Comparing clustering algorithms

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October 18, 2011

1 Introduction

This document is the accumulation of a few somewhat unrelated threads of development:

- During the WTCCC study (phase 1, for which snpMatrix was written), some multi-panel colourised cluster-plot drawing code in R, mainly to verify the validity of cluster across cohorts. e.g. a 3x3 panel works very well for 7 common diseases plus 2 controls. This becomes snp.clust.plot function described later.
- When I noticed from looking at the cluster plots of some Illumina data I was working on that Illuminus is buggy and plainly wrong when calling the sex chromosomes, I wrote some simple low-level composite C/R code to catalogue the differences between Illuminus and GenTrain 1 to find out on which SNP they behaves most differently. This eventually becomes the snp.compare function described later.
- Almost 4 years after GenTrain 1 stabilised (in January 2006), Illumina debuted an alternative genotype-calling algorithm called "GenTrain 2" in December 2009 with the release of GenomeStudio 2009.2; "GenTrain 2" becomes the default with fresh installation of the next release, GenomeStudio 2010.1 (March 2010). SpringOnion added the ability to run "GenTrain 2" very soon afterwards.

Given that "GenTrain 2" is now the default, obviously Illumina staff must believe that it does a better job than "GenTrain 1" (besides its being marginally about 5% faster). The question is, in what way? We have a way of running the same raw data through two different algorithms, a way of cataloguing differences, and a way of displaying them side-by-side. So that's what the rest of the document does.

Similar techniques can be used for studying differences between any two algorithms.

2 Finding the largest differences

Using the 1550 samples of 99 SNPs of the Illuminus example set (the data is neither typical of BeadArray nor very good, but "real" data requires scrubbing and obfuscating). Reading the geno-types in:

```
> library(snpMatrix)
> load(system.file("data/Genotypes.GenTrain1.RData", package = "snpMatrix"))
> load(system.file("data/Genotypes.GenTrain2.RData", package = "snpMatrix"))
```

Reading the raw cluster data in:

```
> ab.signals <- read.wtccc.signals(system.file("extdata/example-new.txt",
+ package = "snpMatrix"), paste("rs", 1:99, sep = ""))
```

Reading /tmp/Rtmpsb5VvI/Rinst3b47450b/snpMatrix/extdata/example-new.txt ... Can take a while...

...Done

We'll just quickly run snpMatrix's SNP summary function to verify:

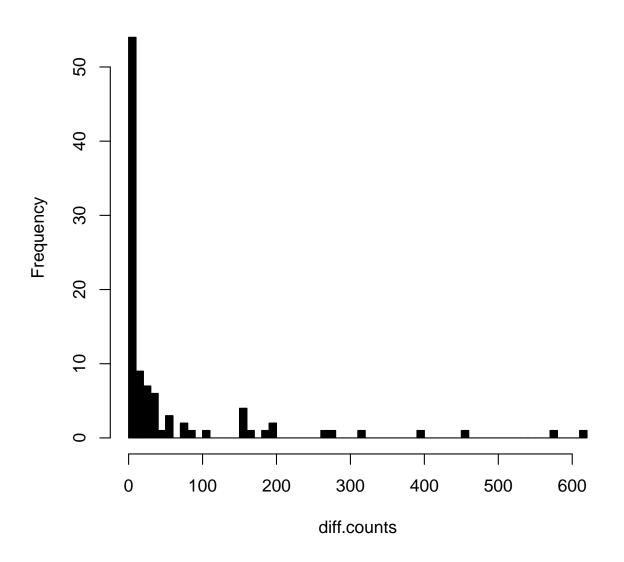
```
> col.summary(GenTrain2)[1:3, 2:7]
```

```
Call.rateMAFP.AAP.ABP.BBz.HWErs10.92571060.42288900.169574320.50662940.32379621.43643827rs20.97286820.13778220.019256310.23705180.7436919-0.08897442rs30.97739020.43721080.190350300.49372110.31592860.12694323
```

Calculate the largest difference count of the two genotype objects, and plot a histogram:

```
> result <- snp.compare(GenTrain1, GenTrain2)
> diff.counts <- result$count
> hist(diff.counts, breaks = 50, col = "black")
```

Histogram of diff.counts



More than half of the SNPs shows zero or close to zero differences between GenTrain 1 and GenTrain 2.

Lesson 1: For most SNPs (which are well-behaved and have 3 well-separated clusters), any half-decent calling algorithms do almost exactly the same thing.

Corollary 1: One must look at where two algorithms differ to see which one is "correct" more often.

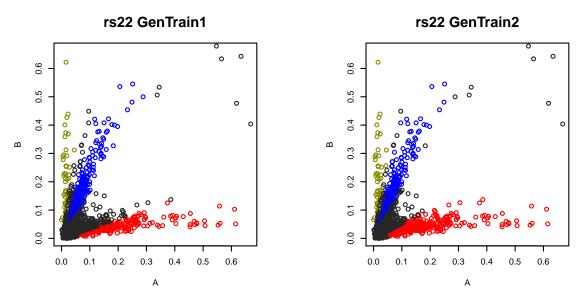
Corollary 2: If one can make sure that where two algorithms differ, one algorithm is more "correct" than the other, then that's what matters.

