dataset. The data contains cells in the pancreatic islets of individuals with early onset diabetes and healthy controls. The object contains single-cell data of 160 images from 8 subjects, with 20 images per subject.

Usage

diabetesData

Format

diabetesData a SegmentedCells object

diabetesData_SCE *Diabetes IMC data in SCE format.*

Description

This is a subset of the Damond et al 2019 imaging mass cytometry dataset. The data contains cells in the pancreatic islets of individuals with early onset diabetes and healthy controls. The object contains single-cell data of 160 images from 8 subjects, with 20 images per subject.

Usage

diabetesData_SCE

Format

diabetesData_SCE a SingleCellExperiment object

Details

Converted into a SingleCellExperiment format.

getPairwise *Get statistic from pairwise L curve of a single image.*

Description

Get statistic from pairwise L curve of a single image.

Usage

```
getPairwise(  
  cells,  
  from = NULL,  
  to = NULL,  
  window = "convex",  
  window.length = NULL,  
  Rs = c(20, 50, 100),  
  sigma = NULL,
```

```

minLambda = 0.05,
edgeCorrect = TRUE,
includeZeroCells = TRUE,
BPPARAM = BiocParallel::SerialParam(),
imageID = "imageID",
cellType = "cellType",
spatialCoords = c("x", "y")
)

```

Arguments

| | |
|------------------|--|
| cells | A SegmentedCells or data frame that contains at least the variables x and y, giving the location coordinates of each cell, and cellType. |
| from | The 'from' cellType for generating the L curve. |
| to | The 'to' cellType for generating the L curve. |
| window | Should the window around the regions be 'square', 'convex' or 'concave'. |
| window.length | A tuning parameter for controlling the level of concavity when estimating concave windows. |
| Rs | A vector of the radii that the measures of association should be calculated. |
| sigma | A numeric variable used for scaling when fitting inhomogeneous L-curves. |
| minLambda | Minimum value for density for scaling when fitting inhomogeneous L-curves. |
| edgeCorrect | A logical indicating whether to perform edge correction. |
| includeZeroCells | A logical indicating whether to include cells with zero counts in the pairwise association calculation. |
| BPPARAM | A BiocParallelParam object. |
| imageID | The imageID if using a SingleCellExperiment or SpatialExperiment. |
| cellType | The cellType if using a SingleCellExperiment or SpatialExperiment. |
| spatialCoords | The spatialCoords if using a SingleCellExperiment or SpatialExperiment. |

Value

Statistic from pairwise L curve of a single image.

Examples

```

data("diabetesData")
pairAssoc <- getPairwise(diabetesData[1, ])

```

| | |
|---------|--|
| getProp | <i>Get proportions from a SegmentedCells, SingleCellExperiment, SpatialExperiment or data.frame.</i> |
|---------|--|

Description

Get proportions from a SegmentedCells, SingleCellExperiment, SpatialExperiment or data.frame.

Usage

```
getProp(cells, feature = "cellType", imageID = "imageID")
```

Arguments

| | |
|---------|---|
| cells | SegmentedCells, SingleCellExperiment, SpatialExperiment or data.frame |
| feature | The feature of interest |
| imageID | The imageID's |

Value

Proportions

Examples

```
data("diabetesData")
prop <- getProp(diabetesData)
```

| | |
|----------------------------------|---|
| plot, SegmentedCells, ANY-method | <i>A basic plot for SegmentedCells object</i> |
|----------------------------------|---|

Description

This function generates a basic x-y plot of the location coordinates and cellType data.

Usage

```
## S4 method for signature 'SegmentedCells, ANY'
plot(x, imageID = NULL)
```

Arguments

| | |
|---------|-----------------------------------|
| x | A SegmentedCells object. |
| imageID | The image that should be plotted. |

Value

A ggplot object.

usage

```
'plot(x, imageID = NULL)'
```

Examples

```
### Something that resembles cellProfiler data

set.seed(51773)

n = 10

cells <- data.frame(row.names = seq_len(n))
cells$ObjectNumber <- seq_len(n)
cells$ImageNumber <- rep(1:2, c(n/2, n/2))
cells$AreaShape_Center_X <- runif(n)
cells$AreaShape_Center_Y <- runif(n)
cells$AreaShape_round <- rexp(n)
cells$AreaShape_diameter <- rexp(n, 2)
cells$Intensity_Mean_CD8 <- rexp(n, 10)
cells$Intensity_Mean_CD4 <- rexp(n, 10)

cellExp <- SegmentedCells(cells, cellProfiler = TRUE)

### Cluster cell types
markers <- cellMarks(cellExp)
kM <- kmeans(markers, 2)
cellType(cellExp) <- paste('cluster', kM$cluster, sep = '')

#plot(cellExp, imageID=1)
```

