

# Package ‘xps’

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**Title** Processing and Analysis of Affymetrix Oligonucleotide Arrays  
including Exon Arrays, Whole Genome Arrays and Plate Arrays

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**Depends** R (>= 2.6.0), methods, utils

**Suggests** tools

**Description** The package handles pre-processing, normalization, filtering and analysis of Affymetrix GeneChip expression arrays, including exon arrays (Exon 1.0 ST: core, extended, full probesets), gene arrays (Gene 1.0 ST) and plate arrays on computers with 1 GB RAM only. It imports Affymetrix .CDF, .CLF, .PGF and .CEL as well as annotation files, and computes e.g. RMA, MAS5, FARMS, DFW, FIRMA, tRMA, MAS5-calls, DABG-calls, I/NI-calls. It is an R wrapper to XPS (eXpression Profiling System), which is based on ROOT, an object-oriented framework developed at CERN. Thus, the prior installation of ROOT is a prerequisite for the usage of this package, however, no knowledge of ROOT is required. ROOT is licensed under LGPL and can be downloaded from <http://root.cern.ch>.

**License** GPL (>= 2.0)

**Collate** utils.R TreeSetClasses.R methods.ProjectInfo.R  
methods.Filter.R methods.PreFilter.R methods.UniFilter.R  
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xps-package	<i>xps Package Overview</i>
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## Description

xps Package Overview

## Details

Important data classes: [SchemeTreeSet](#), [DataTreeSet](#), [ExprTreeSet](#), [CallTreeSet](#), [FilterTreeSet](#), [AnalysisTreeSet](#). Full help on methods and associated functions is available from within class help pages.

Additional data classes: [ProjectInfo](#), [PreFilter](#), [UniFilter](#).

The package handles pre-processing, normalization, filtering and analysis of Affymetrix GeneChip expression arrays, including exon array systems (Exon 1.0 ST: core, extended, full probesets), gene array systems (Gene 1.0 ST) and plate array systems on computers with 1 GB RAM only. It imports Affymetrix .CDF, .CLF, .PGF and .CEL as well as Affymetrix annotation files, and computes e.g. RMA, MAS5, FARMS, DFW, MAS5-calls, DABG-calls, I/NI-calls. It is an R wrapper to XPS (eXpression Profiling System), which is based on ROOT, an object-oriented framework developed at CERN. Thus, the prior installation of ROOT is a prerequisite for the usage of this package, see the README file. However, no knowledge of ROOT is required. ROOT is licensed under LGPL and can be downloaded from <http://root.cern.ch>.

## Author(s)

Christian Stratowa <cstrato@aon.at>

---

addData-methods	<i>Import additional CEL files into a DataTreeSet</i>
-----------------	---

---

## Description

Import additional CEL files into a DataTreeSet and update [ROOT](#) data file.

*Usage*

```
addData(object, celdir = NULL, celfiles = "", celnames = NULL, project = NULL, verbose = TRUE)
```

**Arguments**

object	object of class DataTreeSet.
celdir	system directory containing the CEL-files for corresponding scheme.
celfiles	optional vector of CEL-files to be imported.
celnames	optional vector of names which should replace the CEL-file names.
project	optional class <a href="#">ProjectInfo</a> .
verbose	logical, if TRUE print status information.

**Details**

Import additional CEL-files and update [ROOT](#) data file rootfile.

To import CEL-files from different directories, vector `celfiles` must contain the full path for each CEL-file and `celdir` must be `celdir=NULL`.

**Value**

A DataTreeSet object.

**Author(s)**

Christian Stratowa

**See Also**

[import.data](#), [root.data](#)

**Examples**

```
## get scheme and import subset of CEL-files from package
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- import.data(scheme.test3, "tmp_test3", celdir=paste(path.package("xps"), "raw", sep="/"),
  celfiles=c("TestA1.CEL", "TestB2.CEL"), verbose=FALSE)

unlist(treeNames(data.test3))

## add further subset of CEL-files
data.test3 <- addData(data.test3, celdir=paste(path.package("xps"), "raw", sep="/"),
  celfiles=c("TestA2.CEL", "TestB1.CEL"), verbose=FALSE)

unlist(treeNames(data.test3))
```

**Description**

Functions to detect possible RNA degradation.

**Usage**

```
AffyRNAdeg(xps.data, treename = "*", qualopt = "raw", log.it = TRUE)
```

```
summaryAffyRNAdeg(rna.deg, signif.digits=3)
```

```
plotAffyRNAdeg(rna.deg, transform = "shift.scale", col = NULL, summary = FALSE, add.legend = FALSE)
```

```
xpsRNAdeg(object, ...)
```

**Arguments**

<code>xps.data</code>	object of class <code>QualTreeSet</code> .
<code>treename</code>	vector of tree names to export.
<code>qualopt</code>	option determining the data to which to apply qualification, one of 'raw', 'adjusted', 'normalized'.
<code>log.it</code>	logical, if TRUE, then probe data is log <sub>2</sub> transformed.
<code>rna.deg</code>	list, output from <code>AffyRNAdeg</code> .
<code>signif.digits</code>	number of significant digits to show.
<code>transform</code>	transform data before plotting, one of "shift.scale", "shift.only", "none".
<code>col</code>	vector of colors for plot, length is number of samples.
<code>summary</code>	logical, if TRUE then the slope of <code>summaryAffyRNAdeg</code> will be plotted.
<code>add.legend</code>	logical or integer, if TRUE or larger than zero then a legend with the tree names will be drawn.
<code>object</code>	object of class <code>QualTreeSet</code> .
<code>...</code>	optional arguments to be passed to <code>plotAffyRNAdeg</code> .

**Details**

Since probes within a probeset are ordered directionally from the 5' end to the 3' end, it is possible to estimate the quality (degradation status) of the RNA.

Function `AffyRNAdeg` averages the probe intensities by location in the probeset, with the average taken over all probesets with identical number of probes.

Function `summaryAffyRNAdeg` produces a single summary statistic for each array.

Function `plotAffyRNAdeg` produces a side-by-side plot of the averaged intensities. Option `transform = "none"` shows the averaged intensities for each array while option "shift" staggers the plots for individual arrays vertically to make the display easier to read, and option "scale" normalizes the averaged intensities so that the standard deviation is equal to one.

Setting parameter `add.legend = TRUE` will add a legend containing all tree names to the plot, while setting e.g. `add.legend = 6` will only show the first 6 tree names.

**Value**

`AffyRNAdeg` returns a list with following components:

<code>N</code>	number of probesets with identical number of probes
<code>sample.names</code>	names of samples, derived from affy batch object
<code>mns</code>	average intensity by probe position

ses	standard errors for probe position averages
slope	from linear regression of means.by.number
pvalue	from linear regression of means.by.number

**Author(s)**

Christian Stratowa, adapted from package affy

**Examples**

```
## Not run:
rnadeg <- xpsRNAdeg(r1m.all, treename="*", qualopt="raw")
plotAffyRNAdeg(rnadeg)

rnadeg <- AffyRNAdeg(r1m.all)
result <- summaryAffyRNAdeg(rnadeg)

## plot RNA degradation
plotAffyRNAdeg(rnadeg)

## plot slope of RNA degradation
plotAffyRNAdeg(rnadeg, summary = TRUE)

## End(Not run)
```

---

AnalysisTreeSet-class *Class AnalysisTreeSet*

---

**Description**

This class provides the link to the [ROOT](#) analysis file and the [ROOT](#) trees contained therein. It extends class [ProcesSet](#).

**Objects from the Class**

Objects are currently created using function [unifilter](#).

**Slots**

**fltrset:** Object of class "FilterTreeSet" providing indirect access to the [ExprTreeSet](#) used and the [UniFilter](#) settings.

**scheme:** Object of class "SchemeTreeSet" providing access to [ROOT](#) scheme file.

**data:** Object of class "data.frame". The data.frame contains the data of the unitest stored in [ROOT](#) data trees.

**params:** Object of class "list" representing relevant parameters.

**setname:** Object of class "character" representing the name to the [ROOT](#) file subdirectory where the [ROOT](#) trees are stored, currently 'UniFilterSet'.

**settype:** Object of class "character" describing the type of treeset stored in setname, currently 'unifilter'.

**rootfile:** Object of class "character" representing the name of the [ROOT](#) file, including full path.



**filedir:** Object of class "character" describing the full path to the system directory where rootfile is stored.

**numtrees:** Object of class "numeric" representing the number of [ROOT](#) trees stored in subdirectory setname.

**treenames:** Object of class "list" representing the names of the [ROOT](#) trees stored in subdirectory setname.

### Extends

Class "[ProcesSet](#)", directly. Class "[TreeSet](#)", by class "[ProcesSet](#)", distance 2.

### Methods

**filterTreeset** signature(object = "AnalysisTreeSet"): extracts slot fltrset.

**getTreeData** signature(object = "AnalysisTreeSet"): exports tree data and returns a data.frame.

**validData** signature(object = "AnalysisTreeSet"): extracts data.frame data.

**validFilter** signature(object = "AnalysisTreeSet"): extracts data.frame data from fltrset.

**volcanoplot** signature(x = "AnalysisTreeSet"): creates a volcano-plot.

### Author(s)

Christian Stratowa

### See Also

related classes [FilterTreeSet](#).

### Examples

```
showClass("AnalysisTreeSet")
```

---

attachBgrd-methods      *Attach/Remove Background Intensities*

---

### Description

Attach/remove background intensities to/from [DataTreeSet](#).

*Usage*

```
attachBgrd(object, treenames = "*")
```

```
removeBgrd(object)
```

### Arguments

**object**                    Object of class "[DataTreeSet](#)".

**treenames**                Object of class "list" representing the names of the [ROOT](#) background trees.

**Details**

Whenever one of the [bgcorrect](#) methods will be applied to raw CEL intensities, the background intensities will be stored in [ROOT](#) background trees. However, the background intensities will not be saved as data.frame bgrd, thus avoiding memory problems. Function `attachBgrd` allows to fill slot `bgrd` on demand.

`attachBgrd` exports intensities from background trees from [ROOT](#) data file and saves as data.frame `bgrd`. `treenames` is a vector of tree names to attach; for `treenames="*"` all trees from slot `treenames` will be exported and background intensities attached as data.frame `bgrd`.

`removeBgrd` removes background intensities from [DataTreeSet](#) and replaces data.frame `bgrd` with an empty data.frame of `dim(0,0)`.

**Value**

A [DataTreeSet](#) object.

**Note**

Do not use `attachBgrd` unless you know that your computer has sufficient RAM, especially when using exon arrays. It may be advisable to use a subset of `treenames` only.

**Author(s)**

Christian Stratowa

**See Also**

[attachInten](#), [removeInten](#)

---

attachCall-methods      *Attach/Remove Detecion Call Measures*

---

**Description**

Attach/remove detection call and detection p-value to/from [CallTreeSet](#).

*Usage*

```
attachCall(object, treenames = "*")
```

```
attachPVal(object, treenames = "*")
```

```
removeCall(object)
```

```
removePVal(object)
```

**Arguments**

`object`            Object of class "[CallTreeSet](#)".

`treenames`        Object of class "`list`" representing the names of the [ROOT](#) call trees.

## Details

By default detection calls will be saved in class `CallTreeSet` in slots `data` and `detcall`, respectively, since usually the `data.frames` obtained as result of e.g. `mas5.call` are of reasonable size. However, when computing many arrays, especially exon arrays at probeset levels, it may be better to compute detection calls with `slot add.data=FALSE` thus avoiding memory problems. In this case, functions `attachCall` and `attachPVal` allow to fill slots `detcall` and `data`, respectively, on demand.

`attachCall` exports detection calls from call trees from `ROOT` call file and saves as `data.frame detcall`. `treenames` is a vector of tree names to attach; for `treenames="*"` all trees from slot `treenames` will be exported and detection calls attached as `data.frame detcall`.

`attachPVal` exports detection p-values from call trees from `ROOT` call file and saves as `data.frame data`. `treenames` is a vector of tree names to attach; for `treenames="*"` all trees from slot `treenames` will be exported and detection p-values attached as `data.frame data`.

`removeCall` removes detection calls from `CallTreeSet` and replaces `data.frame detcall` with an empty `data.frame` of `dim(0,0)`.

`removePVal` removes detection p-values from `CallTreeSet` and replaces `data.frame data` with an empty `data.frame` of `dim(0,0)`.

## Value

A `CallTreeSet` object.

## Note

Do not use `attachCall` and `attachPVal` unless you know that your computer has sufficient RAM, especially when using exon arrays. It may be advisable to use a subset of `treenames` only.

## Author(s)

Christian Stratowa

## See Also

[attachExpr](#), [removeExpr](#)

## Examples

```
## first, load ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

## MAS5 detection call
call.mas5 <- mas5.call(data.test3, "tmp_Test3Call0", tmpdir="", add.data=FALSE, verbose=FALSE)

## attach data
call.mas5 <- attachPVal(call.mas5)
call.mas5 <- attachCall(call.mas5)

## get data.frames
pval.mas5 <- pvalData(call.mas5)
pres.mas5 <- presCall(call.mas5)
head(pval.mas5)
head(pres.mas5)
```

```
## remove data
call.mas5 <- removePVal(call.mas5)
call.mas5 <- removeCall(call.mas5)

rm(scheme.test3, data.test3)
gc()
```

---

attachData-methods      *Attach/Remove Data*

---

### Description

Attach/remove data from trees to/from [ProcesSet](#).

#### Usage

```
attachData(object, treenames = character(0), varlist = character(0), outfile = "data.txt")
removeData(object)
```

### Arguments

object	Object of class "ProcesSet".
treename	vector of tree names to export.
varlist	names of tree leaves to export
outfile	name of output file.

### Details

attachData exports varlist from tree(s) with treenames and and saves the result as data.frame in slot data. Possible values of parameter varlist are described in [export](#).

removeData removes data from slot data and replaces data.frame data with an empty data.frame of dim(0,0).

### Value

A [ProcesSet](#) object.

### Author(s)

Christian Stratowa

### See Also

[attachDataXY](#), [attachInten](#)

---

attachDataXY-methods    *Attach/Remove (X,Y)-Coordinates*

---

### Description

Attach/remove (x,y)-coordinates of raw CEL-files to/from [DataTreeSet](#).

#### Usage

```
attachDataXY(object)
removeDataXY(object)
```

### Arguments

object                    Object of class "DataTreeSet".

### Details

attachDataXY exports (x,y)-coordinates only from data tree of [ROOT](#) data file and saves it as data.frame in slot data.

removeDataXY removes (x,y)-coordinates from slot data and replaces data.frame data with an empty data.frame of dim(0,0).

### Value

A [DataTreeSet](#) object.

### Author(s)

Christian Stratowa

### See Also

[attachInten](#), [removeInten](#)

### Examples

```
## first, load ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

## attach (x,y)-coordinates
data.test3 <- attachDataXY(data.test3)

## get data.frame
xy <- treeData(data.test3)
head(xy)

## remove (x,y)-coordinates
data.test3 <- removeDataXY(data.test3)

rm(scheme.test3, data.test3)
gc()
```

---

attachExpr-methods      *Attach/Remove Expression Measures*

---

### Description

Attach/remove expression levels to/from [ExprTreeSet](#).

#### Usage

```
attachExpr(object, treenames = "*")
removeExpr(object)
```

### Arguments

`object`            Object of class "ExprTreeSet".  
`treenames`        Object of class "list" representing the names of the [ROOT](#) expression trees.