> dim(GenTrain1)

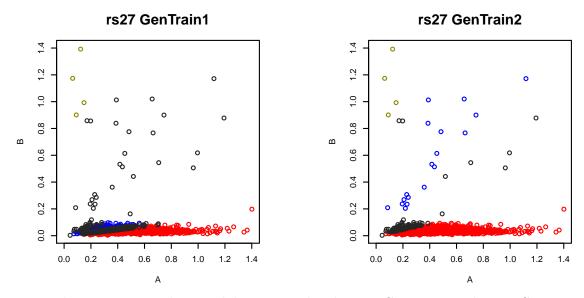
[1] 1550 99

```
> diff.counts[diff.counts > 1550 * 0.1]
     rs8 rs13 rs22 rs24 rs27 rs37 rs62 rs71 rs77 rs80 rs94
 rs5
      263
          160 320 166 576
                              187 617
                                        195 280
                                                  454 398
 192
> diff.counts[diff.counts > 1550 * 0.05]
     rs8 rs13 rs19 rs22 rs24 rs27 rs36 rs37 rs53 rs56 rs59 rs62 rs71 rs77 rs80
 rs5
 192
      263 160 103 320 166 576 151 187
                                              82 151 154 617 195
                                                                      280
                                                                          454
rs94
 398
> worst.snps <- names(diff.counts[diff.counts > 1550 * 0.2])
> worst.snps
[1] "rs22" "rs27" "rs62" "rs80" "rs94"
  There are 5 SNPs with more than 20\% difference. Let's plot these 5 SNPs:
```

```
> par(mfrow = c(1, 2), plt = c(0.25, 0.9, 0.15, 0.8), col.main = "black",
+ col.axis = "black", cex = 0.5, cex.main = 1.7)
> snp.clust.plot(ab.signals[["rs22"]], GenTrain1[, "rs22"], title = "rs22 GenTrain1")
> snp.clust.plot(ab.signals[["rs22"]], GenTrain2[, "rs22"], title = "rs22 GenTrain2")
```



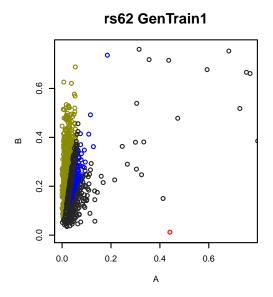
GenTrain 2 does a better job at calling more of the homozygous AA for rs22.

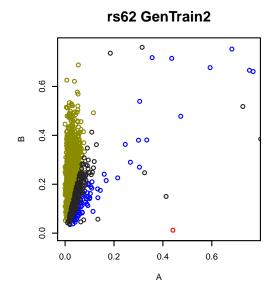


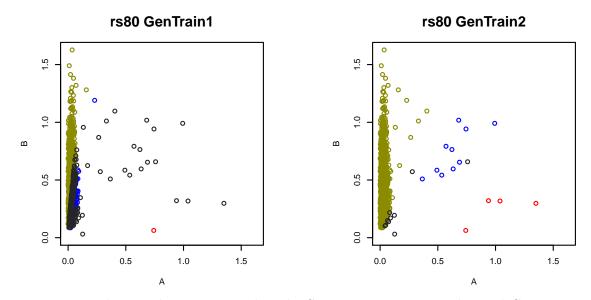
SNP rs27 is almost monomorphic — while one is inclined to say GenTrain 2 is better, GenTrain 2 also mis-behaves more than GenTrain 1 in calling some of the outliers (which were not called by GenTrain 1).

There are often wishful thinking along the line that higher call rates are always better. This bias often comes from the idea that since one already spent the effort for the samples to be processed, it is nicer to get a genotype than not getting one.

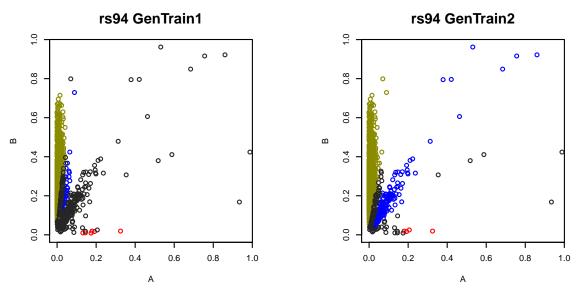
Lesson 2: Being optimistic (calling more) can be wrong.







SNP rs80 is somewhat similar to rs27 in that the SNP is near monomorphic and GenTrain 2 decides (somewhat arbitrarily) to classify outliers into minor clusters.



