

Package ‘atena’

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Title Analysis of Transposable Elements

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Description Quantify expression of transposable elements (TEs) from RNA-seq data through different methods, including ERVmap, Tetranscripts and Telescope. A common interface is provided to use each of these methods, which consists of building a parameter object, calling the quantification function with this object and getting a SummarizedExperiment object as output container of the quantified expression profiles. The implementation allows one to quantify TEs and gene transcripts in an integrated manner.

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atena-package	<i>atena: analysis of transposable elements in R and Bioconductor</i>
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Description

The atena package provides a complete re-implementation in R of three existing methods for the quantification of transposable element (TE) expression in order to facilitate its integration into Bioconductor workflows for the analysis of RNA-seq data. The three methods are TEtranscripts (Jin et al. (2015)), ERVmap (Tokuyama et al. (2018)) and Telescope (Bendall et al.(2019)).

Details

The main functions are:

- `TEtranscriptsParam` - build parameter objects of the class `TEtranscriptsParam-class` for the TEtranscripts expression quantification method
- `ERVmapParam` - build parameter objects of the class `ERVmapParam-class` for the ERVmap expression quantification method
- `TelescopeParam` - build parameter objects of the class `TelescopeParam-class` for the Telescope expression quantification method
- `qtex` - call the TE expression quantification method using a previously built parameter object

For detailed information on usage, see the package vignette, by typing `vignette("atena")`.

All questions and bug reports should be posted to the Bioconductor Support Site:

<https://support.bioconductor.org>

The code of the development version of the package is available at the GitHub repository:

<https://github.com/functionalgenomics/atena>

References

Jin Y et al. TEtranscripts: a package for including transposable elements in differential expression analysis of RNA-seq datasets. *Bioinformatics*. 2015;31(22):3593-3599. DOI: <https://doi.org/10.1093/bioinformatics/btv422>

Tokuyama M et al. ERVmap analysis reveals genome-wide transcription of human endogenous retroviruses. *PNAS*. 2018;115(50):12565-12572. DOI: <https://doi.org/10.1073/pnas.1814589115>

Bendall et al. Telescope: characterization of the retrotranscriptome by accurate estimation of transposable element expression. *PLOS Comp. Biol.* 2019;15(9):e1006453. DOI: <https://doi.org/10.1371/journal.pcbi.1006453>

annotaTEs

Get RepeatMasker UCSC annotations

Description

The `annotaTEs()` function fetches RepeatMasker UCSC transposable element (TE) annotations using [AnnotationHub](#) and parses them.

Usage

```
annotaTEs(genome = "hg38", parsefun = rmskidentity)
```

Arguments

- | | |
|----------|---|
| genome | The genome version of the desired RepeatMasker annotations (e.g. "hg38"). |
| parsefun | A function to parse the annotations: <ul style="list-style-type: none"> • Function <code>rmskidentity</code> returns RepeatMasker annotations as present in AnnotationHub, without processing them. • Function <code>rmskbasicparser</code> parses annotations by removing low complexity regions, simple repeats, satellites, rRNA, scRNA, snRNA, srpRNA and tRNA. Also removes TEs with a strand different than "+" or "-". Modifies "repFamily" and "repClass" columns when a "?" is present or when they are defined as "Unknown" or "Other". Finally, assigns a unique id to each TE instance by adding the suffix "_dup" plus a number at the end of the "repName". • User-defined function. Input and output should be GRanges objects. |

Details

Given a specific genome version, the `annotaTEs()` function fetches RepeatMasker annotations from UCSC Genome Browser using the [AnnotationHub](#) package. Since RepeatMasker not only provides TE annotations but also low complexity DNA sequences and other types of repeats, a specific `parsefun` can be set to parse these annotations (e.g. `rmskbasicparser` or a user-defined function). If no parsing is required, `parsefun` can be set to `rmskidentity`.

Value

A [GRanges](#) object with transposable element annotations.

See Also

[AnnotationHub](#)

Examples

```
rmsk_gr <- annotaTEs(genome = "hg38", parsefun = rmskidentity)
```

atenaParam-class *atena parameter class*

Description

This is a class for storing parameters to quantify TE (and gene) expression using the `atena` method. It is a subclass of the `'QuantifyParam-class'`.

Build an object of the class `atenaParam`.

Usage

```
atenaParam(  
  bfl,  
  teFeatures,  
  aggregateby = character(0),  
  ovMode = "ovUnion",  
  geneFeatures = NA,  
  singleEnd = TRUE,  
  strandMode = 1L,  
  ignoreStrand = FALSE,  
  fragments = FALSE,  
  pi_prior = 0L,  
  theta_prior = 0L,  
  em_epsilon = 1e-07,  
  maxIter = 100L,  
  reassign_mode = "exclude",
```

```

    conf_prob = 0.9
)

## S4 method for signature 'atenaParam'
show(object)