### Details

By default expression levels will be saved in class [ExprTreeSet](#) as slot data, since usually the `data.frame` obtained as result of e.g. `rma` normalization is of reasonable size. However, when normalizing many arrays, especially exon arrays at probeset levels, it may be better to compute `rma` with `slot.add.data=FALSE` thus avoiding memory problems. In this case, function `attachExpr` allows to fill slot data on demand.

`attachExpr` exports expression levels from expression trees from [ROOT](#) expression file and and saves as `data.frame` data. `treenames` is a vector of tree names to attach; for `treenames="*"` all trees from slot `treenames` will be exported and expression levels attached as `data.frame` data.

`removeExpr` removes expression levels from [ExprTreeSet](#) and replaces `data.frame` data with an empty `data.frame` of `dim(0,0)`.

### Value

A [ExprTreeSet](#) object.

### Note

Do not use `attachExpr` unless you know that your computer has sufficient RAM, especially when using exon arrays. It may be advisable to use a subset of `treenames` only.

### Author(s)

Christian Stratowa

### See Also

[attachCall](#), [removeCall](#)

**Examples**

```
## first, load ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

data.rma <- rma(data.test3, "tmp_Test3RMA0", tmpdir="", background="pmonly", normalize=TRUE, add.data=FALSE, verb=TRUE)

## attach data
data.rma <- attachExpr(data.rma)

## get data.frame
expr.rma <- validData(data.rma)
head(expr.rma)

## remove data
data.rma <- removeExpr(data.rma)

rm(scheme.test3, data.test3)
gc()
```

---

attachInten-methods     *Attach/Remove Intensities*

---

**Description**

Attach/remove raw CEL intensities to/from [DataTreeSet](#).

*Usage*

```
attachInten(object, treenames = "*")
removeInten(object)
```

**Arguments**

**object**                Object of class "DataTreeSet".

**treenames**            Object of class "list" representing the names of the [ROOT](#) data trees.

**Details**

When CEL files will be imported using function [import.data](#), the raw intensities will be stored in [ROOT](#) data trees. However, the intensities will not be saved in class [DataTreeSet](#) as slot data, thus avoiding memory problems. Function [attachInten](#) allows to fill slot data on demand.

[attachInten](#) exports intensities from data trees from [ROOT](#) data file and and saves as data.frame data. `treenames` is a vector of tree names to attach; for `treenames="*"` all trees from slot `treenames` will be exported and intensities attached as data.frame data.

[removeInten](#) removes intensities from [DataTreeSet](#) and replaces data.frame data with an empty data.frame of `dim(0,0)`.

**Value**

A [DataTreeSet](#) object.

**Note**

Do not use `attachInten` unless you know that your computer has sufficient RAM, especially when using exon arrays. It may be advisable to use a subset of `treenames` only.

**Author(s)**

Christian Stratowa

**See Also**

[attachBgrd](#), [removeBgrd](#)

**Examples**

```
## load existing ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))
dim(intensity(data.test3))

data.test3 <- attachInten(data.test3)
dim(intensity(data.test3))
head(intensity(data.test3))

data.test3 <- removeInten(data.test3)
dim(intensity(data.test3))
```

---

attachMask-methods      *Attach/Remove Scheme Mask*

---

**Description**

Attach/remove scheme mask to/from [SchemeTreeSet](#) or to slot scheme of [DataTreeSet](#).

**Usage**

```
attachMask(object)
```

```
removeMask(object)
```

**Arguments**

`object`                  Object of class "SchemeTreeSet" or "DataTreeSet".

**Details**

`attachMask` exports mask from scheme tree from `ROOT` scheme file and saves mask as `data.frame` mask of slot scheme.

`removeMask` removes mask from [SchemeTreeSet](#) or from slot scheme of [DataTreeSet](#) and replaces `data.frame` mask with an empty `data.frame` of `dim(0,0)`.

**Value**

A [DataTreeSet](#) object or [SchemeTreeSet](#).



**Note**

Do not use attachMask unless you know that your computer has sufficient RAM, especially for exon array schemes.

**Author(s)**

Christian Stratowa

**See Also**

[import.expr.scheme](#), [import.exon.scheme](#)

**Examples**

```
## load existing ROOT scheme file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
dim(chipMask(scheme.test3))

scheme.test3 <- attachMask(scheme.test3)
dim(chipMask(scheme.test3))
head(chipMask(scheme.test3))

scheme.test3 <- removeMask(scheme.test3)
dim(chipMask(scheme.test3))
```

---

attachProbe-methods     *Attach/Remove Probe Sequence and/or GC Content*

---

**Description**

Attach/remove probe sequence and/or GC content to/from [SchemeTreeSet](#) or to slot scheme of [DataTreeSet](#).

*Usage*

```
attachProbe(object, varlist)
attachProbeContentGC(object)
attachProbeSequence(object)
removeProbe(object)
removeProbeContentGC(object)
removeProbeSequence(object)
```

**Arguments**

object	Object of class "SchemeTreeSet" or "DataTreeSet".
varlist	names of probe tree leaves to import to slot probe.

**Details**

Function attachProbe exports leaves from probe tree of [ROOT](#) scheme file and and saves the data as data.frame probe of slot scheme.

Following varlist parameters are valid:

fPosition: probe interrogation position.  
 fSequence: probe sequence.  
 fNumberGC: number of G/C nucleotides in probe sequence.  
 fTm: probe melting temperature dependent on G/C number.  
 fIsAntisense: probe type (sense/antisense).

Function `attachProbeContentGC` saves `fNumberGC` in data.frame probe of `SchemeTreeSet` or in slot scheme of `DataTreeSet`.

Function `attachProbeSequence` saves `fSequence` in data.frame probe of `SchemeTreeSet`.

Function `removeProbe` removes probe data from `SchemeTreeSet` or from slot scheme of `DataTreeSet` and replaces data.frame probe with an empty data.frame of `dim(0,0)`.

### Value

A `SchemeTreeSet` object or `DataTreeSet`.

### Note

Do not use `attachProbe` unless you know that your computer has sufficient RAM, especially for exon array schemes.

### Author(s)

Christian Stratowa

### See Also

[attachMask](#)

### Examples

```
## load existing ROOT scheme file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
dim(chipProbe(scheme.test3))

scheme.test3 <- attachProbe(scheme.test3, varlist="fSequence:fNumberGC")
dim(chipProbe(scheme.test3))
head(chipProbe(scheme.test3))

scheme.test3 <- removeProbe(scheme.test3)
dim(chipProbe(scheme.test3))
```

---

attachUnitNames-methods

*Attach/Remove Unit Names*

---

### Description

Attach/remove unit names, i.e. the Affymetrix probeset IDs to/from `SchemeTreeSet` or to slot scheme of `DataTreeSet`.

*Usage*

```
attachUnitNames(object, treetype = "idx")
removeUnitNames(object)
```

**Arguments**

object            Object of class "SchemeTreeSet" or "DataTreeSet".  
treetype         the unit tree type, i.e. 'idx' or 'pbs'.

**Details**

attachUnitNames exports "UnitName" from unit tree of [ROOT](#) scheme file and saves it as data.frame in slot unitname.

removeUnitNames removes unitname from slot unitname and replaces data.frame unitname with an empty data.frame of dim(0,0).

For treetype="idx" the internal "UNIT\_ID" will be mapped to the Affymetrix probeset IDs of the expression arrays or to the transcript\_cluster\_ids of the exon arrays, respectively, as "UnitName".

For treetype="pbs" the internal "UNIT\_ID" will be mapped to the Affymetrix probeset\_ids of the exon arrays as "UnitName".

**Value**

A [DataTreeSet](#) object or [SchemeTreeSet](#).

**Note**

Do not use attachUnitNames unless you know that your computer has sufficient RAM, especially for exon array schemes.

**Author(s)**

Christian Stratowa

**See Also**

[attachMask](#), [removeMask](#)

**Examples**

```
## first, load ROOT scheme file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))

## attach unitname
scheme.test3 <- attachUnitNames(scheme.test3)

## get data.frame
unitnames <- unitNames(scheme.test3)
head(unitnames)

## remove unitname
scheme.test3 <- removeUnitNames(scheme.test3)

rm(scheme.test3)
gc()
```

bgcorrect

*Background Correction***Description**

Background corrects probe intensities in an object of class [DataTreeSet](#).

**Usage**

```
bgcorrect(xps.data, filename = character(0), filedir = getwd(), tmpdir = "", update = FALSE, select = "all")
bgcorrect.gc(xps.data, filename = character(0), filedir = getwd(), tmpdir = "", update = FALSE, select = "all")
bgcorrect.mas4(xps.data, filename = character(0), filedir = getwd(), tmpdir = "", update = FALSE, select = "all")
bgcorrect.mas5(xps.data, filename = character(0), filedir = getwd(), tmpdir = "", update = FALSE, select = "all")
bgcorrect.rma(xps.data, filename = character(0), filedir = getwd(), tmpdir = "", update = FALSE, select = "all")
xpsBgCorrect(object, ...)
```

**Arguments**

xps.data	object of class <a href="#">DataTreeSet</a> .
filename	file name of ROOT data file.
filedir	system directory where ROOT data file should be stored.
tmpdir	optional temporary directory where temporary ROOT files should be stored.
update	logical. If TRUE the existing ROOT data file filename will be updated.
select	type of probes to select for background correction.
method	background method to use.
option	type of background correction to use.
exonlevel	exon annotation level determining which probes should be used for summarization; exon/genome arrays only.
params	vector of parameters for background method.
verbose	logical, if TRUE print status information.
object	object of class <a href="#">DataSet</a> .
...	the arguments described above.

**Details**

Background corrects probe intensities in an object of class [DataTreeSet](#).

xpsBgCorrect is the [DataSet](#) method called by function `bgcorrect`, containing the same parameters.

**Value**

An [DataTreeSet](#)

**Author(s)**

Christian Stratowa

**See Also**[express](#)**Examples**

```
## first, load ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

## MAS4 sector background
data.bg.mas4 <- bgcorrect.mas4(data.test3, "tmp_Test3MAS4Bgrd", filedir=getwd(), tmpdir="", verbose=FALSE)

## need to attach background intensities
data.bg.mas4 <- attachBgrd(data.bg.mas4)

## get data.frame
bg.mas4 <- validBgrd(data.bg.mas4)
head(bg.mas4)

## plot images
if (interactive()) {
  image.dev(data.bg.mas4, bg=TRUE, col=rainbow(32))
  image(matrix(bg.mas4[,1], ncol=ncols(schemeSet(data.bg.mas4)), nrow=nrows(schemeSet(data.bg.mas4))))
}

## Not run:
## examples using Affymetrix human tissue dataset (see also xps/examples/script4exon.R)

## example - exon array, e.g. HuEx-1_0-st-v2:
scmdir <- "/Volumes/GigaDrive/CRAN/Workspaces/Schemes"
datdir <- "/Volumes/GigaDrive/CRAN/Workspaces/ROOTData"
scheme.exon <- root.scheme(paste(scmdir, "Scheme_HuEx10stv2r2_na25.root", sep="/"))
data.exon <- root.data(scheme.exon, paste(datdir, "HuTissuesExon_cel.root", sep="/"))

## compute rma background
workdir <- "/Volumes/GigaDrive/CRAN/Workspaces/Exon/hutissues/exon"
data.bg.rma <- bgcorrect(data.exon, "HuExonRMABgrd", filedir=workdir, tmpdir="",
  method="rma", select="antigenomic", option="pmonly:epanechnikov",
  params=c(16384), exonlevel="metacore+affx")

# or alternatively:
data.bg.rma <- bgcorrect.rma(data.exon, "HuExonRMABgrd", filedir=workdir, tmpdir="",
  select="antigenomic", exonlevel="metacore+affx")

## End(Not run)
```

**Description**

Produce box-and-whisker plot(s) of the positive and negative feature intensities.

*Usage*

```
borderplot(x, type = c("pos", "neg"), qualopt = "raw", transfo = log2,
```

**Arguments**

`x` object of class [QualTreeSet](#).  
`type` type of border elements to be used, one of “pos”, “neg”, or both.  
`qualopt` character string specifying whether to draw boxplots for “raw”, “adjusted”, or “normalized” border intensities.  
`transfo` a valid function to transform the data, usually “log2”, or “0”.  
`range` determines how far the plot whiskers extend out from the box.  
`names` optional vector of sample names.  
`ylim` the y limits of the plot.  
`bmar` optional list for bottom margin and axis label magnification `cex.axis`.  
`las` the style of axis labels.  
`...` optional arguments to be passed to `borderplot`.

**Details**

Creates a boxplot of the positive and negative feature intensities for an object of class [QualTreeSet](#).

For `names=NULL` full tree names will be displayed while for `names="namepart"` tree names will be displayed without name extension. If `names` is a vector of tree names, only these columns will be displayed as boxplot.

For `bmar=NULL` the default list `bmar = list(b=6, cex.axis=1.0)` will be used initially. However, both bottom margin `b` and axis label magnification `cex.axis` will be adjusted depending on the number of label characters and the number of samples.

**Author(s)**

Christian Stratowa

**See Also**

[plotBorder](#), [coiplot](#)

**Examples**

```
## Not run:
## border intensities, created by e.g. rmaPLM()
getTreeNames(rootFile(rlm.all), treetype="brd")
borderplot(rlm.all)
borderplot(rlm.all, type="pos")
borderplot(rlm.all, type="neg")

## End(Not run)
```

boxplot-methods

*Box Plots***Description**

Produce box-and-whisker plot(s) of the samples.

*Usage*

```
boxplot(x, which = "", size = 0, transfo = log2, range = 0, names = "namepart", bmar = NULL, ...)
```

**Arguments**

x	object of class <a href="#">DataTreeSet</a> , <a href="#">ExprTreeSet</a> or <a href="#">QualTreeSet</a> .
which	type of probes to be used, for details see <a href="#">validData</a> .
size	length of sequence to be generated as subset.
transfo	a valid function to transform the data, usually “log2”, or “0”.
range	determines how far the plot whiskers extend out from the box.
names	optional vector of sample names.
bmar	optional list for bottom margin and axis label magnification <code>cex.axis</code> .
...	optional arguments to be passed to <code>boxplot</code> .

**Details**

Creates a boxplot for slot data for an object of class [DataTreeSet](#), [ExprTreeSet](#) or [QualTreeSet](#).

For `names=NULL` full column names of slot data will be displayed while for `names="namepart"` column names will be displayed without name extension. If `names` is a vector of column names, only these columns will displayed as boxplot.

For `bmar=NULL` the default list `bmar = list(b=6, cex.axis=1.0)` will be used initially. However, both bottom margin and axis label magnification will be adjusted depending on the number of label characters and the number of samples.

**Note**

For a [DataTreeSet](#) object, data must first be attached using method [attachInten](#).

Alternatively it is possible to use the pre-calculated quantiles stored in the `userinfo` of the data trees by calling `which="userinfo:varlist"`, where the `varlist` to call is described in method [treeInfo](#).

**Author(s)**

Christian Stratowa

**See Also**

[plotBoxplot](#), [boxplot](#)

**Examples**

```
## load existing ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

## need to attach scheme mask and probe intensities only if "userinfo" is not used
data.test3 <- attachMask(data.test3)
data.test3 <- attachInten(data.test3)

if (interactive()) {
  boxplot(data.test3)
}

## optionally remove mask and data to free memory
data.test3 <- removeInten(data.test3)
data.test3 <- removeMask(data.test3)

## alternatively use the quantiles stored in userinfo of trees
if (interactive()) {
  boxplot(data.test3, which="userinfo:fIntenQuant")
}

rm(scheme.test3, data.test3)
gc()
```

---

callFilter-methods      *Detection Call Filter*

---

**Description**

Detection Call Filter.

