

# Introduction to RBM package

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## 1 Overview

This document provides an introduction to the RBM package. The RBM package executes the resampling-based empirical Bayes approach using either permutation or bootstrap tests based on moderated t-statistics through the following steps.

- Firstly, the RBM package computes the moderated t-statistics based on the observed data set for each feature using the `lmFit` and `eBayes` function.
- Secondly, the original data are permuted or bootstrapped in a way that matches the null hypothesis to generate permuted or bootstrapped resamples, and the reference distribution is constructed using the resampled moderated t-statistics calculated from permutation or bootstrap resamples.
- Finally, the p-values from permutation or bootstrap tests are calculated based on the proportion of the permuted or bootstrapped moderated t-statistics that are as extreme as, or more extreme than, the observed moderated t-statistics.

Additional detailed information regarding resampling-based empirical Bayes approach can be found elsewhere (Li et al., 2013).

## 2 Getting started

The RBM package can be installed and loaded through the following R code.  
Install the RBM package with:

```
> if (!requireNamespace("BiocManager", quietly=TRUE))
+   install.packages("BiocManager")
> BiocManager::install("RBM")
```

Load the RBM package with:

```
> library(RBM)
```

## 3 RBM\_T and RBM\_F functions

There are two functions in the RBM package: `RBM_T` and `RBM_F`. Both functions require input data in the matrix format with rows denoting features and columns denoting samples. `RBM_T` is used for two-group comparisons such as study designs with a treatment group and a control group. `RBM_F` can be used for more complex study designs such as more than two groups or time-course studies. Both functions need a vector for group notation, i.e., "1" denotes the treatment group and "0" denotes the control group. For the `RBM_F` function, a contrast vector need to be provided by users to perform pairwise comparisons between groups. For example, if the design has three groups (0, 1, 2), the `aContrast` parameter will be a vector such as ("X1-X0", "X2-X1", "X2-X0") to denote all pairwise comparisons. Users just need to add an extra "X" before the group labels to do the contrasts.

- Examples using the `RBM_T` function: `normdata` simulates a standardized gene expression data and `unifdata` simulates a methylation microarray data. The  $p$ -values from the `RBM_T` function could be further adjusted using the `p.adjust` function in the `stats` package through the Benjamini-Hochberg method.

```
> library(RBM)
> normdata <- matrix(rnorm(1000*6, 0, 1),1000,6)
> mydesign <- c(0,0,0,1,1,1)
> myresult <- RBM_T(normdata,mydesign,100,0.05)
> summary(myresult)
```

	Length	Class	Mode
ordfit_t	1000	-none-	numeric
ordfit_pvalue	1000	-none-	numeric
ordfit_beta0	1000	-none-	numeric
ordfit_beta1	1000	-none-	numeric
permutation_p	1000	-none-	numeric
bootstrap_p	1000	-none-	numeric

```
> sum(myresult$permutation_p<=0.05)
```

```

[1] 16

> which(myresult$permutation_p<=0.05)

[1] 39 49 119 122 155 195 270 314 322 388 392 426 733 785 799 993

> sum(myresult$bootstrap_p<=0.05)

[1] 3

> which(myresult$bootstrap_p<=0.05)

[1] 270 289 483

> permutation_adj_p <- p.adjust(myresult$permutation_p, "BH")
> sum(permutation_adj_p<=0.05)

[1] 0

> bootstrap_adj_p <- p.adjust(myresult$bootstrap_p, "BH")
> sum(bootstrap_adj_p<=0.05)

[1] 0

> unifdata <- matrix(runif(1000*7,0.10, 0.95), 1000, 7)
> mydesign2 <- c(0,0,0, 1,1,1,1)
> myresult2 <- RBM_T(unifdata,mydesign2,100,0.05)
> sum(myresult2$permutatioin_p<=0.05)

[1] 0

> sum(myresult2$bootstrap_p<=0.05)

[1] 29

> which(myresult2$bootstrap_p<=0.05)

[1] 16 60 103 170 183 192 301 319 330 345 364 410 415 418 445 469 531 556 604
[20] 605 656 669 735 835 903 929 933 936 975

> bootstrap2_adj_p <- p.adjust(myresult2$bootstrap_p, "BH")
> sum(bootstrap2_adj_p<=0.05)

[1] 0