```

Arguments

bf1	A BamFile or BamFileList object, or a character string vector of BAM file names.
teFeatures	A GRanges or GRangesList object. Elements in this object should have names, which are used as a grouping factor for genomic ranges forming a common locus. This grouping is performed previous to TE expression quantification, unlike the aggregation of quantifications performed when the aggregateby parameter is specified, which is performed after individual TE instances are quantified.
aggregateby	Character vector with column names from the annotation to be used to aggregate quantifications. By default, this is an empty vector, which means that the names of the input GRanges or GRangesList object given in the teFeatures parameter are used to aggregate quantifications.
ovMode	Character vector indicating the overlapping mode. Available options are: "ovUnion" (default) and "ovIntersectionStrict", which implement the corresponding methods from HTSeq (https://htseq.readthedocs.io/en/release_0.11.1/count.html). Ambiguous alignments (alignments overlapping > 1 feature) are not counted.
geneFeatures	A GRanges or GRangesList object with the gene annotated features to be quantified. Unique reads are first tallied with respect to these gene features whereas multi-mapping reads are preferentially assigned to TEs. Elements should have names indicating the gene name/id. In case that geneFeatures contains a metadata column named type, only the elements with type = exon are considered for the analysis. Then, exon counts are summarized to the gene level.
singleEnd	(Default TRUE) Logical value indicating if reads are single (TRUE) or paired-end (FALSE).
strandMode	(Default 1) Numeric vector which can take values 0, 1 or 2. The strand mode is a per-object switch on GAlignmentPairs objects that controls the behavior of the strand getter. See GAlignmentPairs class for further detail. If singleEnd = TRUE, then strandMode is ignored.
ignoreStrand	(Default FALSE) A logical which defines if the strand should be taken into consideration when computing the overlap between reads and annotated features. When ignoreStrand = FALSE, an aligned read is considered to be overlapping an annotated feature as long as they have a non-empty intersecting genomic range on the same strand, while when ignoreStrand = TRUE the strand is not considered.
fragments	(Default TRUE) A logical; applied to paired-end data only. When fragments=FALSE, the read-counting method only counts 'mated pairs' from opposite strands, while when fragments=TRUE (default), same-strand pairs, singletons, reads with unmapped pairs and other fragments are also counted. fragments=TRUE is equivalent to the original Telescope algorithm. For further details see summarizeOverlaps() .

pi_prior	(Default 0) A positive numeric object indicating the prior on pi. The same prior can be specified for all features setting pi_prior as a scalar, or each feature can have a specific prior by setting pi_prior as a vector with names() corresponding to all feature names. Setting a pi prior is equivalent to adding n unique reads.
theta_prior	(Default 0) A positive numeric object indicating the prior on Q. The same prior can be specified for all features setting theta_prior as a scalar, or each feature can have a specific prior by setting theta_prior as a vector with names() corresponding to all feature names. Equivalent to adding n non-unique reads.
em_epsilon	(Default 1e-7) A numeric scalar indicating the EM Algorithm Epsilon cutoff.
maxIter	A positive integer scalar storing the maximum number of iterations of the EM SQUAREM algorithm (Du and Varadhan, 2020). Default is 100 and this value is passed to the maxIter parameter of the squarem() function.
reassign_mode	(Default 'exclude') Character vector indicating reassignment mode after EM step. Available methods are 'exclude' (reads with more than one best assignment are excluded from the final counts), 'choose' (when reads have more than one best assignment, one of them is randomly chosen), 'average' (the read count is divided evenly among the best assignments) and 'conf' (only assignments that exceed a certain threshold -defined by conf_prob parameter- are accepted, then the read count is proportionally divided among the assignments above conf_prob).
conf_prob	(Default 0.9) Minimum probability for high confidence assignment.
object	A atenaParam object.

Details

This is the constructor function for objects of the class `atenaParam-class`. This type of object is the input to the function `qtex()` for quantifying expression of transposable elements, which will call the `atena` method with this type of object. The `atena` method uses a multiple '`__no_feature`' approach in which as many '`__no_feature`' features as different overlapping patterns of multimapping reads in the overlapping matrix are used to represent alignments mapping outside annotations.

Value

A [atenaParam](#) object.

Slots

`singleEnd` (Default TRUE) Logical value indicating if reads are single (TRUE) or paired-end (FALSE).

`strandMode` (Default 1) Numeric vector which can take values 0, 1 or 2. The strand mode is a per-object switch on [GAlignmentPairs](#) objects that controls the behavior of the strand getter. See [GAlignmentPairs](#) class for further detail. If `singleEnd = TRUE`, then `strandMode` is ignored.

`ignoreStrand` (Default FALSE) A logical which defines if the strand should be taken into consideration when computing the overlap between reads and annotated features. When `ignoreStrand = FALSE`, an aligned read is considered to be overlapping an annotated feature as long as they have a non-empty intersecting genomic range on the same strand, while when `ignoreStrand = TRUE` the strand is not considered.

- `fragments` (Default FALSE) A logical; applied to paired-end data only. When `fragments=FALSE` (default), the read-counting method only counts ‘mated pairs’ from opposite strands, while when `fragments=TRUE`, same-strand pairs, singletons, reads with unmapped pairs and other fragments are also counted. For further details see `summarizeOverlaps()`.
- `pi_prior` (Default 0) A positive numeric object indicating the prior on pi. The same prior can be specified for all features setting `pi_prior` as a scalar, or each feature can have a specific prior by setting `pi_prior` as a vector with `names()` corresponding to all feature names. Setting a pi prior is equivalent to adding n unique reads.
- `theta_prior` (Default 0) A positive numeric object indicating the prior on Q. The same prior can be specified for all features setting `theta_prior` as a scalar, or each feature can have a specific prior by setting `theta_prior` as a vector with `names()` corresponding to all feature names. Equivalent to adding n non-unique reads.
- `em_epsilon` (Default 1e-7) A numeric scalar indicating the EM Algorithm Epsilon cutoff.
- `maxIter` A positive integer scalar storing the maximum number of iterations of the EM SQUAREM algorithm (Du and Varadhan, 2020). Default is 100 and this value is passed to the `maxiter` parameter of the `squarem()` function.
- `reassign_mode` (Default ‘exclude’) Character vector indicating reassignment mode after EM step. Available methods are ‘exclude’ (reads with more than one best assignment are excluded from the final counts), ‘choose’ (when reads have more than one best assignment, one of them is randomly chosen), ‘average’ (the read count is divided evenly among the best assignments) and ‘conf’ (only assignments that exceed a certain threshold -defined by `conf_prob` parameter- are accepted, then the read count is proportionally divided among the assignments above `conf_prob`).
- `conf_prob` (Default 0.9) Minimum probability for high confidence assignment.