The cutoff value defines the upper threshold for allowed detection call p-values. If e.g. the number of samples exceeding this cutoff value is greater than samples then the corresponding expression dataframe row is flagged, i.e. flag = 0.

The Detection Call Filter flags all rows with: flag = (sum(call[i] >= cutoff) >= samples)

*Usage*

```
callFilter(object)
callFilter(object, value)<-
```

**Arguments**

object	object of class PreFilter or UniFilter.
value	character vector c(cutoff, samples, condition).

**Details**

The method callFilter initializes the following parameters:

cutoff:	the cutoff value for the filter: cutoff = 1.0: present/absent call is used. cutoff < 1.0: detection p-value is used as cutoff.
samples:	this value depends on the condition used:
condition:	condition="samples": number of samples (default): condition="percent": percent of samples.



**Value**

An initialized [PreFilter](#) or [UniFilter](#) object.

**Author(s)**

Christian Stratowa

**Examples**

```
## initialize PreFilter
prefltr <- PreFilter()
callFilter(prefltr) <- c(0.02,80.0,"percent")
str(prefltr)

## initialize UniFilter
unifltr <- UniFilter()
callFilter(unifltr) <- c(0.02,80.0,"percent")
str(unifltr)
```

---

callplot-methods      *Barplot of Percent Present and Absent Calls.*

---

**Description**

Creates a barplot of percent Present/Marginal/Absent calls.

*Usage*

```
callplot(x,            beside = TRUE,            names = "namepart",            col    = c("red","green","blue")
```

**Arguments**

x	object of class <a href="#">CallTreeSet</a> .
beside	logical. If FALSE, the columns of height are portrayed as stacked bars, and if TRUE the columns are portrayed as juxtaposed bars.
names	optional vector of sample names.
col	color for P/M/A bars
legend	legend for the plot, defaults to P/M/A.
ylim	the y limits of the plot.
ylab	a label for the y axis.
las	the style of axis labels.
...	optional arguments to be passed to <code>barplot</code> .

**Details**

Creates a barplot of percent Present/Marginal/Absent calls.

For `names=NULL` full column names of slot data will be displayed while for `names="namepart"` column names will be displayed without name extension. If `names` is a vector of column names, only these columns will displayed as callplot.

**Author(s)**

Christian Stratowa

**See Also**

[plotCall](#), [pmpplot](#)

---

CallTreeSet-class      *Class CallTreeSet*

---

**Description**

This class provides the link to the [ROOT](#) call file and the [ROOT](#) trees contained therein. It extends class [ProcesSet](#).

**Objects from the Class**

Objects are created using functions [mas5.call](#) or [dabg.call](#), respectively.

**Slots**

**calltype:** Object of class "character" representing the call type, i.e. 'mas5' or 'dabg'.

**detcall:** Object of class "data.frame". The data.frame can contain the detection calls stored in [ROOT](#) call trees.

**scheme:** Object of class "SchemeTreeSet" providing access to [ROOT](#) scheme file.

**data:** Object of class "data.frame". The data.frame can contain the data (i.e. p-values) stored in [ROOT](#) call trees.

**params:** Object of class "list" representing relevant parameters.

**setname:** Object of class "character" representing the name to the [ROOT](#) file subdirectory where the [ROOT](#) call trees are stored, usually 'CallTreeSet'.

**settype:** Object of class "character" describing the type of treeset stored in setname, usually 'preprocess'.

**rootfile:** Object of class "character" representing the name of the [ROOT](#) call file, including full path.

**filedir:** Object of class "character" describing the full path to the system directory where rootfile is stored.

**numtrees:** Object of class "numeric" representing the number of [ROOT](#) trees stored in subdirectory setname.

**treenames:** Object of class "list" representing the names of the [ROOT](#) trees stored in subdirectory setname.

**Extends**

Class "[ProcesSet](#)", directly. Class "[TreeSet](#)", by class "[ProcesSet](#)", distance 2.

## Methods

**attachCall** signature(object = "CallTreeSet"): exports detection call data from [ROOT](#) call file and and saves as data.frame detcall.

**attachPVal** signature(object = "CallTreeSet"): exports call p-values from [ROOT](#) call file and and saves as data.frame data.

**callplot** signature(x = "CallTreeSet"): creates a barplot of percent present and absent calls.

**presCall** signature(object = "CallTreeSet"): extracts the detection call data.frame.

**presCall<-** signature(object = "CallTreeSet", value = "data.frame"): replaces the detection call data.frame.

**pvalData** signature(object = "CallTreeSet"): extracts the detection p-value data.frame.

**pvalData<-** signature(object = "CallTreeSet", value = "data.frame"): replaces the detection p-value data.frame.

**removeCall** signature(object = "CallTreeSet"): replaces data.frame detcall with an empty data.frame of dim(0,0).

**removePVal** signature(object = "CallTreeSet"): replaces data.frame data with an empty data.frame of dim(0,0).

**validCall** signature(object = "CallTreeSet"): extracts a subset of columns from data.frame detcall.

**validPVal** signature(object = "CallTreeSet"): extracts a subset of columns from data.frame data.

## Author(s)

Christian Stratowa

## See Also

related classes [DataTreeSet](#), [ExprTreeSet](#).

## Examples

```
showClass("CallTreeSet")
```

---

coiplot-methods

*Center-Of-Intensity QC Plots*

---

## Description

Produce Center-Of-Intensity plot(s) of the positive and negative feature intensities.

### Usage

```
coiplot(x, type = c("pos", "neg"), qualopt = "raw", radius = 0.5, linec
```

## Arguments

x	object of class <a href="#">QualTreeSet</a> .
type	type of border elements to be used, one of “pos”, “neg”, or both.
qualopt	character string specifying whether to draw boxplots for “raw”, “adjusted”, or “normalized” border intensities.
radius	determines the radius within which the COI for each array should be located.
linecol	the color of the ablines and the circle to be drawn.
visible	logical, if TRUE then arrays outside the circle with radius will be flagged by labeling the data point with the array name.
...	optional arguments to be passed to <code>coiplot</code> .

## Details

Produces Center-Of-Intensity (COI) plot(s) of the positive and negative feature intensities for an object of class [QualTreeSet](#). This plot is useful for detecting spatial biases in intensities on an array.

Mean intensities for the left, right, top and bottom border elements are calculated, separated into positive and negative controls, and the “center of intensity” is calculated on a relative scale [-1,1]. Arrays with a COI outside a range with radius are considered to be outliers. If `visible = TRUE` then outlier arrays will be flagged by labeling the data point(s) with the array name(s).

## Value

The names of the outlier arrays, otherwise NULL.

## Author(s)

Christian Stratowa

## See Also

[plotCOI](#), [borderplot](#)

## Examples

```
## Not run:
## border intensities, created by e.g. rmaPLM()
coiplot(rlm.all)
coiplot(rlm.all, type="pos")
coiplot(rlm.all, type="neg", radius=0.1)

## End(Not run)
```

---

corplot-methods      *Array-Array Expression Level Correlation Plot*

---

### Description

A heat map of the array-array Spearman rank correlation coefficients.

#### Usage

```
corplot(x, which = "UnitName", transfo = log2, method = "spearman",
```

### Arguments

`x`                    object of class [ExprTreeSet](#).  
`which`                type of probes to be used, for details see [validData](#).  
`transfo`              a valid function to transform the data, usually "log2", or "0".  
`method`              a character string indicating which correlation coefficient is to be computed.  
`col`                  vector of colors for plot, length is number of samples.  
`names`                optional vector of sample names.  
`sort`                 logical, if TRUE the correlation matrix will be sorted decreasingly.  
`reverse`             logical, if TRUE the correlation matrix will be replaced by  $1 - \text{cor}()$ .  
`bmar`                 optional list for bottom margin and axis label magnification `cex.axis`.  
`add.legend`         logical, if TRUE then a color bar will be drawn.  
`...`                 optional arguments to be passed to plot.

### Details

Produces a heat map of the array-array Spearman rank correlation coefficients for slot data for an object of class [ExprTreeSet](#).

For `names=NULL` full column names of slot data will be displayed while for `names="namepart"` column names will be displayed without name extension. If `names` is a vector of column names, only these columns will displayed as `corplot`.

For `bmar=NULL` the default list `bmar = list(b=6, cex.axis=1.0)` will be used initially. However, both bottom margin and axis label magnification will be adjusted depending on the number of label characters and the number of samples.

### Note

Setting `reverse = FALSE` displays the correlation heat map as in package `afyQCReport`.

### Author(s)

Christian Stratowa

### See Also

[plotCorr](#), [madplot](#)

---

cvFilter-methods      *Coefficient of Variation Filter*

---

### Description

This method initializes the Coefficient of Variation Filter.  
The coefficient of variation is the standard deviation divided by the absolute value of the mean.  
The CV Filter flags all rows with: `flag = (cv >= cutoff)`

#### Usage

```
cvFilter(object)
cvFilter(object, value)<-
```

### Arguments

`object`      object of class `PreFilter`.  
`value`      numeric vector `c(cutoff, trim, epsilon)`.

### Details

The method `cvFilter` initializes the following parameters:

`cutoff`:    the cutoff level for the filter.  
`trim`:      the trim value for trimmed mean (default is `trim=0`).  
`epsilon`:    value to replace mean (default is `epsilon=0.01`):  
              `epsilon > 0`: replace `mean=0` with `epsilon`.  
              `epsilon = 0`: always set `mean=1`.

Note, that for `epsilon = 0` the filter flags all rows with: `stdev >= cutoff`

### Value

An initialized `PreFilter` object.

### Author(s)

Christian Stratowa

### Examples

```
prefltr <- PreFilter()
cvFilter(prefltr) <- c(0.3,0.0,0.01)
str(prefltr)
```

dabg.call

*Detection Above Background Call***Description**

Computes the Detection Above Background Call first implemented for the Exon arrays.

**Usage**

```
dabg.call(xps.data, filename = character(0), filedir = getwd(),
          alpha1 = 0.04, alpha2 = 0.06,
          option = "transcript", exonlevel = "", xps.scheme = NULL, add.data = TRUE, verbose = TRUE)

xpsDABGCall(object, ...)
```

**Arguments**

xps.data	object of class DataTreeSet.
filename	file name of ROOT data file.
filedir	system directory where ROOT data file should be stored.
alpha1	a significance threshold in (0,alpha2).
alpha2	a significance threshold in (alpha1,0.5).
option	option determining the grouping of probes for summarization, one of 'transcript', 'exon', 'probeset'; exon arrays only.
exonlevel	exon annotation level determining which probes should be used for summarization; exon/genome arrays only.
xps.scheme	optional alternative SchemeTreeSet.
add.data	logical. If TRUE call data will be added to slots data and detcall.
verbose	logical, if TRUE print status information.
object	object of class DataTreeSet.
...	the arguments described above.

**Details**

This function generates a detection p-value based on comparing the perfect match probe intensity to the intensity distribution provided by background probes sharing the same GC-content as the PM probe under consideration. For exon/genome arrays special 'antigenomic' background probes of defined GC-content are used, while for expression arrays the Mismatch probes will be grouped by their GC-content.

For exon/genome arrays it is necessary to supply option and exonlevel.

Following options are valid for exon arrays only:

transcript:	expression levels are computed for transcript clusters, i.e. probe sets containing the same 'transcript_cluster_id'.
exon:	expression levels are computed for exon clusters, i.e. probe sets containing the same 'exon_id', where each probe set contains all probes of a given exon.
probeset:	expression levels are computed for individual probe sets, i.e. for each 'probeset_id'.

Following exonlevel annotations are valid for exon arrays:

core:	probesets supported by RefSeq and full-length GenBank transcripts.
metacore:	core meta-probesets.
extended:	probesets with other cDNA support.
metaextended:	extended meta-probesets.
full:	probesets supported by gene predictions only.
metafull:	full meta-probesets.
ambiguous:	ambiguous probesets only.
affx:	standard AFFX controls.
all:	combination of above.

Following exonlevel annotations are valid for whole genome arrays:

core:	probesets with category 'unique' and 'mixed'.
metacore:	probesets with category 'unique' only.
affx:	standard AFFX controls.
all:	combination of above.

Exon levels can also be combined, with following combinations being most useful:

exonlevel="metacore+affx":	core meta-probesets plus AFFX controls
exonlevel="core+extended":	probesets with cDNA support
exonlevel="core+extended+full":	supported plus predicted probesets

Exon level annotations are described in the Affymetrix whitepaper 'exon\_probeset\_trans\_clust\_whitepaper.pdf'.

In order to use an alternative [SchemeTreeSet](#) set the corresponding SchemeTreeSet xps.scheme.

xpsDABGCall is the DataTreeSet method called by function dabg.call, containing the same parameters.

## Value

A [CallTreeSet](#)

## Note

Yes, it is possible to compute DABG detection call for expression arrays, but it is very slow and thus not recommended.

## Author(s)

Christian Stratowa

## References

Affymetrix (2005) Exon Probeset Annotations and Transcript Cluster Groupings, Affymetrix Inc., Santa Clara, CA, exon\_probeset\_trans\_clust\_whitepaper.pdf.

## See Also

[mas5.call](#)



**Examples**

```
## first, load ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

## DABG detection call
call.dabg <- dabg.call(data.test3, "tmp_Test3DABG", verbose=FALSE)

## get data.frames
pval.dabg <- pvalData(call.dabg)
pres.dabg <- presCall(call.dabg)
head(pval.dabg)
head(pres.dabg)

## plot results
if (interactive()) {
  callplot(call.dabg)
}

rm(scheme.test3, data.test3)
gc()
```

DataTreeSet-class

*Class DataTreeSet***Description**

This class provides the link to the [ROOT](#) data file and the [ROOT](#) trees contained therein. It extends class [ProcesSet](#).

**Objects from the Class**

Objects can be created using the functions [import.data](#) or [root.data](#).

**Slots**

**bgreenames:** Object of class "list" representing the names of optional [ROOT](#) background trees.

**bgrd:** Object of class "data.frame". The data.frame can contain background intensities stored in [ROOT](#) background trees.

**projectinfo:** Object of class "ProjectInfo" containing information about the project.

**scheme:** Object of class "SchemeTreeSet" providing access to [ROOT](#) scheme file.

**data:** Object of class "data.frame". The data.frame can contain the data (e.g. intensities) stored in [ROOT](#) data trees.

**params:** Object of class "list" representing relevant parameters.

**setname:** Object of class "character" representing the name to the [ROOT](#) file subdirectory where the [ROOT](#) data trees are stored, usually 'DataTreeSet'.

**settype:** Object of class "character" describing the type of treeset stored in setname, usually 'rawdata'.

**rootfile:** Object of class "character" representing the name of the [ROOT](#) data file, including full path.

**filedir:** Object of class "character" describing the full path to the system directory where rootfile is stored.

**numtrees:** Object of class "numeric" representing the number of [ROOT](#) trees stored in subdirectory setname.

**treenames:** Object of class "list" representing the names of the [ROOT](#) trees stored in subdirectory setname.

## Extends

Class "[ProcesSet](#)", directly. Class "[TreeSet](#)", by class "[ProcesSet](#)", distance 2.

## Methods

**addData** signature(object = "DataTreeSet"): import additional CEL-files and update [ROOT](#) data file rootfile.

**attachBgrd** signature(object = "DataTreeSet"): exports background trees from [ROOT](#) data file and and saves as data.frame bgrd.

**attachDataXY** signature(object = "DataTreeSet"): exports (x,y)-coordinates from [ROOT](#) data file and and saves as data.frame data.

**attachInten** signature(object = "DataTreeSet"): exports intensity trees from [ROOT](#) data file and and saves as data.frame data.