```

- Examples using the RBM\_F function: normdata\_F simulates a standardized gene expression data and unifdata\_F simulates a methylation microarray data. In both examples, we were interested in pairwise comparisons.

```

> normdata_F <- matrix(rnorm(1000*9,0,2), 1000, 9)
> mydesign_F <- c(0, 0, 0, 1, 1, 1, 2, 2, 2)
> aContrast <- c("X1-X0", "X2-X1", "X2-X0")
> myresult_F <- RBM_F(normdata_F, mydesign_F, aContrast, 100, 0.05)
> summary(myresult_F)

              Length Class  Mode
ordfit_t      3000   -none- numeric
ordfit_pvalue 3000   -none- numeric
ordfit_beta1   3000   -none- numeric
permutation_p 3000   -none- numeric
bootstrap_p    3000   -none- numeric

> sum(myresult_F$permutation_p[, 1]<=0.05)

[1] 54

> sum(myresult_F$permutation_p[, 2]<=0.05)

[1] 77

> sum(myresult_F$permutation_p[, 3]<=0.05)

[1] 54

> which(myresult_F$permutation_p[, 1]<=0.05)

[1] 17 36 48 92 102 113 119 130 147 149 173 188 193 215 216 233 236 258 289
[20] 306 343 351 361 363 377 397 406 415 478 541 561 591 596 598 609 690 710 719
[39] 726 728 736 783 789 807 826 833 861 875 878 925 931 932 960 987

> which(myresult_F$permutation_p[, 2]<=0.05)

[1] 17 22 35 36 40 48 92 102 113 119 128 130 139 141 143 147 149 165 173
[20] 184 188 193 210 216 236 241 245 258 273 281 289 306 308 318 335 343 351 361
[39] 363 373 377 405 406 415 450 478 500 541 545 561 591 598 609 636 647 651 662
[58] 690 719 726 736 778 783 789 807 826 831 833 861 875 878 922 925 931 932 960
[77] 987

> which(myresult_F$permutation_p[, 3]<=0.05)

[1] 17 36 48 56 102 113 119 128 130 141 147 149 173 188 193 215 216 236 245
[20] 258 289 306 335 343 351 361 363 377 406 415 478 519 541 588 591 598 609 655
[39] 690 726 736 748 789 807 826 833 861 875 878 909 925 931 932 987

> con1_adjp <- p.adjust(myresult_F$permutation_p[, 1], "BH")
> sum(con1_adjp<=0.05/3)

```

```

[1] 11

> con2_adj_p <- p.adjust(myresult_F$permutation_p[, 2], "BH")
> sum(con2_adj_p<=0.05/3)

[1] 18

> con3_adj_p <- p.adjust(myresult_F$permutation_p[, 3], "BH")
> sum(con3_adj_p<=0.05/3)

[1] 11

> which(con2_adj_p<=0.05/3)

[1] 40 48 113 119 289 351 361 415 541 598 636 736 789 807 826 875 878 987

> which(con3_adj_p<=0.05/3)

[1] 36 48 113 343 541 690 736 807 826 925 987

> unifdata_F <- matrix(runif(1000*18, 0.15, 0.98), 1000, 18)
> mydesign2_F <- c(rep(0, 6), rep(1, 6), rep(2, 6))
> aContrast <- c("X1-X0", "X2-X1", "X2-X0")
> myresult2_F <- RBM_F(unifdata_F, mydesign2_F, aContrast, 100, 0.05)
> summary(myresult2_F)

              Length Class  Mode
ordfit_t      3000    -none- numeric
ordfit_pvalue 3000    -none- numeric
ordfit_beta1  3000    -none- numeric
permutation_p 3000    -none- numeric
bootstrap_p   3000    -none- numeric

> sum(myresult2_F$bootstrap_p[, 1]<=0.05)

[1] 58

> sum(myresult2_F$bootstrap_p[, 2]<=0.05)

[1] 51

> sum(myresult2_F$bootstrap_p[, 3]<=0.05)

[1] 57

> which(myresult2_F$bootstrap_p[, 1]<=0.05)