SegmentedCells-class *The SegmentedCells class*

Description

The SegmentedCells S4 class is for storing data from segmented imaging cytometry and spatial omics data. It extends DataFrame and defines methods that take advantage of DataFrame nesting to represent elements of cell-based experiments with spatial orientation that are commonly encountered. This object is able to store information on a cell's spatial location, cellType, morphology, intensity of gene/protein markers as well as image level phenotype information.

Usage

```
SegmentedCells(
  cellData,
  cellProfiler = FALSE,
  spatialCoords = c("x", "y"),
  cellTypeString = "cellType",
  intensityString = "intensity_",
  morphologyString = "morphology_",
  phenotypeString = "phenotype_",
  cellIDString = "cellID",
  cellAnnotations = NULL,
  imageCellIDString = "imageCellID",
  imageIDString = "imageID",
  verbose = TRUE
)
```

Arguments

| | |
|--------------------------------|---|
| <code>cellData</code> | A data frame that contains at least the columns <code>x</code> and <code>y</code> giving the location coordinates of each cell. |
| <code>cellProfiler</code> | A logical indicating that <code>cellData</code> is in a format similar to what <code>cellProfiler</code> outputs. |
| <code>spatialCoords</code> | The column names corresponding to spatial coordinates. eg. <code>x</code> , <code>y</code> , <code>z</code> ... |
| <code>cellTypeString</code> | The name of the column that contains cell type calls. |
| <code>intensityString</code> | A string which can be used to identify the columns which contain marker intensities. (This needs to be extended to take the column names themselves.) |
| <code>morphologyString</code> | A string which can be used to identify the columns which contains morphology information. |
| <code>phenotypeString</code> | A string which can be used to identify the columns which contains phenotype information. |
| <code>cellIDString</code> | The column name for <code>cellID</code> . |
| <code>cellAnnotations</code> | A vector of variables that provide additional annotation of a cell. |
| <code>imageCellIDString</code> | The column name for <code>imageCellID</code> . |
| <code>imageIDString</code> | The column name for <code>imageIDString</code> . |
| <code>verbose</code> | logical indicating whether to output messages. |

Value

A `SegmentedCells` object

Examples

```
### Something that resembles cellProfiler data

set.seed(51773)

n = 10

cells <- data.frame(row.names = seq_len(n))
cells$ObjectNumber <- seq_len(n)
cells$ImageNumber <- rep(seq_len(2),c(n/2,n/2))
cells$AreaShape_Center_X <- runif(n)
cells$AreaShape_Center_Y <- runif(n)
cells$AreaShape_round <- rexp(n)
cells$AreaShape_diameter <- rexp(n, 2)
cells$Intensity_Mean_CD8 <- rexp(n, 10)
cells$Intensity_Mean_CD4 <- rexp(n, 10)

cellExp <- SegmentedCells(cells, cellProfiler = TRUE)

### Cluster cell types
intensities <- cellMarks(cellExp)
kM <- kmeans(intensities,2)
cellType(cellExp) <- paste('cluster',kM$cluster, sep = '')
cellSummary(cellExp)
```

show-SegmentedCells *Show SegmentedCells*

Description

This outputs critical information about a SegmentedCells.

Arguments

object A SegmentedCells.

Value

Information of the number of images, cells, intensities, morphologies and phenotypes.

usage

‘show(object)’

Examples

```
### Something that resembles cellProfiler data

set.seed(51773)

n = 10

cells <- data.frame(row.names = seq_len(n))
cells$ObjectNumber <- seq_len(n)
cells$ImageNumber <- rep(1:2,c(n/2,n/2))
cells$AreaShape_Center_X <- runif(n)
cells$AreaShape_Center_Y <- runif(n)
cells$AreaShape_round <- rexp(n)
cells$AreaShape_diameter <- rexp(n, 2)
cells$Intensity_Mean_CD8 <- rexp(n, 10)
cells$Intensity_Mean_CD4 <- rexp(n, 10)

cellExp <- SegmentedCells(cells, cellProfiler = TRUE)

### Cluster cell types
markers <- cellMarks(cellExp)
kM <- kmeans(markers,2)
cellType(cellExp) <- paste('cluster',kM$cluster, sep = '')

cellExp
```

signifPlot

Plots result of signifPlot.