**attachMask** signature(object = "DataTreeSet"): exports scheme tree from [ROOT](#) scheme file and and saves as data.frame mask of slot scheme.

**attachProbeContentGC** signature(object = "DataTreeSet"): exports probe tree from [ROOT](#) scheme file and and saves fNumberGC as data.frame probe.

**attachUnitNames** signature(object = "DataTreeSet"): exports unit tree from [ROOT](#) scheme file and and saves as data.frame unitname of slot scheme.

**background** signature(object = "DataTreeSet"): extracts slot bgrd.

**background<-** signature(object = "DataTreeSet", value = "data.frame"): replaces slot bgrd.

**bgtreeNames** signature(object = "DataTreeSet"): extracts slot bgtreenames.

**indexUnits** signature(object = "DataTreeSet"): extracts (x,y)-coordinates and corresponding indices for all or selected unitIDs.

**intensity** signature(object = "DataTreeSet"): extracts slot data.

**intensity<-** signature(object = "DataTreeSet", value = "data.frame"): replaces slot data.

**intensity2GCplot** signature(x = "DataTreeSet"): creates a boxplot of probe intensities stratified by GC content.

**mm** signature(object = "DataTreeSet"): extracts the mismatch intensities.

**mmindex** signature(object = "DataTreeSet"): extracts (x,y)-coordinates and corresponding MM indices for all or selected unitIDs.

**ncols** signature(object = "DataTreeSet"): extracts the physical number of array columns from slot scheme.

**nrows** signature(object = "DataTreeSet"): extracts the physical number of array rows from slot scheme.

**pm** signature(object = "DataTreeSet"): extracts the perfect match intensities.

**pmindex** signature(object = "DataTreeSet"): extracts (x,y)-coordinates and corresponding PM indices for all or selected unitIDs.

**pmplot** signature(x = "DataTreeSet"): creates a barplot of mean perfect match and mismatch intensities.

**probesetID2unitID** signature(object = "DataTreeSet"): extracts all or selected probesetIDs from data.frame unitname of slot scheme with UnitName, i.e. probeset ID, as (row)names.

**probesetplot** signature(x = "DataTreeSet"): creates a line plot of probe intensities for a probeset.

**projectInfo** signature(object = "DataTreeSet"): extracts slot projectinfo.

**projectInfo<-** signature(object = "DataTreeSet", value = "ProjectInfo"): replaces slot projectinfo.

**rawCELName** signature(object = "DataTreeSet"): returns the name(s) of the imported raw CEL-files.

**removeBgrd** signature(object = "DataTreeSet"): replaces data.frame bgrd with an empty data.frame of dim(0,0).

**removeDataXY** signature(object = "DataTreeSet"): replaces data.frame data with an empty data.frame of dim(0,0).

**removeInten** signature(object = "DataTreeSet"): replaces data.frame data with an empty data.frame of dim(0,0).

**removeMask** signature(object = "DataTreeSet"): replaces data.frame mask from slot scheme with an empty data.frame of dim(0,0).

**removeProbeContentGC** signature(object = "DataTreeSet"): replaces data.frame probe with an empty data.frame of dim(0,0).

**removeUnitNames** signature(object = "DataTreeSet"): replaces data.frame unitname from slot scheme with an empty data.frame of dim(0,0).

**symbol2unitID** signature(object = "DataTreeSet"): extracts internal UNIT\_ID(s) for one or more gene symbols.

**transcriptID2unitID** signature(object = "DataTreeSet"): extracts all or selected transcriptIDs from data.frame unitname of slot scheme with UnitName, i.e. transcript ID, as (row)names.

**unitID2probesetID** signature(object = "DataTreeSet"): extracts all or selected unitIDs from data.frame unitname of slot scheme with UNIT\_ID as (row)names.

**symbol2unitID** signature(object = "DataTreeSet"): extracts gene symbols for one or more internal UNIT\_ID(s).

**unitID2transcriptID** signature(object = "DataTreeSet"): extracts all or selected unitIDs from data.frame unitname of slot scheme with UNIT\_ID as (row)names.

**validBgrd** signature(object = "DataTreeSet"): extracts the valid data from data.frame bgrd.

**validData** signature(object = "DataTreeSet"): extracts a subset of valid data from data.frame data.

**xpsBgCorrect** signature(object = "DataTreeSet"): applies background correction methods. See [bgcorrect](#).

**xpsDABGCall** signature(object = "DataTreeSet"): computes DABG call.

**xpsFIRMA** signature(object = "DataTreeSet"): computes FIRMA expression level and splice score.

**xpsINICall** signature(object = "DataTreeSet"): computes I/NI call.

**xpsMAS4** signature(object = "DataTreeSet"): computes MAS4 expression levels.

**xpsMAS5** signature(object = "DataTreeSet"): computes MAS5 expression levels.  
**xpsMAS5Call** signature(object = "DataTreeSet"): computes MAS5 detection call.  
**xpsNormalize** signature(object = "DataTreeSet"): applies normalization methods.  
**xpsPreprocess** signature(object = "DataTreeSet"): applies normalization methods.  
**xpsQualify** signature(object = "DataTreeSet"): applies quality control methods.  
**xpsQualityControl** signature(object = "DataTreeSet"): applies quality control methods.  
**xpsRMA** signature(object = "DataTreeSet"): computes RMA expression levels.  
**xpsSummarize** signature(object = "DataTreeSet"): applies summarization methods.

### Author(s)

Christian Stratowa

### See Also

related classes [ExprTreeSet](#), [CallTreeSet](#).

### Examples

```
showClass("DataTreeSet")
```

---

dfw

*Distribution Free Weighted Expression Measure*

---

### Description

This function converts a [DataTreeSet](#) into an [ExprTreeSet](#) using the Distribution Free Weighted Fold Change (DFW) method.

### Usage

```
dfw(xps.data,  
    filename = character(0),  
    filedir  = getwd(),  
    tmpdir   = "",  
    normalize = TRUE,  
    m        = 3,  
    n        = 1,  
    c        = 0.01,  
    option   = "transcript",  
    exonlevel = "",  
    xps.scheme = NULL,  
    add.data  = TRUE,  
    verbose   = TRUE)
```

## Arguments

xps.data	object of class <a href="#">DataTreeSet</a> .
filename	file name of ROOT data file.
filedir	system directory where ROOT data file should be stored.
tmpdir	optional temporary directory where temporary ROOT files should be stored.
normalize	logical. If TRUE normalize data using quantile normalization.
m	positive number as exponent of the weighted range WR.
n	positive number as exponent of the weighted standard deviation WSD.
c	scaling parameter.
option	option determining the grouping of probes for summarization, one of 'transcript', 'exon', 'probeset'; exon arrays only.
exonlevel	exon annotation level determining which probes should be used for summarization; exon/genome arrays only.
xps.scheme	optional alternative SchemeTreeSet.
add.data	logical. If TRUE expression data will be included as slot data.
verbose	logical, if TRUE print status information.

## Details

This function computes the DFW (Distribution Free Weighted Fold Change) expression measure described in Chen et al. for both expression arrays and exon arrays. For exon arrays it is necessary to supply the requested option and exonlevel.

Following options are valid for exon arrays:

transcript:	expression levels are computed for transcript clusters, i.e. probe sets containing the same 'transcript_cluster'.
exon:	expression levels are computed for exon clusters, i.e. probe sets containing the same 'exon_id', where each
probeset:	expression levels are computed for individual probe sets, i.e. for each 'probeset_id'.

Following exonlevel annotations are valid for exon arrays:

core:	probesets supported by RefSeq and full-length GenBank transcripts.
metacore:	core meta-probesets.
extended:	probesets with other cDNA support.
metaextended:	extended meta-probesets.
full:	probesets supported by gene predictions only.
metafull:	full meta-probesets.
affx:	standard AFFX controls.
all:	combination of above (including affx).

Following exonlevel annotations are valid for whole genome arrays:

core:	probesets with category 'unique', 'similar' and 'mixed'.
metacore:	probesets with category 'unique' only.
affx:	standard AFFX controls.
all:	combination of above (including affx).

Exon levels can also be combined, with following combinations being most useful:

exonlevel="metacore+affy":	core meta-probesets plus AFFX controls
exonlevel="core+extended":	probesets with cDNA support
exonlevel="core+extended+full":	supported plus predicted probesets

Exon level annotations are described in the Affymetrix whitepaper [exon\\_probeset\\_trans\\_clust\\_whitepaper.pdf](#): “Exon Probeset Annotations and Transcript Cluster Groupings”.

In order to use an alternative [SchemeTreeSet](#) set the corresponding SchemeSet `xps.scheme`.

## Value

An [ExprTreeSet](#)

## Note

The expression measure obtained with DFW is given in linear scale, analogously to the expression measures computed with [mas5](#) and [rma](#).

For the analysis of many exon arrays it may be better to define a `tmpdir`, since this will store only the results in the main file and not e.g. background and normalized intensities, and thus will reduce the file size of the main file. For quantile normalization memory should not be an issue, however DFW depends on RAM unless you are using a temporary file.

## Author(s)

Christian Stratowa

## References

Chen, Z., McGee M., Liu Q., and Scheuermann, R.H. (2007), A distribution free summarization method for Affymetrix GeneChip arrays. *Bioinformatics* 23(3):321-327

## See Also

[express](#)

## Examples

```
## first, load ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

data.dfw <- dfw(data.test3, "tmp_Test3DFW", verbose=FALSE)

## get data.frame
expr.dfw <- validData(data.dfw)
head(expr.dfw)
```

**Description**

This method initializes the Difference Filter.

The difference is the maximum value minus minimum value for each row of the expression dataframe divided by the mean value of each row.

The Difference Filter flags all rows with:  $\text{flag} = ((\text{max} - \text{min})/\text{mean} \geq \text{cutoff})$

*Usage*

```
diffFilter(object)
diffFilter(object, value)<-
```

**Arguments**

<code>object</code>	object of class <code>PreFilter</code> .
<code>value</code>	numeric vector <code>c(cutoff, trim, epsilon)</code> .

**Details**

The method `diffFilter` initializes the following parameters:

<code>cutoff</code> :	the cutoff level for the filter.
<code>trim</code> :	the trim value for trimmed mean (default is <code>trim=0</code> ).
<code>epsilon</code> :	value to replace mean (default is <code>epsilon=0.01</code> ):
	<code>epsilon &gt; 0</code> : replace <code>mean=0</code> with <code>epsilon</code> .
	<code>epsilon = 0</code> : always set <code>mean=1</code> .

Note, that for `epsilon = 0` the filter flags all rows with:  $(\text{max} - \text{min}) \geq \text{cutoff}$

**Value**

An initialized `PreFilter` object.

**Author(s)**

Christian Stratowa

**Examples**

```
prefltr <- PreFilter()
diffFilter(prefltr) <- c(2.2,0.0,0.01)
str(prefltr)
```

---

existsROOTFile                    *Test for Existing ROOT File*

---

**Description**

Test if a ROOT file does already exist.

**Usage**

```
existsROOTFile(filename, tmp.rm = TRUE)
```

**Arguments**

filename	name of ROOT file, including full path.
tmp.rm	logical, if TRUE then exclude filenames beginning with dQuote(tmp\_).

**Value**

Return TRUE if file filename is an already existing [ROOT](#) file.

**Note**

It is possible to create temporary [ROOT](#) files called “tmp” or with filename starting with “tmp\\_” which can be overwritten. Thus by default temporary files will not be recognized by existsROOTFile. If you want to recognize temporary files, set tmp.rm = TRUE.

**Author(s)**

Christian Stratowa

**See Also**

[isROOTFile](#)

**Examples**

```
existsROOTFile(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
```

---

exonLevel                            *Conversion of Parameter exonlevel to Integer*

---

**Description**

Conversion of parameter exonlevel to an integer vector.

**Usage**

```
exonLevel(exonlevel = "", chiptype = "GeneChip", as.sum = TRUE)
```



**Arguments**

exonlevel	exon annotation level determining which probes should be used for summarization; exon/genome arrays only.
chiptype	chip tpye, one of 'GeneChip', 'GenomeChip', 'ExonChip'.
as.sum	logical, if TRUE an integer vector of size three will be returned, if FALSE then the levels will be split into the basic integer representations.

**Details**

Conversion of parameter exonlevel to an integer; this function is a utility function, which is usually only used internally.

Following exonlevel annotations are valid for exon arrays:

core:	(=8192+1024) probesets supported by RefSeq and full-length GenBank transcripts.
metacore:	(=8192) core meta-probesets.
extended:	(=4096+512) probesets with other cDNA support.
metaextended:	(=4096) extended meta-probesets.
full:	(=2048+256) probesets supported by gene predictions only.
metafull:	(=2048) full meta-probesets.
ambiguous:	(=128) probesets that fall within multiple genes.
affx:	(=60) standard AFFX controls.
all:	(=16316) combination of above (including affx).

Following exonlevel annotations are valid for whole genome arrays:

core:	(=8192+1024) probesets with category 'unique', 'similar' and 'mixed'.
metacore:	(=8192) probesets with category 'unique' only.
affx:	(=60) standard AFFX controls.
all:	(=9276) combination of above (including affx).

Exon levels can also be combined, with following combinations being most useful:

exonlevel="metacore+affx":	core meta-probesets plus AFFX controls
exonlevel="core+extended":	probesets with cDNA support
exonlevel="core+extended+full":	supported plus predicted probesets

Exon level annotations are described in the Affymetrix whitepaper [exon\\_probeset\\_trans\\_clust\\_whitepaper.pdf](#): "Exon Probeset Annotations and Transcript Cluster Groupings".

Parameter exonlevel determines not only which probes are used for medianpolish, but also the probes used for background calculation and for quantile normalization. If you want to use separate probes for background calculation, quantile normalization and medianpolish summarization, you can pass a numeric vector containing three integer values corresponding to the respective exonlevel. These integers must be the sum of the integers shown above, e.g. you can use `exonlevel=c(16316,8252,8252)`, where `8252=8192+60` for "metacore+affx".

**Value**

an integer vector.

**Note**

The following exonlevels are unsupported:

control->bgp->genomic:	(=32768) genomic background probes.
control->bgp->antigenomic:	(=65536) antigenomic background probes.
normgene->intron:	(=131072) intronic controls.
normgene->exon:	(=262144) exonic controls.
rescue->FLmRNA->unmapped:	(=524288) unmapped mRNAs.

For whole genome arrays it is possible (but not recommended) to use all probesets by using `exonlevel=c(992316, 992316)`.  
 For exon arrays it is possible to use e.g. `exonlevel=c(1032124, 1032124, 631868)`.  
 However, please note that these settings are not recommended and not supported.

**Author(s)**

Christian Stratowa

**See Also**

[rma](#), [mas5](#)

**Examples**

```
exonLevel("core", "GenomeChip")
exonLevel("all", "GenomeChip")
exonLevel("core+extended+full", "ExonChip")
exonLevel("core+extended+full", "ExonChip", as.sum=FALSE)
exonLevel(c(16316,8252,8252), "ExonChip")
```

---

export

*Export data as text files*

---

**Description**

Export data from classes [SchemeTreeSet](#), [DataTreeSet](#), [ExprTreeSet](#), or [CallTreeSet](#) to outfile.