Examples

```
bamfiles <- list.files(system.file("extdata", package="atena"),
                      pattern="*.bam", full.names=TRUE)
TE_annot <- readRDS(file = system.file("extdata", "Top28TEs.rds",
                                     package="atena"))
gene_annot <- readRDS(file = system.file("extdata", "Top50genes.rds",
                                       package="atena"))
atpar <- atenaParam(bfl=bamfiles, teFeatures=TE_annot,
                  geneFeatures = gene_annot,
                  singleEnd = TRUE, ignoreStrand=TRUE)
atpar
```

ERVmapParam-class *ERVmap parameter class*

Description

This is a class for storing parameters provided to the ERVmap algorithm. It is a subclass of the ‘QuantifyParam-class’.

Build an object of the class ERVmapParam

Usage

```

ERVmapParam(
  bf1,
  teFeatures,
  aggregateby = character(0),
  ovMode = "ovUnion",
  geneFeatures = NA,
  singleEnd = TRUE,
  ignoreStrand = TRUE,
  strandMode = 1L,
  fragments = !singleEnd,
  maxMismatchRate = 0.02,
  suboptimalAlignmentTag = "auto",
  suboptimalAlignmentCutoff = 5,
  geneCountMode = "all"
)

## S4 method for signature 'ERVmapParam'
show(object)

```

Arguments

<code>bf1</code>	A BamFile or BamFileList object, or a character string vector of BAM file-names.
<code>teFeatures</code>	A GRanges or GRangesList object with the transposable element (TE) annotated features to be quantified. Elements in this object should have names, which are used as a grouping factor for genomic ranges forming a common locus, unless other metadata column names are specified in the <code>aggregateby</code> parameter.
<code>aggregateby</code>	Character vector with column names in the annotation to be used to aggregate quantifications. By default, this is an empty vector, which means that the names of the input GRanges or GRangesList object given in the <code>teFeatures</code> parameter are used to aggregate quantifications.
<code>ovMode</code>	Character vector indicating the overlapping mode. Available options are: "ovUnion" (default) and "ovIntersectionStrict", which implement the corresponding methods from HTSeq (https://htseq.readthedocs.io/en/release_0.11.1/count.html). Ambiguous alignments (alignments overlapping > 1 feature) are addressed as in the original ERVmap algorithm.
<code>geneFeatures</code>	A GRanges or GRangesList object with the gene annotated features to be quantified. Overlaps with unique reads are first tallied with respect to these gene features. Elements should have names indicating the gene name/id. In case that <code>geneFeatures</code> contains a metadata column named <code>type</code> , only the elements with <code>type = exon</code> are considered for the analysis. Then, exon counts are summarized to the gene level.
<code>singleEnd</code>	(Default TRUE) Logical value indicating if reads are single (TRUE) or paired-end (FALSE).
<code>ignoreStrand</code>	(Default TRUE) A logical which defines if the strand should be taken into consideration when computing the overlap between reads and annotated features.

When `ignore_strand = FALSE`, an aligned read is considered to be overlapping an annotated feature as long as they have a non-empty intersecting genomic range on the same strand, while when `ignoreStrand = TRUE` the strand is not considered.

<code>strandMode</code>	(Default 1) Numeric vector which can take values 0, 1 or 2. The strand mode is a per-object switch on GAlignmentPairs objects that controls the behavior of the strand getter. See GAlignmentPairs class for further detail. If <code>singleEnd = TRUE</code> , then <code>strandMode</code> is ignored.
<code>fragments</code>	(Default not <code>singleEnd</code>) A logical; applied to paired-end data only. When <code>fragments=TRUE</code> , the read-counting method in the original ERVmap algorithm is applied: each mate of a paired-end read is counted once and, therefore, two mates mapping to the same element result in adding up a count value of two. When <code>fragments=FALSE</code> , if the two mates of a paired-end read map to the same element, they are counted as a single hit and singletons, reads with unmapped pairs and other fragments, are not counted.
<code>maxMismatchRate</code>	(Default 0.02) Numeric value storing the maximum mismatch rate employed by the ERVmap algorithm to discard aligned reads whose rate of sum of hard and soft clipping or whose rate of the edit distance over the genome reference to the length of the read is above this threshold.
<code>suboptimalAlignmentTag</code>	(Default "auto") Character string storing the tag name in the BAM files that stores the suboptimal alignment score used in the third filter of ERVmap; see Tokuyama et al. (2018) . The default, <code>suboptimalAlignmentTag="auto"</code> , first extracts the name of the read mapper software from one or more BAM files. If BAM files were generated by BWA, the suboptimal alignment scores are obtained from a tag called XS. For other read mappers, the suboptimal alignment score is considered to be missing since, except from BWA, no other aligner provides a tag with suboptimal alignment scores. In this case, the available secondary alignments are used to implement an analogous approach to that of the third ERVmap filter. When <code>suboptimalAlignmentTag="none"</code> , it also performs the latter approach even when the tag XS is available. When this parameter is different from "auto" and "none", a tag with the given name is used to extract the suboptimal alignment score.
<code>suboptimalAlignmentCutoff</code>	(Default 5) Numeric value storing the cutoff above which the difference between the alignment score and the suboptimal alignment score is considered sufficiently large to retain the alignment. When this value is set to NA, the filtering step based on suboptimal alignment scores is skipped.
<code>geneCountMode</code>	(Default "all") Character string indicating if the ERVmap read filters applied to quantify TEs expression should also be applied when quantifying gene expression ("ervmap") or not ("all"), in which case all primary alignments mapping to genes are counted.
<code>object</code>	A ERVmapParam object.