Description

Plots result of signifPlot.

Usage

```
signifPlot(
  results,
  fdr = FALSE,
  type = "bubble",
  breaks = NULL,
  comparisonGroup = NULL,
  colours = c("#4575B4", "white", "#D73027"),
  marksToPlot = NULL,
  cutoff = 0.05
)
```

Arguments

| | |
|-----------------|--|
| results | Data frame obtained from spicy. |
| fdr | TRUE if FDR correction is used. |
| type | Where to make a bubble plot or heatmap. |
| breaks | Vector of 3 numbers giving breaks used in pheatmap. The first number is the minimum, the second is the maximum, the third is the number of breaks. |
| comparisonGroup | A string specifying the name of the outcome group to compare with the base group. |
| colours | Vector of colours to use in pheatmap. |
| marksToPlot | Vector of marks to include in pheatmap. |
| cutoff | significance threshold for circles in bubble plot |

Value

a pheatmap object

Examples

```
data(spicyTest)
signifPlot(spicyTest, breaks = c(-3, 3, 0.5))
```

spicyBoxPlot

Plots boxplot for a specified cell-cell relationship

Description

Plots boxplot for a specified cell-cell relationship

Usage

```
spicyBoxPlot(results, from = NULL, to = NULL, rank = NULL)
```

Arguments

| | |
|---------|--|
| results | Data frame obtained from spicy. |
| from | Cell type which you would like to compare to the to cell type. |
| to | Cell type which you would like to compare to the from cell type. |
| rank | Ranking of cell type in terms of p-value, the smaller the p-value the higher the rank. |

Value

a ggplot2 boxplot

Examples

```
data(spicyTest)

spicyBoxPlot(spicyTest,
             rank = 1)
```

SpicyResults-class *Performs spatial tests on spatial cytometry data.*

Description

Performs spatial tests on spatial cytometry data.

Usage

```
spicy(
  cells,
  condition = NULL,
  subject = NULL,
  covariates = NULL,
  from = NULL,
  to = NULL,
  alternateResult = NULL,
  verbose = TRUE,
  weights = TRUE,
  weightsByPair = FALSE,
  weightFactor = 1,
  window = "convex",
  window.length = NULL,
  BPPARAM = BiocParallel::SerialParam(),
  sigma = NULL,
  Rs = NULL,
  minLambda = 0.05,
  edgeCorrect = TRUE,
  includeZeroCells = FALSE,
  imageID = "imageID",
  cellType = "cellType",
  spatialCoords = c("x", "y"),
  ...
)
```

Arguments

cells A SegmentedCells or data frame that contains at least the variables x and y, giving the location coordinates of each cell, and cellType.

| | |
|------------------|---|
| condition | Vector of conditions to be tested corresponding to each image if cells is a data frame. |
| subject | Vector of subject IDs corresponding to each image if cells is a data frame. |
| covariates | Vector of covariate names that should be included in the mixed effects model as fixed effects. |
| from | vector of cell types which you would like to compare to the to vector |
| to | vector of cell types which you would like to compare to the from vector |
| alternateResult | An pairwise association statistic between each combination of celltypes in each image. |
| verbose | logical indicating whether to output messages. |
| weights | logical indicating whether to include weights based on cell counts. |
| weightsByPair | logical indicating whether weights should be calculated for each cell type pair. |
| weightFactor | numeric that controls the convexity of the weight function. |
| window | Should the window around the regions be 'square', 'convex' or 'concave'. |
| window.length | A tuning parameter for controlling the level of concavity when estimating concave windows. |
| BPPARAM | A BiocParallelParam object. |
| sigma | A numeric variable used for scaling when fitting inhomogeneous L-curves. |
| Rs | A vector of radii that the measures of association should be calculated. |
| minLambda | Minimum value for density for scaling when fitting inhomogeneous L-curves. |
| edgeCorrect | A logical indicating whether to perform edge correction. |
| includeZeroCells | A logical indicating whether to include cells with zero counts in the pairwise association calculation. |
| imageID | The image ID if using SingleCellExperiment. |
| cellType | The cell type if using SingleCellExperiment. |
| spatialCoords | The spatial coordinates if using a SingleCellExperiment. |
| ... | Other options. |

Value

Data frame of p-values.