**Usage**

```
export.scheme(xps.scheme, treetype = character(0), varlist = "*", outfile = character(0), sep = "\n")
export.data(xps.data, treename = "*", treetype = "cel", varlist = "*", outfile = character(0), sep = "\n")
export.expr(xps.expr, treename = "*", treetype = character(0), varlist = "*", outfile = character(0), sep = "\n")
export.call(xps.call, treename = "*", treetype = character(0), varlist = "*", outfile = character(0), sep = "\n")
export(object, ...)
```

**Arguments**

xps.scheme	an object of type <a href="#">SchemeTreeSet</a> .
xps.data	an object of type <a href="#">DataTreeSet</a> .
xps.expr	an object of type <a href="#">ExprTreeSet</a> .
xps.call	an object of type <a href="#">CallTreeSet</a> .
treename	vector of tree names to export.
treetype	type of tree(s) to export, see <a href="#">validTreetype</a>
varlist	names of tree leaves to export
outfile	name of output file.
sep	column separator
as.dataframe	if TRUE a data.frame will be returned.
verbose	logical, if TRUE print status information.
object	object of class <a href="#">DataTreeSet</a> .
...	arguments <a href="#">treenames</a> , <a href="#">treetype</a> , <a href="#">varlist</a> , <a href="#">outfile</a> , <a href="#">sep</a> , <a href="#">as.dataframe</a> .

**Details**

Export data from classes [SchemeTreeSet](#), [DataTreeSet](#), [ExprTreeSet](#), or [CallTreeSet](#) to outfile.

Parameter `varlist` lists the parameters to export:

- parameters are separated by ":", e.g. `varlist="fInten:fStdev"`.
- for `varlist="*"` all valid parameters will be exported.

For class [DataTreeSet](#) the following `varlist` parameters are valid:

fInten:	intensities from e.g. <code>tree.cel</code> .
fStdev:	standard deviation from e.g. <code>tree.cel</code> .
fNPixels:	number of pixels from e.g. <code>tree.cel</code> .
fBg:	background values (background trees only).

For classes [ExprTreeSet](#) and [CallTreeSet](#) `varlist` can contain annotation parameters and parameters of the resulting data.

Following `varlist` annotation parameters are valid:

fUnitName:	unit name (probeset ID).
fTranscriptID:	transcript_id (probeset ID).
fName:	gene name.
fSymbol:	gene symbol.
fAccession:	mRNA accession such as Refseq ID.
fEntrezID:	entrez ID.
fChromosome:	chromosome.
fStart:	start position.
fStop:	stop position.
fStrand:	strand on chromosome.
fCytoBand:	cytoband.

Following `varlist` parameters are valid for [ExprTreeSet](#):

fLevel:	expression level.
---------	-------------------

fStdev: standard deviation.  
fNPairs: number of pairs.

Following varlist parameters are valid for CallTreeSet:

fCall: detection call.  
fPValue: detection p-value.

Following varlist parameters are valid for QualTreeSet:

fLevel: expression level.  
fStderr: standard error.  
fNUSE: normalized unscaled standard error.  
fRLE: relative log expression value.

An example: `varlist="fUnitName:fName:fSymbol:fLevel:fStdev:fEntrezID"`

`export` is a generic method to export data from [ROOT](#) trees as text file.

### Value

If `as.dataframe` is `TRUE`, the data will be imported into the current R session as `data.frame`. Otherwise, `NULL` will be returned.

### Author(s)

Christian Stratowa

### See Also

[export-methods](#)

### Examples

```
## load existing ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

## export as table only
export(scheme.test3, treetype="idx", outfile="Test3_idx.txt", verbose=FALSE)

## export as table and import as data.frame
ann <- export.scheme(scheme.test3, treetype="ann", outfile="Test3_ann.txt", as.dataframe=TRUE, verbose=FALSE)
head(ann)
data <- export.data(data.test3, outfile="Test3_cel.txt", as.dataframe=TRUE, verbose=FALSE)
head(data)
```

---

export.filter	<i>Export filter data as text files</i>
---------------	---

---

## Description

Export data from classes [FilterTreeSet](#) or [AnalysisTreeSet](#) to outfile.

## Usage

```
export.filter(xps.fltr, treename = "*", treetype = character(0), varlist = "*", outfile = character(0))
```

## Arguments

xps.fltr	an object of type <a href="#">FilterTreeSet</a> or <a href="#">AnalysisTreeSet</a> .
treename	tree name to export.
treetype	type of tree(s) to export, 'pfr', 'ufr' or 'stt'.
varlist	names of tree leaves to export.
outfile	name of output file.
sep	column separator
as.dataframe	if TRUE a data.frame will be returned.
verbose	logical, if TRUE print status information.

## Details

Export data from classes [FilterTreeSet](#), or [AnalysisTreeSet](#) to outfile.

Parameter `varlist` lists the parameters to export:

- parameters are separated by ":", e.g. `varlist="fUnitName:fFlag"`.
- for `varlist="*"` all valid parameters will be exported.

For class [FilterTreeSet](#) the following `varlist` parameters are valid:

fUnitName:	unit name (probeset ID).
fFlag:	mask.

For class [AnalysisTreeSet](#) `varlist` can contain annotation parameters and parameters of the resulting data.

Following `varlist` annotation parameters are valid:

fUnitName:	unit name (probeset ID).
fTranscriptID:	transcript\_id (probeset ID).
fName:	gene name.
fSymbol:	gene symbol.
fAccession:	mRNA accession such as Refseq ID.
fEntrezID:	entrez ID.
fChromosome:	chromosome.
fStart:	start position.
fStop:	stop position.
fStrand:	strand on chromosome.
fCytoBand:	cytoband.

For class `AnalysisTreeSet` the following `varlist` parameters are valid:

```

mn1:  mean of group 1.
mn2:  mean of group 2.
fc:   fold-change  $fc=mn2/mn1$ .
se:   standard error.
df:   degree of freedom.
stat: t-statistic.
pval: p-value.
nper: number of permutations.
pcha: p-chance.
padj: adjusted p-value.
flag: flag.
mask: only rows with flag=1 will be exported.

```

### Value

If `as.dataframe` is `TRUE`, the data will be imported into the current R session as `data.frame`. Otherwise, `NULL` will be returned.

### Author(s)

Christian Stratowa

### See Also

[export-methods](#)

---

`export.root`

*Export data from ROOT file*

---

### Description

Export data as text files directly from a [ROOT](#) file.

### Usage

```
export.root(datafile = character(0), schemefile = character(0), treeset = character(0), treename =
```

### Arguments

<code>datafile</code>	name of ROOT data file including full path
<code>schemefile</code>	name of ROOT scheme file including full path
<code>treeset</code>	name of subdirectory in ROOT file where trees are stored
<code>treename</code>	name of ROOT tree to export.
<code>treetype</code>	type of tree(s) to export, see <a href="#">validTreetype</a> .
<code>varlist</code>	names of tree leaves to export.
<code>outfile</code>	name of output file.
<code>sep</code>	column separator
<code>as.dataframe</code>	if <code>TRUE</code> a <code>data.frame</code> will be returned.
<code>verbose</code>	logical, if <code>TRUE</code> print status information.

**Details**

Export data as text files directly from a [ROOT](#) file.

**Value**

If `as.dataframe` is `TRUE`, the data will be imported into the current R session as `data.frame`. Otherwise, `NULL` will be returned.

**Author(s)**

Christian Stratowa

**See Also**

[export](#), [export-methods](#)

**Examples**

```
## export data directly from root file
schemefile <- paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/")
datafile   <- paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/")
data <- export.root(datafile, schemefile, "DataSet", "*", "cel", "*", "DataOutFile.txt", as.dataframe = TRUE,
head(data)
```

---

express

*Compute expression levels from raw data*

---

**Description**

This function allows to combine different algorithms to compute expression levels, or to return the result for different algorithms only.

**Usage**

```
express(xps.data,
        filename = character(),
        filedir = getwd(),
        tmpdir = "",
        update = FALSE,
        # background correction
        bgcorrect.method = NULL,
        bgcorrect.select = character(),
        bgcorrect.option = character(),
        bgcorrect.params = list(),
        # normalization
        normalize.method = NULL,
        normalize.select = character(),
        normalize.option = character(),
        normalize.logbase = character(),
        normalize.params = list(),
        # expression values
        summarize.method = NULL,
```

```

summarize.select = character(),
summarize.option = character(),
summarize.logbase = character(),
summarize.params = list(),
# reference values
reference.index = 0,
reference.method = "mean",
reference.params = list(),
# misc.
exonlevel = "",
xps.scheme = NULL,
add.data = TRUE,
bufsize = 32000,
verbose = TRUE)

```

```
xpsPreprocess(object, ...)
```

### Arguments

xps.data	object of class DataTreeSet.
filename	file name of ROOT data file.
filedir	system directory where ROOT data file should be stored.
tmpdir	optional temporary directory where temporary ROOT files should be stored.
update	logical. If TRUE the existing ROOT data file filename will be updated.
bgcorrect.method	background method to use.
bgcorrect.select	type of probes to select for background correction.
bgcorrect.option	type of background correction to use.
bgcorrect.params	vector of parameters for background method.
normalize.method	normalization method to use.
normalize.select	type of probes to select for normalization.
normalize.option	normalization option.
normalize.logbase	logarithm base as character, one of '0', 'log', 'log2', 'log10'.
normalize.params	vector of parameters for normalization method.
summarize.method	summarization method to use.
summarize.select	type of probes to select for summarization.
summarize.option	option determining the grouping of probes for summarization, one of 'transcript', 'exon', 'probeset'; exon arrays only.



<code>summarize.logbase</code>	logarithm base as character, one of '0', 'log', 'log2', 'log10'.
<code>summarize.params</code>	vector of parameters for summarization method.
<code>reference.index</code>	index of reference tree to use, or 0.
<code>reference.method</code>	for <code>refindex=0</code> , either trimmed mean or median of trees.
<code>reference.params</code>	vector of parameters for reference method.
<code>exonlevel</code>	exon annotation level determining which probes should be used for summarization; exon/genome arrays only.
<code>xps.scheme</code>	optional alternative <code>SchemeSet</code> .
<code>add.data</code>	logical. If TRUE expression data will be included as slot data.
<code>bufsize</code>	integer which sets the buffer size of the tree branch baskets (default is 32000).
<code>verbose</code>	logical, if TRUE print status information.
<code>object</code>	object of class <code>DataTreeSet</code> .
<code>...</code>	the arguments described above.

## Details

This function allows to combine different algorithms to compute expression levels, or to return the result for different algorithms only.

Please have a look at vignette “`xpsPreprocess.pdf`” for details on how to use function `express`.

`xpsPreprocess` is the `DataTreeSet` method called by function `express`, containing the same parameters.

## Value

An object of type `DataTreeSet` or `ExprTreeSet`.

## Author(s)

Christian Stratowa

## See Also

[bgcorrect](#), [normalize](#), [summarize](#)

## Examples

```
## load existing ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

## compute rma with a single call to express()
expr.rma <- express(data.test3, "tmp_Test3Exprs", filedir=getwd(), tmpdir="", update=FALSE,
  bgcorrect.method="rma", bgcorrect.select="none", bgcorrect.option="pmonly:epanechnikov", bgcorrect.p
  normalize.method="quantile", normalize.select="pmonly", normalize.option="transcript:together:none"
  summarize.method="medianpolish", summarize.select="pmonly", summarize.option="transcript", summarize
  verbose=FALSE)
```

```

## get expression data.frame
expr <- exprs(expr.rma)
head(expr)

## plot expression levels
if (interactive()) {
  boxplot(expr.rma)
  boxplot(log2(expr[,3:6]))
}

## Not run:
## examples using Affymetrix human tissue dataset (see also xps/examples/script4exon.R)

## example - exon array, e.g. HuEx-1_0-st-v2:
scmdir <- "/Volumes/GigaDrive/CRAN/Workspaces/Schemes"
datdir <- "/Volumes/GigaDrive/CRAN/Workspaces/ROOTData"
scheme.exon <- root.scheme(paste(scmdir, "Scheme_HuEx10stv2r2_na25.root", sep="/"))
data.exon <- root.data(scheme.exon, paste(datdir, "HuTissuesExon_cel.root", sep="/"))

workdir <- "/Volumes/GigaDrive/CRAN/Workspaces/Exon/hutissues/exon"
expr.rma <- express(data.exon, "HuExonExprs", filedir=workdir, tmpdir="", update=F,
  bgcorrect.method="rma", bgcorrect.select="antigenomic", bgcorrect.option="pmonly:epanechnikov", bgco
  normalize.method="quantile", normalize.select="pmonly", normalize.option="transcript:together:none"
  summarize.method="medianpolish", summarize.select="pmonly", summarize.option="transcript", summarize
  exonlevel="metacore+affx")

## End(Not run)

```

---

 exprs-methods

*Get/Set Expression Values*


---

## Description

Get/set expression values from/for class `ExprTreeSet`.

*Usage*

```

exprs(object)
exprs(object, treenames = NULL) <- value

```

## Arguments

<code>object</code>	object of class <code>ExprTreeSet</code> .
<code>treenames</code>	character vector containing optional tree names to be used as subset.
<code>value</code>	data.frame containing expression values.

## Details

Get the expression values from slot data or set slot data to value.

Method `exprs` returns the expression values from slot data as data.frame, while replacement method `exprs<-` allows to replace slot data with a data.frame.

In order to create an `ExprTreeSet` containing only a subset of slot data, first export slot data using method `exprs`, create a character vector containing only `treenames` to be used in the subset, and

then use replacement method `exprs<-` to replace slot data with the subset. Slots `treenames` and `numtrees` will be updated automatically.

Note: When creating character vector `treenames` it is sufficient to use the name part of the tree name w/o the extension.

Note: If you do not want to replace your current object, create first a copy of type `ExprTreeSet` by simply writing `newobj <- oldobj`, and use `newobj` for replacement. This is important since `exprs<-` does also update slots `treenames` and `numtrees` as already mentioned.

### Author(s)

Christian Stratowa

### See Also

[pvalData](#), [presCall](#)

### Examples

```
## Not run:
## load ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

## create an ExprTreeSet
data.rma <- rma(data.test3, "tmp_TestRMA", tmpdir="", background="pmonly", normalize=TRUE, verbose=FALSE)

## get expression values
value <- exprs(data.rma)

## selected treenames only
treenames <- c("TestA2", "TestB1")

## make a copy of your object if you do not want to replace it
subset.rma <- data.rma

## replace slot data with subset
exprs(subset.rma, treenames) <- value
str(subset.rma)

## End(Not run)
```

---

ExprTreeSet-class

*Class ExprTreeSet*

---

### Description

This class provides the link to the [ROOT](#) expression file and the [ROOT](#) trees contained therein. It extends class [ProcesSet](#).

### Objects from the Class

Objects are created using functions [express](#), [summarize](#) or [normalize](#), or the specialized functions [rma](#), [mas5](#) or [mas4](#).

**Slots**

- exprtype**: Object of class "character" representing the expression type, i.e. 'rma', 'mas5', 'mas4' or 'custom'.
- normtype**: Object of class "character" representing the normalization type, i.e. 'mean', 'median', 'lowess', 'supsmu'.
- scheme**: Object of class "SchemeTreeSet" providing access to [ROOT](#) scheme file.
- data**: Object of class "data.frame". The data.frame can contain the data (e.g. expression levels) stored in [ROOT](#) data trees.
- params**: Object of class "list" representing relevant parameters.
- setname**: Object of class "character" representing the name to the [ROOT](#) file subdirectory where the [ROOT](#) data trees are stored, usually 'PreprocesSet'.
- settype**: Object of class "character" describing the type of treeset stored in setname, usually 'preprocess'.
- rootfile**: Object of class "character" representing the name of the [ROOT](#) data file, including full path.
- filedir**: Object of class "character" describing the full path to the system directory where rootfile is stored.
- numtrees**: Object of class "numeric" representing the number of [ROOT](#) trees stored in subdirectory setname.
- treenames**: Object of class "list" representing the names of the [ROOT](#) trees stored in subdirectory setname.