Details

This is the constructor function for objects of the class `ERVmapParam-class`. This type of object is the input to the function `qtex()` for quantifying expression of transposable elements using the ERVmap method [Tokuyama et al. \(2018\)](#). The ERVmap algorithm processes reads following conservative filtering criteria to provide reliable raw count data for each TE.

Value

A `ERVmapParam` object.

Slots

- `readMapper` The name of the software used to align reads, obtained from the BAM file header.
- `singleEnd` (Default FALSE) Logical value indicating if reads are single (TRUE) or paired-end (FALSE).
- `strandMode` (Default 1) Numeric vector which can take values 0, 1 or 2. The strand mode is a per-object switch on `GAlignmentPairs` objects that controls the behavior of the strand getter. See `GAlignmentPairs` class for further detail. If `singleEnd = TRUE`, then `strandMode #'` is ignored.
- `ignoreStrand` (Default TRUE) A logical which defines if the strand should be taken into consideration when computing the overlap between reads and TEs in the annotations. When `ignore_strand = FALSE`, only those reads which overlap the TE and are on the same strand are counted. On the contrary, when `ignore_strand = TRUE`, any read overlapping an element in `teFeatures` is counted regardless of the strand.
- `fragments` (Default not `singleEnd`) A logical; applied to paired-end data only. When `fragments=TRUE` (default), the read-counting method in the original ERVmap algorithm will be applied: each mate of a paired-end read is counted once and, therefore, two mates mapping to the same element result in adding up a count value of two. When `fragments=FALSE`, if the two mates of a paired-end read map to the same element, they are counted as a single hit and singletons, reads with unmapped pairs and other fragments, are not counted.
- `maxMismatchRate` (Default 0.02) Numeric value storing the maximum mismatch rate employed by the ERVmap algorithm to discard aligned reads whose rate of sum of hard and soft clipping, or of the edit distance over the genome reference, to the length of the read is above this threshold.
- `suboptimalAlignmentTag` (Default "auto") Character string storing the tag name in the BAM files that stores the suboptimal alignment score used in the third filter of ERVmap; see [Tokuyama et al. \(2018\)](#). The default, `suboptimalAlignmentTag="auto"`, assumes that either the BAM files were generated by BWA and include a tag called `XS` that stores the suboptimal alignment score or, if the `XS` tag is not available, then it uses the available secondary alignments to implement an analogous approach to that of the third ERVmap filter. When `suboptimalAlignmentTag="none"`, it also performs the latter approach even when the tag `XS` is available. When this parameter is different from "auto" and "none", a tag with the given name is used to extract the suboptimal alignment score. The absence of that tag will prompt an error.
- `suboptimalAlignmentCutoff` (Default 5) Numeric value storing the cutoff above which the difference between the alignment score and the suboptimal alignment score is considered sufficiently large to retain the alignment. When this value is set to `NA`, then the filtering step based on suboptimal alignment scores is skipped.

geneCountMode (Default "all") Character string indicating if the ERVmap read filters applied to quantify TEs expression should also be applied when quantifying gene expression ("ervmap") or not ("all"), in which case all primary alignments mapping to genes are counted.

References

Tokuyama M et al. ERVmap analysis reveals genome-wide transcription of human endogenous retroviruses. PNAS. 2018;115(50):12565-12572. DOI: <https://doi.org/10.1073/pnas.1814589115>

Tokuyama M et al. ERVmap analysis reveals genome-wide transcription of human endogenous retroviruses. PNAS. 2018;115(50):12565-12572. DOI: <https://doi.org/10.1073/pnas.1814589115>

Examples

```
bamfiles <- list.files(system.file("extdata", package="atena"),
                      pattern="*.bam", full.names=TRUE)
TE_annot <- readRDS(file = system.file("extdata", "Top28TEs.rds",
                                       package="atena"))
empar <- ERVmapParam(bamfiles, teFeatures = TE_annot, singleEnd = TRUE,
                    ignoreStrand = TRUE, suboptimalAlignmentCutoff=NA)
empar
```

 ovUnion

Pre-defined overlapping mode functions

Description

The following functions control the way in which overlaps between aligned reads and annotated features are resolved when an aligned read overlaps more than one feature on the same locus:

Usage

```
ovUnion(reads, features, ignoreStrand, inter.feature = TRUE)
```

```
ovIntersectionStrict(reads, features, ignoreStrand, inter.feature = TRUE)
```

Arguments

reads	A GAlignments, GAlignmentList or a GAlignmentPairs object.
features	A GRanges object with annotated features.
ignoreStrand	(Default FALSE) A logical which defines if the strand should be taken into consideration when computing the overlap between reads and annotated features. When ignoreStrand = FALSE, an aligned read will be considered to be overlapping an annotated feature as long as they have a non-empty intersecting genomic ranges on the same strand, while when ignoreStrand = TRUE the strand will not be considered.

`inter.feature` When TRUE, ambiguous alignments (alignments overlapping > 1 features) are removed and not counted. When `inter.feature` is set to FALSE, these ambiguous overlaps are taken into account and addressed differently depending on the TE quantification.

Details

- `ovUnion()`: (default)
- `ovIntersectionStrict()`:
- User supplied: a function taking the same parameters as the previous three functions and returning a `Hits` object.

They take the following parameters:

These functions are given to the `mode` parameter of the `qtex()` function and are similar to the functions `Union()` and `IntersectionStrict()` from the `GenomicAlignments` package, with the difference that instead of returning counts of reads overlapping annotated features, they return the actual overlaps, because the counting is deferred to other algorithms that follow some specific strategy when a read maps to more than one feature. For this same reason, these functions lack the `inter.feature` argument found in the corresponding functions from the `GenomicAlignments` package.

