

hpar: The Human Protein Atlas in R

Laurent Gatto

lg390@cam.ac.uk

Cambridge Center for Proteomics
University of Cambridge

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Abstract

The Human Protein Atlas (HPA) is a systematic study of the human proteome using antibody-based proteomics. Multiple tissues and cell lines are systematically assayed using affinity-purified antibodies and confocal microscopy. The **hpar** package is an R interface to the HPA project. It distributes three data sets, provides functionality to query these and to access detailed information pages, including confocal microscopy images available on the HPA web page.

Keywords: infrastructure, bioinformatics, proteomics, microscopy

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1 Introduction

1.1 The HPA project

From the Human Protein Atlas¹ (Uhlén et al., 2005; Uhlen et al., 2010) site:

The Swedish Human Protein Atlas project, funded by the Knut and Alice Wallenberg Foundation, has been set up to allow for a systematic exploration of the human proteome using Antibody-Based Proteomics. This is accomplished by combining high-throughput generation of affinity-purified antibodies with protein profiling in a multitude of tissues and cells assembled in tissue microarrays. Confocal microscopy analysis using human cell lines is performed for more detailed protein localisation. The program hosts the Human Protein Atlas portal with expression profiles of human proteins in tissues and cells.

The `hpar` package provides functionality to use HPA data from the R interface. It also distributes three data sets available from the HPA site.

Normal tissue data Expression profiles for proteins in human tissues based on immunohistochemistry using tissue micro arrays. The `dataframe` includes Ensembl gene identifier ("Gene"), tissue name ("Tissue"), annotated cell type ("Cell.type"), expression value ("Level"), the type of annotation (annotated protein expression (APE), based on more than one antibody, or staining, based on one antibody only) ("Expression.type"), and the reliability or validation of the expression value ("Reliability").

Subcellular location data Subcellular localisation of proteins based on immunofluorescently stained cells. The `dataframe` includes Ensembl gene identifier ("Gene"), main subcellular location of the protein ("Main.location"), other locations ("Other.location"), the type of annotation (annotated protein expression (APE), based on more than one antibody, or staining, based on one antibody only) ("Expression.type"), and the reliability or validation of the expression value ("Reliability").

RNA data RNA levels in three different cell lines, based on RNA-seq. The `dataframe` includes Ensembl gene identifier ("Gene"), analysed cell line ("Cell.line"), number of reads per kilobase gene model and million reads ("RPKM"), and abundance class ("Abundance").

1.2 HPA data usage policy

The use of data and images from the HPA in publications and presentations is permitted provided that the following conditions are met:

- The publication and/or presentation are solely for informational and non-commercial purposes.
- The source of the data and/or image is referred to the HPA site (www.proteinatlas.org) and/or one or more of our publications are cited.

1.3 Installation

`hpar` is available through the Bioconductor project. Details about the package and the installation procedure can be found on its page². To install using the dedicated Bioconductor infrastructure, run :

¹<http://www.proteinatlas.org/>

²<http://bioconductor.org/packages/devel/bioc/html/hpar.html>

```

> source("http://bioconductor.org/biocLite.R")
> ## or, if you have already used the above before
> library("BiocInstaller") ## and to install the package
> biocLite("hpar")

```

After installation, hpar will have to be explicitly loaded with

```

> library("hpar")

This is hpar 1.0.1. For more information,
please type '?hpar' or 'vignette('hpar')'.

```

so that all the package's functionality and data is available to the user.

2 The hpar package

2.1 Data sets

The three data sets, named `hpaNormalTissue`, `hpaSubcellularLoc` and `hpaRna` in the package can be loaded with the `data` function, as illustrated below for `hpaNormalTissue` below. Each data set is a `dataframe` and can be easily manipulated using standard R functionality. The code chunk below illustrates some of its properties.

```

> data(hpaNormalTissue)
> dim(hpaNormalTissue)

[1] 1101631      6

> names(hpaNormalTissue)

[1] "Gene"          "Tissue"          "Cell.type"
[4] "Level"         "Expression.type" "Reliability"

> ## Number of genes
> length(unique(hpaNormalTissue$Gene))

[1] 14079

> ## Number of cell types
> length(unique(hpaNormalTissue$Cell.type))

[1] 43

> head(levels(hpaNormalTissue$Cell.type))

[1] "adipocytes"          "bile duct cells"
[3] "cells in endometrial stroma" "cells in glomeruli"
[5] "cells in granular layer" "cells in molecular layer"

> ## Number of tissues
> length(unique(hpaNormalTissue$Tissue))

[1] 48

> head(levels(hpaNormalTissue$Tissue))