**Extends**

Class "[ProcesSet](#)", directly. Class "[TreeSet](#)", by class "[ProcesSet](#)", distance 2.

**Methods**

- attachExpr** signature(object = "ExprTreeSet"): exports expression trees from [ROOT](#) expression file and saves as data.frame data.
- corplot** signature(x = "ExprTreeSet"): creates a correlation heat map.
- exprType** signature(object = "ExprTreeSet"): extracts slot exprtype.
- exprType<-** signature(object = "ExprTreeSet", value = "character"): replaces slot exprtype.
- exprs** signature(object = "ExprTreeSet"): extracts the expression data.frame.
- exprs<-** signature(object = "ExprTreeSet", value = "data.frame"): replaces the expression data.frame.
- madplot** signature(x = "ExprTreeSet"): creates a false color display of between arrays distances.
- mvaplot** signature(x = "ExprTreeSet"): creates an MvA-plot.
- normType** signature(object = "ExprTreeSet"): extracts slot normtype.
- normType<-** signature(object = "ExprTreeSet", value = "character"): replaces slot normtype.
- nuseplot** signature(x = "ExprTreeSet"): creates a NUSE-plot.
- pcaplot** signature(x = "ExprTreeSet"): plots first two principal components of PCA.
- rleplot** signature(x = "ExprTreeSet"): creates a RLE-plot.

**removeExpr** signature(object = "ExprTreeSet"): replaces data.frame data with an empty data.frame of dim(0,0).

**se.exprs** signature(object = "ExprTreeSet"): extracts the standard deviation data.frame.

**validExpr** signature(object = "ExprTreeSet"): extracts a subset of columns from data.frame data.

**validSE** signature(object = "ExprTreeSet"): extracts data columns from data.frame se.exprs.

**xpsNormalize** signature(object = "ExprTreeSet"): applies normalization methods.

**xpsPreFilter** signature(object = "ExprTreeSet"): applies prefiltering methods.

**xpsUniFilter** signature(object = "ExprTreeSet"): applies unfiltering methods.

### Author(s)

Christian Stratowa

### See Also

related classes [DataTreeSet](#), [CallTreeSet](#), [QualTreeSet](#).

### Examples

```
showClass("ExprTreeSet")
```

---

extenPart

*Get Extension of Tree Names*

---

### Description

Get the extension(s) of (tree) names.

### Usage

```
extenPart(names, as.unique=TRUE)
```

### Arguments

names                vector of names.

as.unique            if TRUE return only unique extensions.

### Details

Extracts the extension part of names, e.g. of tree names of treename.treetype stored in a [ROOT](#) file.

### Value

A vector of (unique) extensions.

### Author(s)

Christian Stratowa

**See Also**[namePart](#)**Examples**

```
names <- c("TestA1.int", "TestA2.int")
extenPart(names)
extenPart(names, as.unique=FALSE)
```

farms

*Factor Analysis for Robust Microarray Summarization Expression Measure*

**Description**

This function converts a [DataTreeSet](#) into an [ExprTreeSet](#) using the Factor Analysis for Robust Microarray Summarization (FARMS) method.

**Usage**

```
farms(xps.data,
      filename = character(0),
      filedir  = getwd(),
      tmpdir   = "",
      normalize = TRUE,
      weight   = 0.5,
      mu       = 0.0,
      scale    = 1.0,
      tol      = 0.00001,
      cyc      = 100,
      weighted = TRUE,
      version  = "1.3.1",
      option   = "transcript",
      exonlevel = "",
      xps.scheme = NULL,
      add.data  = TRUE,
      verbose  = TRUE)
```

**Arguments**

xps.data	object of class <a href="#">DataTreeSet</a> .
filename	file name of ROOT data file.
filedir	system directory where ROOT data file should be stored.
tmpdir	optional temporary directory where temporary ROOT files should be stored.
normalize	logical. If TRUE normalize data using quantile normalization.
weight	hyperparameter, usually set to 0.5 for version="1.3.1" and to 8.0 for version="1.3.0".
mu	hyperparameter allowing to correct for potential bias.
scale	scaling parameter, usually set to 1.0 for version="1.3.1" and to 2.0 for version="1.3.0".
tol	termination tolerance for EM algorithm.

cyc	maximum number of cycles of EM algorithm.
weighted	logical, used only with <code>version="1.3.1"</code> . Default is TRUE.
version	version of original farms package. Currently, <code>version="1.3.1"</code> and <code>version="1.3.0"</code> are implemented. Default is <code>version="1.3.1"</code> .
option	option determining the grouping of probes for summarization, one of 'transcript', 'exon', 'probeset'; exon arrays only.
exonlevel	exon annotation level determining which probes should be used for summarization; exon/genome arrays only.
xps.scheme	optional alternative SchemeTreeSet.
add.data	logical. If TRUE expression data will be included as slot data.
verbose	logical, if TRUE print status information.

## Details

This function computes the FARMS (Factor Analysis for Robust Microarray Summarization) expression measure described in Hochreiter et al. for both expression arrays and exon arrays.

Parameter `version` currently allows the user to choose between the original implementation of FARMS as implemented in package 'farms\_1.3.0' or enhanced FARMS as implemented in package 'farms\_1.3.1'. By default `version="1.3.1"` is used.

Parameter `weight` is a hyperparameter which determines the influence of the prior. For `version="1.3.1"` the value in the range of [0,1].

Parameter `mu` is a hyperparameter which allows to quantify different aspects of potential prior knowledge. Values near zero assume that most genes do not contain a signal and introduce a bias for loading matrix elements near zero.

Parameter `weighted` is a logical and indicates whether a weighted mean or a least square fit is used to summarize the loading matrix. It is applicable only to `version="1.3.1"`.

For exon arrays it is necessary to supply the requested `option` and `exonlevel`.

Following options are valid for exon arrays:

<code>transcript:</code>	expression levels are computed for transcript clusters, i.e. probe sets containing the same 'transcript_cluster'
<code>exon:</code>	expression levels are computed for exon clusters, i.e. probe sets containing the same 'exon_id', where each
<code>probeset:</code>	expression levels are computed for individual probe sets, i.e. for each 'probeset_id'.

Following `exonlevel` annotations are valid for exon arrays:

<code>core:</code>	probesets supported by RefSeq and full-length GenBank transcripts.
<code>metacore:</code>	core meta-probesets.
<code>extended:</code>	probesets with other cDNA support.
<code>metaextended:</code>	extended meta-probesets.
<code>full:</code>	probesets supported by gene predictions only.
<code>metafull:</code>	full meta-probesets.
<code>affx:</code>	standard AFFX controls.
<code>all:</code>	combination of above (including <code>affx</code> ).

Following `exonlevel` annotations are valid for whole genome arrays:

<code>core:</code>	probesets with category 'unique', 'similar' and 'mixed'.
--------------------	--

metacore: probesets with category 'unique' only.  
 affx: standard AFFX controls.  
 all: combination of above (including affx).

Exon levels can also be combined, with following combinations being most useful:

exonlevel="metacore+affy": core meta-probesets plus AFFX controls  
 exonlevel="core+extended": probesets with cDNA support  
 exonlevel="core+extended+full": supported plus predicted probesets

Exon level annotations are described in the Affymetrix whitepaper [exon\\_probeset\\_trans\\_clust\\_whitepaper.pdf](#):  
 "Exon Probeset Annotations and Transcript Cluster Groupings".

In order to use an alternative [SchemeTreeSet](#) set the corresponding SchemeSet `xps.scheme`.

### Value

An [ExprTreeSet](#)

### Note

The expression measure obtained with FARMS is given in linear scale, analogously to the expression measures computed with [mas5](#) and [rma](#).

For the analysis of many exon arrays it may be better to define a `tmpdir`, since this will store only the results in the main file and not e.g. background and normalized intensities, and thus will reduce the file size of the main file. For quantile normalization memory should not be an issue, however DFW depends on RAM unless you are using a temporary file.

### Author(s)

Christian Stratowa

### References

Hochreiter, S., Clevert D.-A., and Obermayer, K. (2006), A new summarization method for Affymetrix probe level data. *Bioinformatics* 22(8):943-949

### See Also

[express](#)

### Examples

```
## first, load ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

data.farms <- farms(data.test3, "tmp_Test3FARMS", verbose=FALSE)

## get data.frame
expr.farms <- validData(data.farms)
head(expr.farms)
```



---

fcFilter-methods      *Fold-Change Filter*

---

### Description

This method initializes the Fold-Change Filter.  
The fold-change is determined by the mean value of group 2 divided by the mean value of group 1.  
The Fold-Change Filter flags all rows with: `flag = (fc >= cutoff)`

#### Usage

```
fcFilter(object)
fcFilter(object, value)<-
```

### Arguments

object	object of class <code>UniFilter</code> .
value	numeric vector <code>c(cutoff, direction)</code>

### Details

The method `fcFilter` initializes the following parameters:

cutoff:	the cutoff level for the filter.
direction:	<code>direction="both"</code> (default): select up and downregulated genes. <code>direction="up"</code> : select upregulated genes only. <code>direction="down"</code> : select downregulated genes only.

### Value

An initialized `UniFilter` object.

### Author(s)

Christian Stratowa

### Examples

```
unifltr <- UniFilter()
fcFilter(unifltr) <- c(1.5,"both")
str(unifltr)
```

---

Filter-class      *Base Class Filter*

---

### Description

Base class for classes `PreFilter` and `UniFilter`.

### Slots

`numfilters`: Object of class "numeric" giving the number of filters applied.

**Methods**

**numberFilters** signature(object = "Filter"): number of filters applied.

**Author(s)**

Christian Stratowa

**See Also**

related classes [PreFilter](#), [UniFilter](#).

**Examples**

```
showClass("Filter")
```

---

FilterTreeSet-class    *Class FilterTreeSet*

---

**Description**

This class provides the link to the [ROOT](#) filter file and the [ROOT](#) trees contained therein. It extends class [ProcesSet](#).

**Objects from the Class**

Objects are currently created using function [prefilter](#).

**Slots**

**filter:** Object of class "Filter" currently providing access to the [PreFilter](#) settings.

**exprset:** Object of class "ExprTreeSet" providing direct access to the [ExprTreeSet](#) used for filtering.

**callset:** Object of class "CallTreeSet" providing direct access to the optional [CallTreeSet](#) used for filtering.

**scheme:** Object of class "SchemeTreeSet" providing access to [ROOT](#) scheme file.

**data:** Object of class "data.frame". The data.frame contains the data of the filter stored in [ROOT](#) filter trees.

**params:** Object of class "list" representing relevant parameters.

**setname:** Object of class "character" representing the name to the [ROOT](#) file subdirectory where the [ROOT](#) trees are stored, currently 'PreFilterSet'.

**settype:** Object of class "character" describing the type of treeset stored in setname, currently 'prefilter'.

**rootfile:** Object of class "character" representing the name of the [ROOT](#) file, including full path.

**filedir:** Object of class "character" describing the full path to the system directory where rootfile is stored.

**numtrees:** Object of class "numeric" representing the number of [ROOT](#) trees stored in subdirectory setname.

**treenames:** Object of class "list" representing the names of the [ROOT](#) trees stored in subdirectory setname.

## Extends

Class "[ProcesSet](#)", directly. Class "[TreeSet](#)", by class "ProcesSet", distance 2.

## Methods

**callTreeset** signature(object = "FilterTreeSet"): extracts slot callset.

**exprTreeset** signature(object = "FilterTreeSet"): extracts slot exprset.

**getTreeData** signature(object = "FilterTreeSet"): exports tree data and returns a data.frame.

**validData** signature(object = "FilterTreeSet"): extracts data.frame data.

## Author(s)

Christian Stratowa

## See Also

related classes [AnalysisTreeSet](#).

## Examples

```
showClass("FilterTreeSet")
```

---

firma

*Finding Isoforms using Robust Multichip Analysis*

---

## Description

This function converts a [DataTreeSet](#) for exon arrays into an [ExprTreeSet](#) using the Finding Isoforms using Robust Multichip Analysis (FIRMA).

## Usage

```
firma(xps.data,  
      filename = character(0),  
      filedir  = getwd(),  
      tmpdir   = "",  
      background = "antigenomic",  
      normalize = TRUE,  
      option   = "probeset",  
      exonlevel = "metacore",  
      method   = "mdp",  
      params   = list(16384, 0.0, 1.0, 10, 0.01, 1.0),  
      xps.scheme = NULL,  
      add.data  = TRUE,  
      verbose  = TRUE)
```

```
xpsFIRMA(object, ...)
```

## Arguments

xps.data	object of class <code>DataTreeSet</code> .
filename	file name of ROOT data file.
filedir	system directory where ROOT data file should be stored.
tmpdir	optional temporary directory where temporary ROOT files should be stored.
background	probes used to compute background, one of 'genomic', 'antigenomic'
normalize	logical. If TRUE normalize data using quantile normalization.
option	option determining the grouping of probes for summarization, one of 'exon', 'probeset'.
exonlevel	exon annotation level determining which probes should be used for summarization.
method	method to be used for summarization, currently 'mdp'.
params	list of (default) parameters for rma.
xps.scheme	optional alternative SchemeTreeSet.
add.data	logical. If TRUE expression data will be included as slot data.
verbose	logical, if TRUE print status information.
object	object of class <code>DataTreeSet</code> .
...	the arguments described above.

## Details

This function computes FIRMA (Finding Isoforms using Robust Multichip Analysis) for detecting differential alternative splicing for exon arrays, as described in Purdom et al.

Following options are valid for exon arrays:

probeset: expression levels are computed for individual probe sets, i.e. for each 'probeset\_id'.

exon: expression levels are computed for exon clusters, i.e. probe sets containing the same 'exon\_id', where each e

Following exonlevel annotations are valid for exon arrays:

core:	probesets supported by RefSeq and full-length GenBank transcripts.
metacore:	core meta-probesets.
extended:	probesets with other cDNA support.
metaextended:	extended meta-probesets.
full:	probesets supported by gene predictions only.
metafull:	full meta-probesets.
ambiguous:	ambiguous probesets only.
affx:	standard AFFX controls.
all:	combination of above (including affx).

Exon levels can also be combined, with following combinations being most useful:

exonlevel="metacore+affx":	core meta-probesets plus AFFX controls
exonlevel="core+extended":	probesets with cDNA support
exonlevel="core+extended+full":	supported plus predicted probesets

Exon level annotations are described in the Affymetrix whitepaper `exon_probeset_trans_clust_whitepaper.pdf`: “Exon Probeset Annotations and Transcript Cluster Groupings”.

Method `xpsFIRMA` is the `DataTreeSet` method called by function `firma`, containing the same parameters.

## Value

An `ExprTreeSet`

## Note

In contrary to other implementations of (FI)RMA the expression measure of FIRMA is given in linear scale, analogously to the expression measures computed with `mas5` and `mas4`.

Please note that the current implementation of FIRMA is based on median-polish only, see: <http://www.aroma-project.org/node/81>

Please note that the default settings of `params` gives results which are identical to the results obtained with APT (Affymetrix Power Tools) and with package `affy_1.14.2` or earlier. If you want to obtain results which are identical to the results obtained with `affy_1.16.0` or later then you need to set `params = list(16384, 0.0, 0.4, 10, 0.01, 1.0)`.

By setting parameter `background="none"` it is possible to skip background correction .

For the analysis of many exon arrays it may be better to define a `tmpdir`, since this will store only the results in the main file and not e.g. background and normalized intensities, and thus will reduce the file size of the main file. For quantile normalization memory should not be an issue, however medianpolish depends on RAM unless you are using a temporary file.