Value

A `Hits` object; see the [Hits-class](#) manual page.

Examples

```
bamfiles <- list.files(system.file("extdata", package="atena"),
                      pattern="*.bam", full.names=TRUE)
TE_annot <- readRDS(file = system.file("extdata", "Top28TEs.rds",
                                      package="atena"))
tspar <- TelescopeParam(bfl=bamfiles, teFeatures=TE_annot,
                       singleEnd = TRUE, ignoreStrand=TRUE,
                       minOverlFract=0L)
tsSE <- qtex(tspar, mode=ovIntersectionStrict)
```

qtex,ERVmapParam-method

Quantify transposable element expression

Description

The `qtex()` method quantifies transposable element expression.

Usage

```
## S4 method for signature 'ERVmapParam'
qtex(
  x,
  phenodata = NULL,
  mode = ovUnion,
  yieldSize = 1000000L,
  verbose = 1,
  BPPARAM = SerialParam(progressbar = ifelse(verbose == 1, TRUE, FALSE))
)
```

```
## S4 method for signature 'TEtranscriptsParam'
qtex(
  x,
  phenodata = NULL,
  mode = ovUnion,
  yieldSize = 1000000L,
  BPPARAM = SerialParam(progressbar = TRUE)
)
```

```
## S4 method for signature 'TelescopeParam'
qtex(
  x,
  phenodata = NULL,
  mode = ovUnion,
  yieldSize = 1000000L,
  BPPARAM = SerialParam(progressbar = TRUE)
)
```

```
## S4 method for signature 'atenaParam'
qtex(
  x,
  phenodata = NULL,
  mode = ovUnion,
  yieldSize = 1000000L,
  BPPARAM = SerialParam(progressbar = TRUE)
)
```

Arguments

x	An <code>AtenaParam</code> object of one of the following subclasses: <ul style="list-style-type: none"> • A <code>ERVmapParam</code> object built using the constructor function <code>ERVmapParam()</code>. This object will trigger <code>qtex()</code> to use the algorithm by Tokuyama et al. (2018). • A <code>TelescopeParam</code> object built using the constructor function <code>TelescopeParam()</code>. This object will trigger <code>qtex()</code> to use the algorithm by Bendall et al. (2019).
phenodata	A <code>data.frame</code> or <code>DataFrame</code> object storing phenotypic data to include in the

	resulting SummarizedExperiment object. If phenodata is set, its row names will become the column names of the resulting <code>SummarizedExperiment</code> object.
mode	One of the pre-defined overlapping methods such as <code>ovUnion()</code> , <code>ovIntersectionStrict</code> or a user-supplied overlapping function. For a user-supplied overlapping function, the input parameters must match those of the pre-defined methods and the function must return a <code>Hits</code> object with subject hits matching the annotated features. This parameter is analogous to the mode parameter of the <code>summarizeOverlaps()</code> function from the <code>GenomicAlignments</code> package.
yieldSize	Field inherited from <code>BamFile</code> . The method for signature <code>ERVmapParam()</code> reads the BAM file by chunks. <code>yieldSize</code> represents the number of records (chunk size) to yield each time the file is read.
verbose	(Default 1). When <code>verbose > 1</code> , detailed information on the quantification steps is provided. Warnings are always present regardless of the value of <code>verbose</code> .
BPPARAM	An object of a <code>BiocParallelParam</code> subclass to configure the parallel execution of the code. By default, a <code>SerialParam</code> object is used, which does not use any parallelization, with the flag <code>progress=TRUE</code> to show progress through the calculations.

Details

Giving some `AtenaParam` object sub-class as input, the `qtex()` method quantifies the expression of transposable elements (TEs). The particular algorithm to perform the quantification will be selected depending on the specific sub-class of input `AtenaParam` object, see argument `x` above.

Value

A `SummarizedExperiment` object.

References

- Tokuyama M et al. ERVmap analysis reveals genome-wide transcription of human endogenous retroviruses. *PNAS*, 115(50):12565-12572, 2018. <https://doi.org/10.1073/pnas.1814589115>
- Bendall ML et al. Telescope: characterization of the retrotranscriptome by accurate estimation of transposable element expression. *PLOS Computational Biology*, 15:e1006453, 2019. <https://doi.org/10.1371/journal.pcbi.1006453>

See Also

[ERVmapParam TelescopeParam](#)

Examples

```
bamfiles <- list.files(system.file("extdata", package="atena"),
                      pattern="*.bam", full.names=TRUE)
TE_annot <- readRDS(file = system.file("extdata", "Top28TEs.rds",
                                      package="atena"))
gene_annot <- readRDS(file = system.file("extdata", "Top50genes.rds",
                                       package="atena"))
tspar <- TelescopeParam(bfl=bamfiles, teFeatures=TE_annot,
```

```

geneFeatures = gene_annot,
singleEnd = TRUE, ignoreStrand=TRUE)
tsSE <- qtex(tspar)

```

QuantifyParam-class *Atena parameter class*

Description

This is a virtual class from which other classes are derived for storing parameters provided to quantification methods of transposable elements from RNA-seq data.

Usage

```

## S4 method for signature 'QuantifyParam'
path(object)

## S4 method for signature 'QuantifyParam'
features(object)

```

Arguments

object A [QuantifyParam](#) object.

Value

path(): Filesystem paths to the BAM files in the input parameter object.

features(): The GenomicRanges or GenomicRangesList object with the features in the input parameter object.