```

```
[1] "adrenal gland" "appendix"      "bone marrow"   "breast"
[5] "bronchus"     "cerebellum"

> table(hpaNormalTissue$Expression.type)

      APE Staining
254533    847098
```

2.2 HPA interface

The package provides a interface to the HPA data. The `getHpa` allows to query the data sets described in section 2.1. It takes three arguments, `id`, `hpadata` and `type`, that control the query, what data set to interrogate and how to report results respectively. The HPA data uses Ensembl gene identifiers and `id` must be a valid identifier. `hpadata` must be one of "NormalTissue", "Rna" or "SubcellularLoc". `type` can be `data` or `details`. The former is the default and returns a `dataframe` containing the information relevant to `id`. It is also possible to obtain detailed information, (including cell images) as web pages, directly from the HPA web page, using `details`.

We will illustrate this functionality with using the E74-like factor 3 gene (ENSG00000163435) as example.

```
> id <- "ENSG00000163435"
> head(getHpa(id, hpadata = "NormalTissue"))
```

	Gene	Tissue	Cell.type	Level
670255	ENSG00000163435	adrenal gland	glandular cells	None
670256	ENSG00000163435	appendix	glandular cells	High
670257	ENSG00000163435	appendix	lymphoid tissue	None
670258	ENSG00000163435	bone marrow	hematopoietic cells	None
670259	ENSG00000163435	breast	adipocytes	None
670260	ENSG00000163435	breast	glandular cells	None

```
      Expression.type Reliability
670255      APE      High
670256      APE      High
670257      APE      High
670258      APE      High
670259      APE      High
670260      APE      High

> getHpa(id, hpadata = "SubcellularLoc")
```

	Gene	Main.location
7025	ENSG00000163435	Nucleus but not nucleoli;Cytoplasm

```
      Other.location Expression.type Reliability
7025      APE      High

> head(getHpa(id, hpadata = "Rna"))
```

	Gene	sample	Value	Unit	Abundance
32722	ENSG00000163435	A-431	54.1	FPKM	high
32723	ENSG00000163435	U-2 OS	0.1	FPKM	low
32724	ENSG00000163435	U-251 MG	0.5	FPKM	low

If we ask for `detail`, a browser page pointing to the relevant page is open (see figure 1)

```
> getHpa(id, type = "details")
```

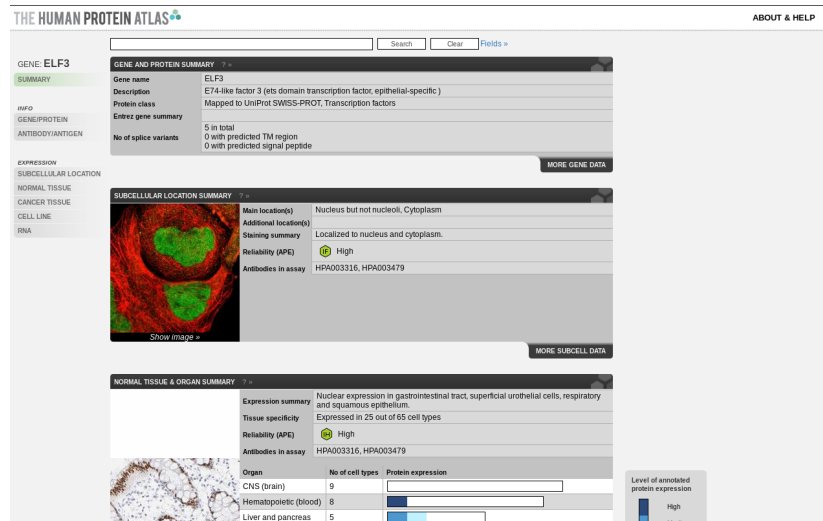


Figure 1: The HPA web page for the E74-like factor 3 gene (ENSG00000163435) gene.

If a user is interested specifically in one data set, it is possible to set `hpardata` globally and omit it in `getHpa`. This is done by setting the `hpar` options `hpardata` with the `setHparOptions` function. The current default data set can be tested with `getHparOptions`.

```
> getHparOptions()

$hpar
$hpar$hpardata
[1] "NormalTissue"

> setHparOptions(hpardata = "SubcellularLoc")
> getHparOptions()

$hpar
$hpar$hpardata
[1] "SubcellularLoc"

> getHpa(id)

      Gene                               Main.location
7025 ENSG00000163435 Nucleus but not nucleoli;Cytoplasm
      Other.location Expression.type Reliability
7025                               APE           High
```

2.3 HPA release information

Information about the HPA release used to build the installed `hpar` package can be accessed with `getHpaVersion`, `getHpaDate` and `getHpaEnsembl`. Full release details can be found on the HPA release history³ page.