Parameter `exonlevel` determines not only which probes are used for medianpolish, but also the probes used for background calculation and for quantile normalization. If you want to use separate probes for background calculation, quantile normalization and medianpolish summarization, you can pass a numeric vector containing three integer values corresponding to the respective `exonlevel`, e.g. you can use `exonlevel=c(16316,8252,8252)`, see function `exonLevel` for more details.

## Author(s)

Christian Stratowa

## References

Purdum, E., Simpson K.M., Robinson M.D., Conboy J.G., Lapuk A.V. and Speed, T.P. (2008), FIRMA: a method for detection of alternative splicing from exon array data. *Bioinformatics* 24(15):1707-1714

## Examples

```
## Not run:
## load ROOT scheme file
scmdir <- "/Volumes/GigaDrive/CRAN/Workspaces/Schemes"
scheme.exon <- root.scheme(paste(scmdir, "Scheme_HuEx10stv2r2_na27.root", sep="/"))

## load subset of ROOT data file
datdir <- "/Volumes/GigaDrive/CRAN/Workspaces/ROOTData"
subnames <- c("HeartA", "HeartB", "HeartC", "MuscleA", "MuscleB", "MuscleC")
sub.exon <- root.data(scheme.exon, rootFile(data.exon), celnames=subnames)
```

```

## firma
outdir <- getwd()
sub.firma.ps <- firma(sub.exon,"HeartMuscleFIRMAcorePS",filedir=outdir,tmpdir="",background="antigenomic",
                    normalize=TRUE,option="probeset",exonlevel="core")

## get transcript expression levels for all transcripts or transcript=2429277
expr.firma <- firma.expr(sub.firma.ps, probeset=NULL, option="transcript")
expr.firma <- firma.expr(sub.firma.ps, probeset=2429277, option="transcript")

## get probeset expression levels for all probeset or probeset=2429278 or transcript=2429277
expr.firma <- firma.expr(sub.firma.ps, probeset=NULL, option="probeset")
expr.firma <- firma.expr(sub.firma.ps, probeset=2429278, option="probeset")
expr.firma <- firma.expr(sub.firma.ps, probeset=2429277, option="probeset")

## get probeset splice scores for all probeset or probeset=2429278 or transcript=2429277
score.firma <- firma.score(sub.firma.ps, probeset=NULL, option="probeset")
score.firma <- firma.score(sub.firma.ps, probeset=2429278, option="probeset")
score.firma <- firma.score(sub.firma.ps, probeset=2429277, option="probeset")

## different plots
boxplot(sub.firma.ps, which="UnitName:LEVEL_PS")
boxplot(sub.firma.ps, which="UnitName:LEVEL_TS")

hist(sub.firma.ps, which="UnitName:LEVEL_PS")
hist(sub.firma.ps, which="UnitName:LEVEL_TS")

rleplot(sub.firma.ps, which="UnitName:LEVEL_PS")
rleplot(sub.firma.ps, which="UnitName:LEVEL_TS")

nuseplot(sub.firma.ps, which="UnitName:STDEV_PS")
nuseplot(sub.firma.ps, which="UnitName:STDEV_TS")

## End(Not run)

```

---

 firma.expr

*Get Expression Levels from FIRMA*


---

## Description

Extracts FIRMA expression levels from data.frame data.

## Usage

```

firma.expr(xps.data,
           probeset = NULL,
           option   = "probeset")

```

## Arguments

xps.data	object of class <a href="#">ExprTreeSet</a> .
probeset	transcriptID or probesetID or NULL.
option	option determining the probeset type for which to extract expression levels, one of 'transcript', 'probeset', 'exon'.

## Details

Function `firma.expr` returns the expression levels from slot data for a given probeset, or for all probesets or transcripts in case of `probeset=NULL`. Row names will be the Affymetrix transcriptIDs, probesetIDs or exonIDs, respectively, dependent on the selected option.

## Value

A `data.frame`.

## Note

For option="probeset" parameter probeset should usually be the transcriptID in order to get the expression levels for all probesetIDs of the corresponding transcriptID.

## Author(s)

Christian Stratowa

## See Also

[firma](#)

## Examples

```
## Not run:
## get transcript expression levels for all transcripts or for transcript=2429277
expr.firma <- firma.expr(sub.firma.ps, probeset=NULL, option="transcript")
expr.firma <- firma.expr(sub.firma.ps, probeset=2429277, option="transcript")

## get probeset expression levels for all probeset or for probeset=2429278
expr.firma <- firma.expr(sub.firma.ps, probeset=NULL, option="probeset")
expr.firma <- firma.expr(sub.firma.ps, probeset=2429278, option="probeset")

## get probeset expression levels for all probesets corresponding to transcript=2429277
expr.firma <- firma.expr(sub.firma.ps, probeset=2429277, option="probeset")

## End(Not run)
```

---

`firma.score`

*Get Splice Score from FIRMA*

---

## Description

Extracts the FIRMA splice score from `data.frame` data.

## Usage

```
firma.score(xps.data,
            probeset = NULL,
            option   = "probeset")
```

## Arguments

xps.data	object of class <a href="#">ExprTreeSet</a> .
probeset	probesetID or NULL.
option	option determining the probeset type for which to extract expression levels, one of 'probeset', 'exon'.

## Details

Function `firma.score` returns the FIRMA splice score described in Purdom et al. from slot data for a given probeset, or for all probesets in case of `probeset=NULL`. Row names will be the Affymetrix probesetIDs or exonIDs, respectively, dependent on the selected option.

## Value

A [data.frame](#).

## Note

For `option="probeset"` parameter `probeset` should usually be the transcriptID in order to get the splice scores for all probesetIDs of the corresponding transcriptID.

## Author(s)

Christian Stratowa

## References

Purdom, E., Simpson K.M., Robinson M.D., Conboy J.G., Lapuk A.V. and Speed, T.P. (2008), FIRMA: a method for detection of alternative splicing from exon array data. *Bioinformatics* 24(15):1707-1714

## See Also

[firma](#)

## Examples

```
## Not run:
## get probeset splice scores for all probeset or for probeset=2429278
score.firma <- firma.score(sub.firma.ps, probeset=NULL, option="probeset")
score.firma <- firma.score(sub.firma.ps, probeset=2429278, option="probeset")

## get probeset splice scores for all probesets corresponding to transcript=2429277
score.firma <- firma.score(sub.firma.ps, probeset=2429277, option="probeset")

## End(Not run)
```



**Description**

This function allows to combine different algorithms to compute background correction, normalization and fit a multichip model for summarization.

**Usage**

```
fitQC(xps.data,
      filename = character(),
      filedir = getwd(),
      tmpdir = "",
      update = FALSE,
      # background correction
      bgcorrect.method = "rma",
      bgcorrect.select = "none",
      bgcorrect.option = "pmonly:epanechnikov",
      bgcorrect.params = c(16384),
      # normalization
      normalize.method = "quantile",
      normalize.select = "pmonly",
      normalize.option = "transcript:together:none",
      normalize.logbase = "0",
      normalize.params = c(0.0),
      # quality control
      qualify.method = "rlm",
      qualify.select = "pmonly",
      qualify.qualopt = "all",
      qualify.option = "transcript",
      qualify.estimator = "huber",
      qualify.logbase = "log2",
      qualify.params = list(10, 0.01, 1.0),
      # reference values
      reference.index = 0,
      reference.method = "mean",
      reference.params = list(0.0),
      # misc.
      exonlevel = "",
      xps.scheme = NULL,
      add.data = FALSE,
      bufsize = 32000,
      verbose = TRUE)
```

```
xpsQualityControl(object, ...)
```

**Arguments**

xps.data	object of class DataTreeSet.
filename	file name of ROOT data file.

<code>filedir</code>	system directory where ROOT data file should be stored.
<code>tmpdir</code>	optional temporary directory where temporary ROOT files should be stored.
<code>update</code>	logical. If TRUE the existing ROOT data file filename will be updated.
<code>bgcorrect.method</code>	background method to use.
<code>bgcorrect.select</code>	type of probes to select for background correction.
<code>bgcorrect.option</code>	type of background correction to use.
<code>bgcorrect.params</code>	vector of parameters for background method.
<code>normalize.method</code>	normalization method to use.
<code>normalize.select</code>	type of probes to select for normalization.
<code>normalize.option</code>	normalization option.
<code>normalize.logbase</code>	logarithm base as character, one of '0', 'log', 'log2', 'log10'.
<code>normalize.params</code>	vector of parameters for normalization method.
<code>qualify.method</code>	qualification method to use, currently <code>rlm</code> .
<code>qualify.select</code>	type of probes to select for qualification.
<code>qualify.qualopt</code>	option determining the data to which to apply qualification, one of 'raw', 'adjusted', 'normalized', 'all'.
<code>qualify.option</code>	option determining the grouping of probes for qualification, one of 'transcript', 'exon', 'probeset'; exon arrays only.
<code>qualify.estimator</code>	option determining the M-estimator to use, one of 'huber', 'fair', 'cauchy', 'ge-manmcclore', 'welsch', 'tukey', 'andrew'.
<code>qualify.logbase</code>	logarithm base as character, one of '0', 'log', 'log2', 'log10'.
<code>qualify.params</code>	vector of parameters for qualification method.
<code>reference.index</code>	index of reference tree to use, or 0.
<code>reference.method</code>	for <code>refindex=0</code> , either trimmed mean or median of trees.
<code>reference.params</code>	vector of parameters for reference method.
<code>exonlevel</code>	exon annotation level determining which probes should be used for summarization; exon/genome arrays only.
<code>xps.scheme</code>	optional alternative <code>SchemeSet</code> .
<code>add.data</code>	logical. If TRUE expression data will be included as slot data.
<code>bufsize</code>	integer which sets the buffer size of the tree branch baskets (default is 32000).
<code>verbose</code>	logical, if TRUE print status information.
<code>object</code>	object of class <code>DataTreeSet</code> .
<code>...</code>	the arguments described above.

## Details

This function allows to combine different algorithms to compute background correction, normalization and fit a multichip model for summarization.

`xpsQualityControl` is the `DataTreeSet` method called by function `fitQC`, containing the same parameters.

## Value

An object of type `QualTreeSet`.

## Author(s)

Christian Stratowa

## See Also

[fitRLM](#), [qualify](#), [express](#)

## Examples

```
## Not run:
## load existing ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

## qualification - rlm
rlm.all <- fitQC(data.test3, "tmp_Test3RLMall", filedir=getwd(), tmpdir="",
                qualify.method="rlm", qualify.qualopt="all", qualify.option="transcript", add.data=FALSE)

## get expression data.frame
expr.rlm.all <- validData(rlm.all)

## get borders
brd.rlm.all <- borders(rlm.all)

## get residuals
res.rlm.all <- residuals(rlm.all)

## get weights
w.rlm.all <- weights(rlm.all)

## plot expression levels
if (interactive()) {
  coiplot(rlm.all)
  borderplot(rlm.all)
  nuseplot(rlm.all)
  rleplot(rlm.all)
  image(rlm.all, type="resids")
}

## End(Not run)
```

**Description**

Convert Affymetrix probe level data to expression levels by fitting RMA as multichip model.

**Usage**

```
fitRLM(xps.data,
       filename = character(),
       filedir  = getwd(),
       tmpdir   = "",
       background = "pmonly",
       normalize = TRUE,
       qualopt  = "all",
       option   = "transcript",
       exonlevel = "",
       params   = list(16384, 0.0, 1.0, 10, 0.01, 1),
       xps.scheme = NULL,
       add.data  = FALSE,
       bufsize   = 32000,
       verbose   = TRUE)
```

```
rmaPLM(xps.data,
       filename = character(),
       filedir  = getwd(),
       tmpdir   = "",
       background = "pmonly",
       normalize = TRUE,
       qualopt  = "all",
       option   = "transcript",
       exonlevel = "",
       params   = list(16384, 0.0, 1.0, 10, 0.01, 1),
       xps.scheme = NULL,
       add.data  = FALSE,
       bufsize   = 32000,
       verbose   = TRUE)
```

**Arguments**

xps.data	object of class DataTreeSet.
filename	file name of ROOT data file.
filedir	system directory where ROOT data file should be stored.
tmpdir	optional temporary directory where temporary ROOT files should be stored.
background	probes used to compute background, one of 'pmonly', 'mmonly', 'both'; for genome/exon arrays one of 'genomic', 'antigenomic'
normalize	logical. If TRUE normalize data using quantile normalization.

qualopt	option determining the data to which to apply qualification, one of 'raw', 'adjusted', 'normalized', 'all'.
option	option determining the grouping of probes for qualification, one of 'transcript', 'exon', 'probeset'; exon arrays only.
exonlevel	exon annotation level determining which probes should be used for summarization; exon/genome arrays only.
params	list of (default) parameters for rma.
xps.scheme	optional alternative SchemeSet.
add.data	logical. If TRUE expression data will be included as slot data.
bufsize	integer which sets the buffer size of the tree branch baskets (default is 32000).
verbose	logical, if TRUE print status information.

### Details

Convert Affymetrix probe level data to expression levels by fitting RMA as multichip model.

### Value

An object of type [QualTreeSet](#).

### Author(s)

Christian Stratowa

### See Also

[fitQC](#), [qualify](#), [express](#)

### Examples

```
## Not run:
## load existing ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

## qualification - rlm
rlm.all <- rmaPLM(data.test3, "tmp_Test3RLMall", filedir=getwd(), tmpdir="", qualopt="all", option="transcrip

## get borders
brd.rlm.all <- borders(rlm.all)

## get residuals
res.rlm.all <- residuals(rlm.all)

## get weights
w.rlm.all <- weights(rlm.all)

## plot expression levels
if (interactive()) {
  coiplot(rlm.all)
  borderplot(rlm.all)
  nuseplot(rlm.all)
  rleplot(rlm.all)
}
```

```

image(rlm.all, type="resids")
}

## End(Not run)

```

---

gapFilter-methods      *Gap Filter*

---

### Description

This method initializes the Gap Filter.

The `gapFilter` looks for genes that might usefully discriminate between two groups. To do this we look for a gap in the ordered expression values. The gap should come in the central portion, thus a parameter window is defined to exclude jumps in the initial window values and the final window values.

The Gap Filter flags all rows with: `flag = ((gap[i+1] - gap[i])/mean >= cutoff)`

```

gapFilter(object)
gapFilter(object, value)<-

```

### Arguments

<code>object</code>	object of class <code>PreFilter</code> .
<code>value</code>	numeric vector <code>c(cutoff, window, trim, epsilon)</code> .

### Details

The method `gapFilter` initializes the following parameters:

<code>cutoff</code> :	the cutoff level for the filter.
<code>window</code> :	trim value for the ordered expression levels (default is <code>window=0.05</code> ).
<code>trim</code> :	the trim value for trimmed mean (default is <code>trim=0</code> ).
<code>epsilon</code> :	value to replace mean (default is <code>epsilon=0.01</code> ):
	<code>epsilon &gt; 0</code> : replace mean=0 with epsilon.
	<code>epsilon = 0</code> : always set mean=1.

Note, that for `epsilon = 0` the filter flags all rows with: `(gap[i+1] - gap[i]) >= cutoff`

### Value

An initialized `PreFilter` object.

### Author(s)

Christian Stratowa

### Examples

```

prefltr <- PreFilter()
gapFilter(prefltr) <- c(0.3,0.05,0.0,0.01)
str(prefltr)

```

---

getChipName	<i>Get Chip Name</i>
-------------	----------------------

---

**Description**

Get chip name from ROOT scheme file.

**Usage**

```
getChipName(rootfile)
```

**Arguments**

rootfile            name of ROOT scheme file, including full path.