Slots

bfl A [BamFileList](#) object.

features A [GRanges](#) object.

aggregateby Character vector with column names in the annotation to be used to aggregate quantifications.

ovMode Character vector indicating the overlapping mode. Available options are: "ovUnion" (default) and "ovIntersectionStrict", which implement the corresponding methods from HTSeq (https://htseq.readthedocs.io/en/release_0.11.1/count.html). In the Tetranscripts, ERVmap and Telescope methods ambiguous alignments (alignments overlapping > 1 feature) are addressed differently depending on the method. In the atena method, those overlaps are not counted.

See Also

[ERVmapParam-class](#) [TelescopeParam-class](#) [TetranscriptsParam-class](#) [atenaParam-class](#)

Examples

```
bamfiles <- list.files(system.file("extdata", package="atena"),
                       pattern="*.bam", full.names=TRUE)
TE_annot <- readRDS(file = system.file("extdata", "Top28TEs.rds",
                                       package="atena"))
ttpar <- TetranscriptsParam(bamfiles, teFeatures=TE_annot, singleEnd=TRUE,
                           ignoreStrand=TRUE, aggregateby = c("repName"))
path(ttpar)
```

rmskbasicparser

Parser of RepeatMasker annotations

Description

Parser of RepeatMasker annotations

Usage

```
rmskbasicparser(gr)
```

Arguments

`gr` A [GRanges](#) object with RepeatMasker annotations from [AnnotationHub](#)

Details

Parses annotations by removing low complexity regions, simple repeats, satellites, rRNA, scRNA, snRNA, srpRNA and tRNA. Also removes TEs with a strand different than "+" or "-". Modifies "repFamily" and "repClass" columns when a "?" is present or when they are defined as "Unknown" or "Other". Finally, assigns a unique id to each TE instance by adding the suffix "_dup" plus a number at the end of the "repName".

Value

A [GRanges](#) object.

Examples

```
rmsk_gr <- annotaTEs(genome = "dm6", parsefun = rmskbasicparser)
```

rmskidentity	<i>Identity function for parsefun</i>
--------------	---------------------------------------

Description

Identity function for parsefun

Usage

```
rmskidentity(gr)
```

Arguments

gr A [GRanges](#) object.

Details

Identity function: returns the [GRanges](#) object without any modification.

Value

A [GRanges](#) object.

Examples

```
rmsk_gr <- annotaTEs(genome = "dm6", parsefun = rmskidentity)
```

TelescopeParam-class	<i>Telescope parameter class</i>
----------------------	----------------------------------

Description

This is a class for storing parameters provided to the Telescope algorithm.

Build an object of the class TelescopeParam.

Usage

```
TelescopeParam(  
  bfl,  
  teFeatures,  
  aggregateby = character(0),  
  ovMode = "ovUnion",  
  geneFeatures = NA,  
  singleEnd = TRUE,  
  strandMode = 1L,
```

```

ignoreStrand = FALSE,
fragments = FALSE,
minOverlFract = 0.2,
pi_prior = 0L,
theta_prior = 0L,
em_epsilon = 1e-07,
maxIter = 100L,
reassign_mode = "exclude",
conf_prob = 0.9
)

## S4 method for signature 'TelescopeParam'
show(object)

```

Arguments

<code>bfl</code>	A BamFile or BamFileList object, or a character string vector of BAM file-names.
<code>teFeatures</code>	A GRanges or GRangesList object. Elements in this object should have names, which are used as a grouping factor for genomic ranges forming a common locus (equivalent to "locus" column in Telescope). This grouping is performed previous to TE expression quantification, unlike the aggregation of quantifications performed when the <code>aggregateby</code> parameter is specified, which is performed after individual TE instances are quantified.
<code>aggregateby</code>	Character vector with column names from the annotation to be used to aggregate quantifications. By default, this is an empty vector, which means that the names of the input GRanges or GRangesList object given in the <code>teFeatures</code> parameter are used to aggregate quantifications.
<code>ovMode</code>	Character vector indicating the overlapping mode. Available options are: "ovUnion" (default) and "ovIntersectionStrict", which implement the corresponding methods from HTSeq (https://htseq.readthedocs.io/en/release_0.11.1/count.html). Ambiguous alignments (alignments overlapping > 1 feature) are addressed as in the original Telescope method: the overlap with the longest overlapping length is kept.
<code>geneFeatures</code>	A GRanges or GRangesList object with the gene annotated features to be quantified. The TETRanscripts approach for gene expression quantification is used, in which overlaps with multi-mapping reads are preferentially assigned to TEs. Elements should have names indicating the gene name/id. In case that <code>geneFeatures</code> contains a metadata column named <code>type</code> , only the elements with <code>type = exon</code> are considered for the analysis. Then, exon counts are summarized to the gene level.
<code>singleEnd</code>	(Default TRUE) Logical value indicating if reads are single (TRUE) or paired-end (FALSE).
<code>strandMode</code>	(Default 1) Numeric vector which can take values 0, 1 or 2. The strand mode is a per-object switch on <code>GAlignmentPairs</code> objects that controls the behavior of the strand getter. See <code>GAlignmentPairs</code> class for further detail. If <code>singleEnd = TRUE</code> , then <code>strandMode</code> is ignored.