³<http://www.proteinatlas.org/about/releases>

```

> getHpaVersion()
[1] "Protein Atlas version 10.0"

> getHpaDate()
[1] "2012.09.12"

> getHpaEnsembl()
[1] "67.37"

```

3 A small use case

Let's compare the subcellular localisation annotation obtained from the HPA subcellular location data set and the information available in the Bioconductor annotation packages. The HPA query shown below indicates that the HECW1 (ENSG00000002746) gene main locations are nucleus (but not nucleoli) and cytoplasm.

```

> id <- "ENSG00000002746"
> getHpa(id, "SubcellularLoc")

```

	Gene	Main.location	Other.location
24	ENSG00000002746	Nucleus but not nucleoli;	Cytoplasm
	Expression.type	Reliability	
24	APE	High	

Below, we first extract all cellular component GO terms available for ENSG00000002746 from the org.Hs.eg.db human annotation and then retrieve their term definitions using the GO.db database, indicating concordant results. The IDA evidence code indicates that this information is inferred from direct assay.

```

> library(org.Hs.eg.db)
> library(GO.db)
> ans <- select(org.Hs.eg.db, keys = id, cols = c("ENSEMBL", "GO", "ONTOLOGY"),
+             keytype = "ENSEMBL")

```

Warning: 'select' resulted in 1:many mapping between keys and return rows

```

> ans <- ans[ans$ONTOLOGY == "CC", ]
> ans

```

	ENSEMBL	GO	EVIDENCE	ONTOLOGY
2	ENSG00000002746	GO:0005634	IDA	CC
3	ENSG00000002746	GO:0005737	IDA	CC

```

> sapply(as.list(GOTERM[ans$GO]), slot, "Term")

```

GO:0005634	GO:0005737
"nucleus"	"cytoplasm"

Session information

- R version 2.15.2 (2012-10-26), i386-apple-darwin9.8.0
- Locale: C/en_US.US-ASCII/en_US.US-ASCII/C/en_US.US-ASCII/en_US.US-ASCII
- Base packages: base, datasets, grDevices, graphics, methods, stats, utils
- Other packages: AnnotationDbi 1.20.3, Biobase 2.18.0, BiocGenerics 0.4.0, DBI 0.2-5, GO.db 2.8.0, RSQLite 0.11.2, codetools 0.2-8, hpar 1.0.1, knitr 1.0, org.Hs.eg.db 2.8.0
- Loaded via a namespace (and not attached): IRanges 1.16.4, digest 0.6.0, evaluate 0.4.3, formatR 0.7, parallel 2.15.2, stats4 2.15.2, stringr 0.6.2, tools 2.15.2

References

Mathias Uhlén, Erik Björling, Charlotta Agaton, Cristina Al-Khalili A. Szigartyo, Bahram Amini, Elisabet Andersen, Ann-Catrin C. Andersson, Pia Angelidou, Anna Asplund, Caroline Asplund, Lisa Berglund, Kristina Bergström, Harry Brumer, Dijana Cerjan, Marica Ekström, Adila Elobeid, Cecilia Eriksson, Linn Fagerberg, Ronny Falk, Jenny Fall, Mattias Forsberg, Marcus Gry G. Björklund, Kristoffer Gumbel, Asif Halimi, Inga Hallin, Carl Hamsten, Marianne Hansson, My Hedhammar, Görel Hercules, Caroline Kampf, Karin Larsson, Mats Lindskog, Wald Lodewyckx, Jan Lund, Joakim Lundberg, Kristina Magnusson, Erik Malm, Peter Nilsson, Jenny Odling, Per Oksvold, Ingmarie Olsson, Emma Oster, Jenny Ottosson, Linda Paavilainen, Anja Persson, Rebecca Rimini, Johan Rockberg, Marcus Runeson, Asa Sivertsson, Anna Sköllerö, Johanna Steen, Maria Stenvall, Fredrik Sterky, Sara Strömberg, Mårten Sundberg, Hanna Tegel, Samuel Tourle, Eva Wahlund, Annelie Waldén, Jinghong Wan, Henrik Wernérus, Joakim Westberg, Kenneth Wester, Ulla Wrethagen, Lan Lan L. Xu, Sophia Hober, and Fredrik Pontén. A human protein atlas for normal and cancer tissues based on antibody proteomics. *Molecular & cellular proteomics : MCP*, 4(12): 1920–1932, December 2005. ISSN 1535-9476. doi: 10.1074/mcp.M500279-MCP200. URL <http://dx.doi.org/10.1074/mcp.M500279-MCP200>.

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