Examples

```
data("diabetesData")

# Test with random effect for patient on a pairwise combination of cell
# types.
spicy(diabetesData,
      condition = "stage", subject = "case",
      from = "Tc", to = "Th")
```

```

)

# Test all pairwise combinations of cell types without random effect of
# patient.
# spicityTest <- spicity(diabetesData, condition = "stage", subject = "case")

# Test all pairwise combination of cell types with random effect of patient.
# spicity(diabetesData, condition = "condition", subject = "subject")

```

spicityTest

Results from spicity for diabetesData

Description

Results from the call: `spicityTest <- spicity(diabetesData, condition = "condition", subject = "subject")`

Usage

```
spicityTest
```

Format

spicityTest a spicity object

topPairs

A table of the significant results from spicity tests

Description

A table of the significant results from spicity tests

Usage

```
topPairs(x, coef = NULL, n = 10, adj = "fdr", cutoff = NULL, figures = NULL)
```

Arguments

| | |
|---------|--|
| x | The output from spicity. |
| coef | Which coefficient to list. |
| n | Extract the top n most significant pairs. |
| adj | Which p-value adjustment method to use, argument for <code>p.adjust()</code> . |
| cutoff | A p-value threshold to extract significant pairs. |
| figures | Round to ‘figures’ significant figures. |

Value

A data.frame

Examples

```
data(spicyTest)
topPairs(spicyTest)
```

Index

* datasets

diabetesData, 7
diabetesData_SCE, 8
spicyTest, 18

Accessors, 2

as.data.frame.SegmentedCells, 4

bind, 5

cellAnnotation (Accessors), 2

cellAnnotation, SegmentedCells-method
(Accessors), 2

cellAnnotation<- (Accessors), 2

cellAnnotation<- , SegmentedCells-method
(Accessors), 2

cellID (Accessors), 2

cellID, SegmentedCells-method
(Accessors), 2

cellID<- (Accessors), 2

cellID<- , SegmentedCells-method
(Accessors), 2

cellMarks (Accessors), 2

cellMarks, SegmentedCells-method
(Accessors), 2

cellMarks<- (Accessors), 2

cellMarks<- , SegmentedCells-method
(Accessors), 2

cellMorph (Accessors), 2

cellMorph, SegmentedCells-method
(Accessors), 2

cellMorph<- (Accessors), 2

cellMorph<- , SegmentedCells-method
(Accessors), 2

cellSummary (Accessors), 2

cellSummary, SegmentedCells-method
(Accessors), 2

cellSummary<- (Accessors), 2

cellSummary<- , SegmentedCells-method
(Accessors), 2

cellType (Accessors), 2

cellType, SegmentedCells-method
(Accessors), 2

cellType<- (Accessors), 2

cellType<- , SegmentedCells-method
(Accessors), 2

colTest, 6

convPairs, 6

diabetesData, 7

diabetesData_SCE, 8

filterCells (Accessors), 2

filterCells, SegmentedCells-method
(Accessors), 2

getPairwise, 8

getProp, 10

imageCellID (Accessors), 2

imageCellID, SegmentedCells-method
(Accessors), 2

imageCellID<- (Accessors), 2

imageCellID<- , SegmentedCells-method
(Accessors), 2

imageID (Accessors), 2

imageID, SegmentedCells-method
(Accessors), 2

imagePheno (Accessors), 2

imagePheno, SegmentedCells-method
(Accessors), 2

imagePheno<- (Accessors), 2

imagePheno<- , SegmentedCells-method
(Accessors), 2

plot (plot, SegmentedCells, ANY-method),
10

plot, SegmentedCells
(plot, SegmentedCells, ANY-method),
10

plot, SegmentedCells, ANY-method, 10

SegmentedCells (SegmentedCells-class),
11

SegmentedCells, SegmentedCells-method
(SegmentedCells-class), 11

SegmentedCells-class, 11

show (show-SegmentedCells), 13

show-SegmentedCells, 13

signifPlot, 14

spicy (SpicyResults-class), 16

spicy, spicy-method
(SpicyResults-class), 16

spicyBoxPlot, 15

SpicyResults, list, ANY-method
(SpicyResults-class), 16

SpicyResults-class, 16

spicyTest, 18

topPairs, 18

topPairs, SpicyResults-method
(topPairs), 18