ignoreStrand	(Default FALSE) A logical which defines if the strand should be taken into consideration when computing the overlap between reads and annotated features. When ignoreStrand = FALSE, an aligned read is considered to be overlapping an annotated feature as long as they have a non-empty intersecting genomic range on the same strand, while when ignoreStrand = TRUE the strand is not considered.
fragments	(Default TRUE) A logical; applied to paired-end data only. When fragments=FALSE, the read-counting method only counts 'mated pairs' from opposite strands, while when fragments=TRUE (default), same-strand pairs, singletons, reads with unmapped pairs and other fragments are also counted. fragments=TRUE is equivalent to the original Telescope algorithm. For further details see summarizeOverlaps() .
minOverlFract	(Default 0.2) A numeric scalar. minOverlFract is multiplied by the read length and the resulting value is used to discard alignments for which the overlapping length (number of base pairs the alignment and the feature overlap) is lower. When no minimum overlap is required, set minOverlFract = 0.
pi_prior	(Default 0) A positive integer scalar indicating the prior on pi. This is equivalent to adding n unique reads.
theta_prior	(Default 0) A positive integer scalar storing the prior on Q. Equivalent to adding n non-unique reads.
em_epsilon	(Default 1e-7) A numeric scalar indicating the EM Algorithm Epsilon cutoff.
maxIter	A positive integer scalar storing the maximum number of iterations of the EM SQUAREM algorithm (Du and Varadhan, 2020). Default is 100 and this value is passed to the maxIter parameter of the squarem() function.
reassign_mode	(Default 'exclude') Character vector indicating reassignment mode after EM step. Available methods are 'exclude' (reads with more than one best assignment are excluded from the final counts), 'choose' (when reads have more than one best assignment, one of them is randomly chosen), 'average' (the read count is divided evenly among the best assignments) and 'conf' (only assignments that exceed a certain threshold -defined by conf_prob parameter- are accepted, then the read count is proportionally divided among the assignments above conf_prob).
conf_prob	(Default 0.9) Minimum probability for high confidence assignment.
object	A TelescopeParam object.

Details

This is the constructor function for objects of the class `TelescopeParam-class`. This type of object is the input to the function [qtex\(\)](#) for quantifying expression of transposable elements, which will call the Telescope algorithm [Bendall et al. \(2019\)](#) with this type of object.

Value

A [TelescopeParam](#) object.

Slots

- `singleEnd` (Default TRUE) Logical value indicating if reads are single (TRUE) or paired-end (FALSE).
- `strandMode` (Default 1) Numeric vector which can take values 0, 1 or 2. The strand mode is a per-object switch on `GAlignmentPairs` objects that controls the behavior of the strand getter. See `GAlignmentPairs` class for further detail. If `singleEnd = TRUE`, then `strandMode` is ignored.
- `ignoreStrand` (Default FALSE) A logical which defines if the strand should be taken into consideration when computing the overlap between reads and annotated features. When `ignoreStrand = FALSE`, an aligned read is considered to be overlapping an annotated feature as long as they have a non-empty intersecting genomic range on the same strand, while when `ignoreStrand = TRUE` the strand is not considered.
- `fragments` (Default FALSE) A logical; applied to paired-end data only. When `fragments=FALSE` (default), the read-counting method only counts ‘mated pairs’ from opposite strands, while when `fragments=TRUE`, same-strand pairs, singletons, reads with unmapped pairs and other fragments are also counted. For further details see `summarizeOverlaps()`.
- `minOverlFract` (Default 0.2) A numeric scalar. `minOverlFract` is multiplied by the read length and the resulting value is used to discard alignments for which the overlapping length (number of base pairs the alignment and the feature overlap) is lower. When no minimum overlap is required, set `minOverlFract = 0`.
- `pi_prior` (Default 0) A positive integer scalar indicating the prior on pi. This is equivalent to adding n unique reads.
- `theta_prior` (Default 0) A positive integer scalar storing the prior on Q. Equivalent to adding n non-unique reads.
- `em_epsilon` (Default 1e-7) A numeric scalar indicating the EM Algorithm Epsilon cutoff.
- `maxIter` A positive integer scalar storing the maximum number of iterations of the EM SQUAREM algorithm (Du and Varadhan, 2020). Default is 100 and this value is passed to the `maxiter` parameter of the `squarem()` function.
- `reassign_mode` (Default ‘exclude’) Character vector indicating reassignment mode after EM step. Available methods are ‘exclude’ (reads with more than one best assignment are excluded from the final counts), ‘choose’ (when reads have more than one best assignment, one of them is randomly chosen), ‘average’ (the read count is divided evenly among the best assignments) and ‘conf’ (only assignments that exceed a certain threshold -defined by `conf_prob` parameter- are accepted, then the read count is proportionally divided among the assignments above `conf_prob`).
- `conf_prob` (Default 0.9) Minimum probability for high confidence assignment.

References

- Bendall et al. Telescope: characterization of the retrotranscriptome by accurate estimation of transposable element expression. PLOS Comp. Biol. 2019;15(9):e1006453. DOI: <https://doi.org/10.1371/journal.pcbi.1006453>
- Bendall et al. Telescope: characterization of the retrotranscriptome by accurate estimation of transposable element expression. PLOS Comp. Biol. 2019;15(9):e1006453. DOI: <https://doi.org/10.1371/journal.pcbi.1006453>

Examples

```

bamfiles <- list.files(system.file("extdata", package="atena"),
                      pattern="*.bam", full.names=TRUE)
TE_annot <- readRDS(file = system.file("extdata", "Top28TEs.rds",
                                       package="atena"))
gene_annot <- readRDS(file = system.file("extdata", "Top50genes.rds",
                                       package="atena"))
tspar <- TelescopeParam(bfl=bamfiles, teFeatures=TE_annot,
                       geneFeatures = gene_annot,
                       singleEnd = TRUE, ignoreStrand=TRUE)

tspar

```

TEtranscriptsParam-class

TEtranscripts parameter class

Description

This is a class for storing parameters provided to the TEtranscripts algorithm. It is a subclass of the 'QuantifyParam-class'.