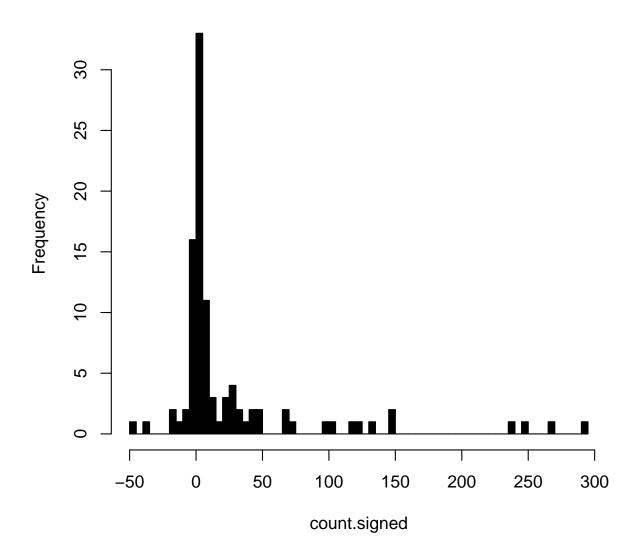
GenTrain 2 works very well on SNP rs94.

2.1 Does GenTrain 2 ever work less well than GenTrain1?

GenTrain 2 gives almost uniformly higher call rate across all SNPs:

```
> count.signed <- result$count.signed
> hist(count.signed, breaks = 50, col = "black")
```



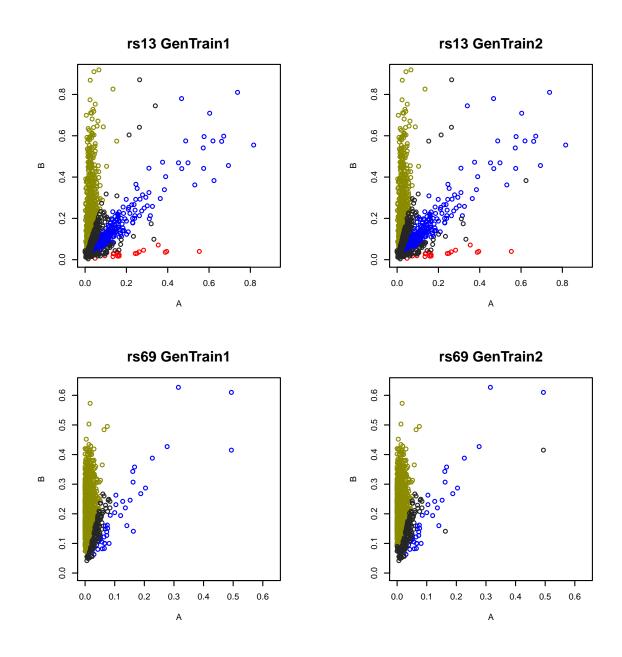


The two SNPs which give somewhat lower call rates under GenTrain 2 are:

```
> result$count.signed[result$count.signed < -30]</pre>
```

rs13 rs69 -36 -46

The differences are some what subtle when the cluster plots are examined:



3 Call rates and HWE

Let's have a look at the call rates, etc of those 5 worst SNPs again:

> col.summary(GenTrain1)[worst.snps, 2:7]

| Call.rate | MAF | P.AA | P.AB | P.BB | z.HWE |
|----------------|------------|--------------|------------|-------------|----------|
| rs22 0.4547804 | 0.31321023 | 0.4474431818 | 0.47869318 | 0.073863636 | 2.989549 |
| rs27 0.7571059 | 0.10878840 | 0.7858361775 | 0.21075085 | 0.003412969 | 2.973807 |
| rs62 0.7622739 | 0.13898305 | 0.0025423729 | 0.27288136 | 0.724576271 | 4.815062 |

rs80 0.8049096 0.06581059 0.0008025682 0.13001605 0.869181380 2.025882 rs94 0.7855297 0.04934211 0.0057565789 0.08717105 0.907072368 -2.469540

> col.summary(GenTrain2)[worst.snps, 2:7]

Call.rateMAFP.AAP.ABP.BBz.HWErs220.61498710.2310924370.5987394960.3403361340.060924370-1.305913rs270.93152450.0090152570.9847434120.0124826630.002773925-11.445045rs620.85852710.0312264860.0022573360.0579382990.939804364-1.545210rs800.99354010.0058517560.0026007800.0065019510.990897269-17.301685rs940.93863050.0715760500.0034411560.1362697870.8602890570.964802

In all cases, GenTrain 2 gives higher rates. We also see that GenTrain 2 allows HWE to deviate from normal a lot. Looking back at the cluster plots, we can see how that happens: GenTrain 2 calls more outliers when MAF is low and the SNP is near monomorphic. Or, conversely, nearmonomorphic SNPs (which could also be clustering errors, i.e. where two of the clusters are mis-clustered as one, and lone outliers are classified as the 3rd cluster) shows up as low call-rates under GenTrain 1 (which tends to put an arbitrary cut in the middle), while GenTrain 2 draws in outliers and the SNP shows up with strong deviation from HWE.

Under the conventional filtering rules: a sufficiently high call rate (say 95%) and good HWE (|HWE| < 5), all 5 are rather poor under GenTrain 1, but rs94 becomes almost acceptable (as is shown in the cluster plot) under GenTrain 2.

In any case, as a final remark: GenTrain 2 is the new default — it doesn't necessarily mean it is any good, but 3rd-party alternative needs to compared well with it under "fair" conditions where both are run optimally.