```

```

[1] 43 48 49 56 75 83 93 97 98 107 115 127 132 134 135 137 138 147 180
[20] 197 199 223 275 298 301 313 329 384 400 427 461 471 473 494 535 563 714 736
[39] 756 780 787 806 813 876 880 889 911 912 926 928 929 946 952 963 974 986 993
[58] 999

> which(myresult2_F$bootstrap_p[, 2]<=0.05)

[1] 43 48 75 76 83 93 97 107 115 127 132 135 137 147 165 197 213 275 298
[20] 313 329 383 390 427 445 471 473 494 520 535 710 714 736 756 771 778 780 806
[39] 813 876 879 911 912 917 926 928 929 946 952 974 986

> which(myresult2_F$bootstrap_p[, 3]<=0.05)

[1] 43 48 75 83 93 97 107 134 135 137 138 147 149 165 180 183 197 199 223
[20] 275 298 301 313 329 364 383 384 390 401 427 445 473 494 520 535 710 714 736
[39] 780 787 806 813 876 880 889 911 912 917 926 928 929 946 952 963 974 986 999

> con21_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 1], "BH")
> sum(con21_adj_p<=0.05/3)

[1] 9

> con22_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 2], "BH")
> sum(con22_adj_p<=0.05/3)

[1] 7

> con23_adj_p <- p.adjust(myresult2_F$bootstrap_p[, 3], "BH")
> sum(con23_adj_p<=0.05/3)

[1] 10

```

## 4 Ovarian cancer methylation example using the RBM\_T function

Two-group comparisons are the most common contrast in biological and biomedical field. The ovarian cancer methylation example is used to illustrate the application of RBM\_T in identifying differentially methylated loci. The ovarian cancer methylation example is taken from the genome-wide DNA methylation profiling of United Kingdom Ovarian Cancer Population Study (UKOPS). This study used Illumina Infinium 27k Human DNA methylation Beadchip v1.2 to obtain DNA methylation profiles on over 27,000 CpGs in whole blood cells from 266 ovarian cancer women and 274 age-matched healthy controls. The data are downloaded from the NCBI GEO website with access number GSE19711. For illustration purpose, we chose the first 1000 loci in 8 randomly selected women with 4 ovarian cancer cases (pre-treatment) and 4 healthy controls. The following codes show the process of generating significant differential DNA methylation loci using the RBM\_T function and presenting the results for further validation and investigations.

```
> system.file("data", package = "RBM")
```

```
[1] "/private/tmp/RtmpPcJupN/Rinstfc3274b1ebb3/RBM/data"
```

```
> data(ovarian_cancer_methylation)
> summary(ovarian_cancer_methylation)
```

IlmnID	Beta	exmdata2[, 2]	exmdata3[, 2]
cg00000292: 1	Min. :0.01058	Min. :0.01187	Min. :0.009103
cg00002426: 1	1st Qu.:0.04111	1st Qu.:0.04407	1st Qu.:0.041543
cg00003994: 1	Median :0.08284	Median :0.09531	Median :0.087042
cg00005847: 1	Mean :0.27397	Mean :0.28872	Mean :0.283729
cg00006414: 1	3rd Qu.:0.52135	3rd Qu.:0.59032	3rd Qu.:0.558575
cg00007981: 1	Max. :0.97069	Max. :0.96937	Max. :0.970155
(Other) :994		NA's :4	