Build an object of the class TEtranscriptsParam

Usage

```

TEtranscriptsParam(
  bfl,
  teFeatures,
  aggregateby = character(0),
  ovMode = "ovUnion",
  geneFeatures = NA,
  singleEnd = TRUE,
  ignoreStrand = FALSE,
  strandMode = 1L,
  fragments = TRUE,
  tolerance = 1e-04,
  maxIter = 100L
)

## S4 method for signature 'TEtranscriptsParam'
show(object)

```

Arguments

bfl a character string vector of BAM file names.

teFeatures	A GRanges or GRangesList object with the TE annotated features to be quantified. Elements in this object should have names, which are used as a grouping factor for genomic ranges forming a common locus, unless other metadata column names are specified in the aggregateby parameter.
aggregateby	Character vector with column names from the annotation to be used to aggregate quantifications. By default, this is an empty vector, which means that the names of the input GRanges or GRangesList object given in the teFeatures parameter are used to aggregate quantifications.
ovMode	Character vector indicating the overlapping mode. Available options are: "ovUnion" (default) and "ovIntersectionStrict", which implement the corresponding methods from HTSeq (https://htseq.readthedocs.io/en/release_0.11.1/count.html). Ambiguous alignments (alignments overlapping > 1 feature) are addressed as in the original TEtranscripts method.
geneFeatures	A GRanges or GRangesList object with the gene annotated features to be quantified. Following the TEtranscripts algorithm, overlaps with unique reads are first tallied with respect to these gene features. Elements should have names indicating the gene name/id. In case that geneFeatures contains a metadata column named type, only the elements with type = exon are considered for the analysis. Then, exon counts are summarized to the gene level.
singleEnd	(Default TRUE) Logical value indicating if reads are single (TRUE) or paired-end (FALSE).
ignoreStrand	(Default FALSE) A logical which defines if the strand should be taken into consideration when computing the overlap between reads and annotated features. When ignoreStrand = FALSE, an aligned read is considered to be overlapping an annotated feature as long as they have a non-empty intersecting genomic range on the same strand, while when ignoreStrand = TRUE the strand is not considered.
strandMode	(Default 1) Numeric vector which can take values 0, 1 or 2. The strand mode is a per-object switch on GAlignmentPairs objects that controls the behavior of the strand getter. See GAlignmentPairs class for further detail. If singleEnd = TRUE, then strandMode is ignored.
fragments	(Default TRUE) A logical; applied to paired-end data only. When fragments=FALSE, the read-counting method only counts 'mated pairs' from opposite strands, while when fragments=TRUE (default), same-strand pairs, singletons, reads with unmapped pairs and other fragments are also counted. fragments=TRUE is equivalent to the behavior of the TEtranscripts algorithm. For further details see summarizeOverlaps() .
tolerance	A positive numeric scalar storing the minimum tolerance above which the SQUAREM algorithm (Du and Varadhan, 2020) keeps iterating. Default is 1e-4 and this value is passed to the tol parameter of the squarem() function.
maxIter	A positive integer scalar storing the maximum number of iterations of the SQUAREM algorithm (Du and Varadhan, 2020). Default is 100 and this value is passed to the maxIter parameter of the squarem() function.
object	A TEtranscriptsParam object.

Details

This is the constructor function for objects of the class `TEtranscriptsParam`-class. This type of object is the input to the function `qtex()` for quantifying expression of transposable elements using the TEtranscripts method [Jin et al. \(2015\)](#). The TEtranscripts algorithm quantifies TE expression by using an EM algorithm to optimally distribute ambiguously mapped reads.

Value

A `TEtranscriptsParam` object.

Slots

`singleEnd` (Default FALSE) Logical value indicating if reads are single (TRUE) or paired-end (FALSE).

`ignoreStrand` (Default FALSE) A logical which defines if the strand should be taken into consideration when computing the overlap between reads and annotated features. When `ignoreStrand = FALSE`, an aligned read will be considered to be overlapping an annotated feature as long as they have a non-empty intersecting genomic ranges on the same strand, while when `ignoreStrand = TRUE` the strand will not be considered.

`strandMode` (Default 1) Numeric vector which can take values 0, 1 or 2. The strand mode is a per-object switch on `GAlignmentPairs` objects that controls the behavior of the strand getter. See `GAlignmentPairs` class for further detail. If `singleEnd = TRUE`, then use either `strandMode = NULL` or do not specify the `strandMode` parameter.

`fragments` (Default TRUE) A logical; applied to paired-end data only. When `fragments=TRUE` (default), the read-counting method will also count reads without mates, while when `fragments=FALSE` those reads will not be counted. For further details see `summarizeOverlaps()`.

`tolerance` A positive numeric scalar storing the minimum tolerance above which the SQUAREM algorithm (Du and Varadhan, 2020) keeps iterating. Default is $1e-4$ and this value is passed to the `tol` parameter of the `squarem()` function.

`maxIter` A positive integer scalar storing the maximum number of iterations of the SQUAREM algorithm (Du and Varadhan, 2020). Default is 100 and this value is passed to the `maxiter` parameter of the `squarem()` function.

References

Jin Y et al. TEtranscripts: a package for including transposable elements in differential expression analysis of RNA-seq datasets. *Bioinformatics*. 2015;31(22):3593-3599. DOI: <https://doi.org/10.1093/bioinformatics/btv422>

Jin Y et al. TEtranscripts: a package for including transposable elements in differential expression analysis of RNA-seq datasets. *Bioinformatics*. 2015;31(22):3593-3599. DOI: <https://doi.org/10.1093/bioinformatics/btv422>

Examples

```
bamfiles <- list.files(system.file("extdata", package="atena"),
                      pattern="*.bam", full.names=TRUE)
TE_annot <- readRDS(file = system.file("extdata", "Top28TEs.rds",
```


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