  

exmdata4[, 2]	exmdata5[, 2]	exmdata6[, 2]	exmdata7[, 2]
Min. :0.01019	Min. :0.01108	Min. :0.01937	Min. :0.01278
1st Qu.:0.04092	1st Qu.:0.04059	1st Qu.:0.05060	1st Qu.:0.04260
Median :0.09042	Median :0.08527	Median :0.09502	Median :0.09362
Mean :0.28508	Mean :0.28482	Mean :0.27348	Mean :0.27563
3rd Qu.:0.57502	3rd Qu.:0.57300	3rd Qu.:0.52099	3rd Qu.:0.52240
Max. :0.96658	Max. :0.97516	Max. :0.96681	Max. :0.95974
	NA's :1		

  

```
exmdata8[, 2]
Min. :0.01357
1st Qu.:0.04387
Median :0.09282
Mean :0.28679
3rd Qu.:0.57217
Max. :0.96268
```

```
> ovarian_cancer_data <- ovarian_cancer_methylation[, -1]
> label <- c(1, 1, 0, 0, 1, 1, 0, 0)
> diff_results <- RBM_T(aData=ovarian_cancer_data, vec_trt=label, repetition=100, alpha=0.05)
> summary(diff_results)
```

	Length	Class	Mode
ordfit_t	1000	-none-	numeric
ordfit_pvalue	1000	-none-	numeric
ordfit_beta0	1000	-none-	numeric
ordfit_beta1	1000	-none-	numeric
permutation_p	1000	-none-	numeric
bootstrap_p	1000	-none-	numeric

```
> sum(diff_results$ordfit_pvalue<=0.05)
```

```
[1] 45
```

```
> sum(diff_results$permutation_p<=0.05)
```

```

[1] 47

> sum(diff_results$bootstrap_p<=0.05)

[1] 42

> ordfit_adj_p <- p.adjust(diff_results$ordfit_pvalue, "BH")
> sum(ordfit_adj_p<=0.05)

[1] 0

> perm_adj_p <- p.adjust(diff_results$permutation_p, "BH")
> sum(perm_adj_p<=0.05)

[1] 5

> boot_adj_p <- p.adjust(diff_results$bootstrap_p, "BH")
> sum(boot_adj_p<=0.05)

[1] 0

> diff_list_perm <- which(perm_adj_p<=0.05)
> diff_list_boot <- which(boot_adj_p<=0.05)
> sig_results_perm <- cbind(ovarian_cancer_methylation[diff_list_perm, ], diff_results$ordfit_t)
> print(sig_results_perm)

```

	IlmnID	Beta	exmdata2[, 2]	exmdata3[, 2]	exmdata4[, 2]
5	cg00006414	0.07635468	0.07442468	0.15698040	0.08676092
245	cg00224508	0.04479948	0.04972043	0.04152814	0.04189373
346	cg00331237	0.05972383	NA	0.08204769	0.08345662
627	cg00612467	0.04777553	0.03783457	0.05380982	0.05582291
764	cg00730260	0.90471270	0.90542290	0.91002680	0.91258610

  

	exmdata5[, 2]	exmdata6[, 2]	exmdata7[, 2]	exmdata8[, 2]
5	0.07982556	0.08111396	0.08271889	0.08045977
245	0.04208405	0.05284988	0.03775905	0.03955271
346	0.05372019	0.06241126	0.06955040	0.09140985
627	0.04740551	0.05332965	0.05775211	0.05579710
764	0.90575890	0.88760470	0.90756300	0.90946790

  

	diff_results\$ordfit_t[diff_list_perm]
5	-1.389459
245	1.962457
346	-3.767916
627	-2.239498
764	-1.808081

  

	diff_results\$permutation_p[diff_list_perm]
5	0
245	0
346	0
627	0
764	0



```

> sig_results_boot <- cbind(ovarian_cancer_methylation[diff_list_boot, ], diff_results$ordfit_t
> print(sig_results_boot)

[1] IlmnID
[2] Beta
[3] exmdata2[, 2]
[4] exmdata3[, 2]
[5] exmdata4[, 2]
[6] exmdata5[, 2]
[7] exmdata6[, 2]
[8] exmdata7[, 2]
[9] exmdata8[, 2]
[10] diff_results$ordfit_t[diff_list_boot]
[11] diff_results$bootstrap_p[diff_list_boot]
<0 rows> (or 0-length row.names)

```