**Details**

Extracts the chip name directly from [ROOT](#) scheme file rootfile.

**Value**

a character with the chip name.

**Author(s)**

Christian Stratowa

**See Also**

[getChipType](#), [getNameType](#)

**Examples**

```
## correct usage
getChipName(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
## incorrect usage
getChipName(paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))
```

---

getChipType	<i>Get Chip Type</i>
-------------	----------------------

---

**Description**

Get chip type from ROOT scheme file.

**Usage**

```
getChipType(rootfile)
```

**Arguments**

rootfile            name of ROOT scheme file, including full path.

**Details**

Extracts the chip type directly from [ROOT](#) scheme file rootfile.

**Value**

a character with the chip type, either 'GeneChip' or 'ExonChip'.

**Author(s)**

Christian Stratowa

**See Also**

[getChipName](#), [getNameType](#)

**Examples**

```
## correct usage
getChipType(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
## incorrect usage
getChipType(paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))
```

---

getDatatype

*Get Data Type*

---

**Description**

Get data type corresponding to tree type.

**Usage**

```
getDatatype(treetype)
```

**Arguments**

treetype      tree type.

**Details**

Get data type corresponding to tree type. Valid tree types are described in [validTreetype](#).

**Value**

a character with the correct data type, i.e. 'rawdata', 'preprocess' or 'normation'.

**Author(s)**

Christian Stratowa

**See Also**

[type2Exten](#), [validTreetype](#)



**Examples**

```
getDatatype("cel")
getDatatype("tbw")
```

---

getNameType	<i>Get Chip Name and Type</i>
-------------	-------------------------------

---

**Description**

Get chip name and type from ROOT scheme file.

**Usage**

```
getNameType(rootfile)
```

**Arguments**

rootfile            name of ROOT scheme file, including full path.

**Details**

Extracts the chip name and type directly from [ROOT](#) scheme file rootfile.

**Value**

a list with parameters:

chipname            chip name.  
chiptype            chip type, either 'GeneChip' or 'ExonChip'.

**Author(s)**

Christian Stratowa

**See Also**

[getChipName](#), [getChipType](#)

**Examples**

```
## correct usage
getNameType(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
## incorrect usage
getNameType(paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))
```

---

getNumberTrees	<i>Get Number of Trees</i>
----------------	----------------------------

---

**Description**

Get number of trees stored in a ROOT file.

**Usage**

```
getNumberTrees(rootfile, treetype = "*", setname = NULL)
```

**Arguments**

rootfile	name of ROOT file, including full path.
treetype	tree type.
setname	name of ROOT subdirectory containing trees.

**Details**

Extracts the number of trees of treetype stored in [ROOT](#) file rootfile.

Valid tree types are listed in [validTreetype](#). For treetype="\*" the total number of trees in rootfile are returned.

If setname is provided, only trees in subdirectory setname are counted.

**Value**

Number of trees.

**Author(s)**

Christian Stratowa

**Examples**

```
getNumberTrees(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
getNumberTrees(paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))
```

---

getProbeInfo	<i>Get Probe Information</i>
--------------	------------------------------

---

**Description**

Get GeneChip probe information from root scheme file.

**Usage**

```
getProbeInfo(rootfile)
```

**Arguments**

rootfile	name of ROOT scheme file, including full path.
----------	--

**Details**

Extracts GeneChip probe information directly from [ROOT](#) scheme file rootfile.

**Value**

a list with parameters:

nrows	physical number of rows in the array.
ncols	physical number of columns in the array.
nprobes	number of probes on the array.
ncontrols	number of controls on the array.
ngenes	number of genes on the array.
nunits	number of units on the array.
nprobesets	umber of probesets on the array.
naffx	number of AFFX controls on the array.

**Author(s)**

Christian Stratowa

**Examples**

```
getProbeInfo(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
```

---

getTreeData-methods    *Export Tree Data*

---

**Description**

Exports tree data from [ROOT](#) data file and and saves as data.frame.

*Usage*

```
getTreeData(object, treetype = "cel", varlist = "fInten")
```

**Arguments**

object	Object of class "ProcesSet".
treetype	type of tree to export, see <a href="#">validTreetype</a>
varlist	names of tree leaves to export.

**Details**

Exports tree leaves from [ROOT](#) data file and and saves as data.frame.

**Value**

A [data.frame](#).

**Author(s)**

Christian Stratowa

**See Also**

[export](#)

---

getTreeNames

*Get Tree Names*

---

**Description**

Get tree names stored in a ROOT file.

**Usage**

```
getTreeNames(rootfile, treetype = "*", setname = NULL, gettitle = FALSE)
```

**Arguments**

rootfile	name of ROOT file, including full path.
treetype	tree type.
setname	name of ROOT subdirectory containing trees.
gettitle	If TRUE the titles of the trees will be returned.

**Details**

Extracts the tree names of treetype stored in ROOT file rootfile.

Valid tree types are listed in [validTreetype](#). For treetype="\*" names for all trees in rootfile are returned.

If setname is provided, only tree names in subdirectory setname are returned.

**Value**

A vector of tree names. For gettitle=TRUE a vector of tree titles.

**Author(s)**

Christian Stratowa

**Examples**

```
getTreeNames(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
getTreeNames(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"), "scm")
getTreeNames(paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))
```

---

**highFilter-methods**      *Upper Threshold Filter*

---

**Description**

This method initializes the Upper Threshold Filter.

The cutoff value defines the upper threshold for allowed expression levels. If e.g. the number of samples exceeding this cutoff value is greater than parameter then the corresponding dataframe row is flagged, i.e. `flag = 0`.

The Upper Threshold Filter flags all rows with: `flag = (sum(expression[i] <= cutoff) >= parameter)`

*Usage*

```
highFilter(object)
highFilter(object, value)<-
```

**Arguments**

object	object of class <code>PreFilter</code> .
value	character vector <code>c(cutoff, parameter, condition)</code> .

**Details**

The method `highFilter` initializes the following parameters:

cutoff:	the upper threshold level for the filter.
parameter:	this value depends on the condition used:
condition:	condition="samples": number of samples (default):
	condition="percent": percent of samples.
	condition="mean": mean value of samples.
	condition="percentile": percentile of samples.

**Value**

An initialized `PreFilter` object.

**Author(s)**

Christian Stratowa

**Examples**

```
prefltr <- PreFilter()
highFilter(prefltr) <- c(14.5,75.0,"percent")
str(prefltr)
```

**Description**

Plot the density estimates for each sample.

*Usage*

```
hist(x, which = "", size = 0, transfo = log2, xlab = "log intensity",
```

**Arguments**

x	object of class <a href="#">DataTreeSet</a> or <a href="#">ExprTreeSet</a> .
which	type of probes to be used, for details see <a href="#">validData</a> .
size	length of sequence to be generated as subset.
transfo	a valid function to transform the data, usually “log2”, or “0”.
xlab	a title for the x axis.
ylab	a title for the y axis.
names	optional vector of sample names.
type	type for the plot.
col	colors to use for the different arrays.
lty	line types to use for the different arrays.
add.legend	logical, if TRUE then a legend will be drawn.
verbose	logical, if TRUE print status information.
...	optional arguments to be passed to plot.

**Details**

Plots the non-parametric density estimates for each sample.

For names=NULL full column names of slot data will be displayed while for names="namepart" column names will be displayed without name extension. If names is a vector of column names, only these columns will displayed as callplot.

**Note**

For objects of class [DataTreeSet](#) it is no longer necessary to attachInten since each data tree will be imported separately.

**Author(s)**

Christian Stratowa

**See Also**

[plotDensity](#)

**Examples**

```
## load existing ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

if (interactive()) {
  hist(data.test3)
}
```

image-methods

*Display an Image***Description**

Creates an image of intensities or residuals, respectively, for each sample.

*Usage*

```
image(x,          bg          = FALSE,          transfo  = log2,          col        = NULL,
image(x,          type        = c("resids", "pos.resids", "neg.resids", "sign.resids", "weights"),
```

**Arguments**

x	object of class <a href="#">ProcesSet</a> .
bg	logical. If FALSE, intensities from slot data will be used; if TRUE, background intensities from slot bgrd will be used.
type	character string specifying the type of residual image.
qualopt	character string specifying whether to draw residual image for “raw”, “adjusted”, or “normalized” intensities.
transfo	a valid function to transform the data, usually “log2”, or “0”.
col	color range for intensities.
names	optional vector of sample names.
xlab	a label for the x axis.
ylab	a label for the y axis.
add.legend	logical, if TRUE then a color bar will be drawn.
...	optional arguments to be passed to <code>image</code> .

**Details**

Creates an image of intensities or residuals, respectively, for each array, i.e. ‘pseudo chip images’.

If x belongs to class `DataTreeSet` then images of raw intensities will be drawn.

If x belongs to class `ExprTreeSet` and `bg=FALSE` then images of background corrected intensities will be drawn.

If x belongs to class `ExprTreeSet` and `bg=TRUE` the distribution of the background intensities will be shown; this can be useful to see potential density gradients caused by hybridization conditions. For the computation of background intensities see function `bgcorrect`; it is suggested to use `bgcorrect.mas4` to identify density gradients.

If x belongs to class `QualTreeSet` then images of the residuals or the probe weights, respectively, will be drawn. For `col=NULL` the same colors will be used as described in vignette “QualityAssess.pdf” of package `affyPLM`, using internally function `pseudoPalette` described in `affyPLM`.

For `names=NULL` full tree names will be displayed while for `names="namepart"` column names will be displayed without name extension. If `names` is a vector of tree names then data from these trees only will displayed as `image(s)`.

**Author(s)**

Christian Stratowa

**See Also**[plotImage](#)**Examples**

```
## Not run:
## images of raw intensities as imported using import.data()
unlist(treeNames(data.test3)) # show available tree names
image(data.test3, names="TestA2.cel")
image(data.test3)

## images of background adjusted or background intensities, created by e.g. rma()
getTreeNames(rootFile(data.rma))
image(data.rma, names="TestA2.int")
image(data.rma, names="TestA2.rbg", bg=TRUE)

## residual images, created by e.g. rmaPLM()
getTreeNames(rootFile(rlm.all), treetype="res")
image(rlm.all, type="resids")
image(rlm.all, type="resids", names="TestA2_raw.res", add.legend=TRUE)
image(rlm.all, type="pos.resids", names="TestA2_raw.res", add.legend=TRUE)
image(rlm.all, type="neg.resids", names="TestA2_raw.res", add.legend=TRUE)
image(rlm.all, type="sign.resids", names="TestA2_raw.res", add.legend=TRUE)
image(rlm.all, type="weights", names="TestA2_raw.res", add.legend=TRUE)
image(rlm.all, type="resids", qualopt="adjusted", names="TestA2_adjusted.res", add.legend=TRUE)

## End(Not run)
```

---

`import.data`*Import CEL files into a DataTreeSet*

---

**Description**

Import the Affymetrix CEL files into a ROOT file and create S4 class DataTreeSet

**Usage**

```
import.data(xps.scheme,
            filename = character(0),
            filedir  = getwd(),
            celdir   = NULL,
            celfiles = "*",
            celnames = NULL,
            project  = NULL,
            verbose  = TRUE)
```

**Arguments**

<code>xps.scheme</code>	a <a href="#">SchemeTreeSet</a> containing the correct scheme for the CEL-files
<code>filename</code>	file name of ROOT data file.
<code>filedir</code>	system directory where ROOT data file should be stored.
<code>celdir</code>	system directory containing the CEL-files for corresponding scheme.



celfiles	optional vector of CEL-files to be imported.
celnames	optional vector of names which should replace the CEL-file names.
project	optional class <a href="#">ProjectInfo</a> .
verbose	logical, if TRUE print status information.

### Details

`import.data` is used to import CEL-files from directory `celdir` into a [ROOT](#) data file. To import only a subset of CEL-files, list these CEL-files as vector `celfiles`.

To import CEL-files from different directories, vector `celfiles` must contain the full path for each CEL-file and `celdir` must be `celdir=NULL`.

The optional parameter `celnames` allows you to replace the original CEL-file names with names of your choice, otherwise the names of the CEL-files will be used as `celnames`.

Currently, the following types of Affymetrix CEL-files can be imported: text (version 3), xml, binary (xda), generic (agcc,calvin)

An S4 class [DataTreeSet](#) will be created, serving as R wrapper to the [ROOT](#) data file filename.

Use function [root.data](#) to access the [ROOT](#) data file from new R sessions to avoid creating a new [ROOT](#) data file for every session.

### Value

A [DataTreeSet](#) object.

### Note

As mentioned above, use function [root.data](#) to access the [ROOT](#) data file from new R sessions to avoid creating a new [ROOT](#) data file for every R session.

Do not separate filename of [ROOT](#) files with dots, use underscores, e.g. do not use `filename="Data.Test3"` but use `filename="Data_Test3"` or `filename="DataTest3"` instead.

To every [ROOT](#) data file the extension `"_cel"` is attached to filename to easily recognize [ROOT](#) data files containing the raw CEL data, e.g. for `filename="Data_Test3"` the final name is `"Data_Test3_cel.root"`. Extension `"root"` is added automatically, so that [ROOT](#) is able to recognize the file as [ROOT](#) file.

Once a [ROOT](#) file is created it can not be overwritten, it must be deleted manually first. Only [ROOT](#) files called `"tmp"` or with filename starting with `"tmp_"` will be re-created automatically.

If CEL-file names contain dots, colons, parenthesis, etc. as characters, these characters will be replaced by underscores. It is recommended to use parameter `celnames` to create shorter CEL names and to replace special characters.

### Author(s)

Christian Stratowa

### See Also

[root.data](#), [DataTreeSet](#)

**Examples**

```
## get scheme and import CEL-files from package
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- import.data(scheme.test3, "tmp_data_test3", celdir=paste(path.package("xps"), "raw", sep="/"))
unlist(treeNames(data.test3))

## import only subset of CEL-files
subdata.test3 <- import.data(scheme.test3, "tmpdt_data_test3", celdir=paste(path.package("xps"), "raw", sep="/"),
                             celfiles=c("TestA1.CEL", "TestB2.CEL"), verbose=FALSE)
unlist(treeNames(subdata.test3))
```

---

import.exon.scheme      *Import CLF, PGF and annotation files into a SchemeTreeSet*

---

**Description**

Import the Affymetrix CLF, PGF, and probeset and transcript annotation files into a ROOT file and create S4 class SchemeTreeSet

**Usage**

```
import.exon.scheme(filename = character(0),
                   filedir  = getwd(),
                   layoutfile = character(0),
                   schemefile = character(0),
                   probeset  = character(0),
                   transcript = character(0),
                   control   = "",
                   add.mask  = FALSE,
                   verbose   = TRUE)
```

**Arguments**

filename	file name of ROOT scheme file.
filedir	system directory where ROOT scheme file should be stored.
layoutfile	name of CLF-file, including full path.
schemefile	name of PGF-file, including full path.
probeset	name of probeset annotation-file, including full path.
transcript	name of transcript annotation-file, including full path.
control	optional name of controls.ps-file, including full path.
add.mask	logical. If TRUE mask information will be included as slot mask.
verbose	logical, if TRUE print status information.

**Details**

`import.exon.scheme` is used to import all information for an Affymetrix exon array into a [ROOT](#) scheme file, including CLF and PGF-files, and the current Affymetrix probeset and transcript annotation files.

An S4 class `SchemeTreeSet` will be created, serving as R wrapper to the [ROOT](#) scheme file `filename`. Since a new [ROOT](#) scheme file needs only to be created when new annotation files are available from the Affymetrix website, it is recommended to store all [ROOT](#) scheme files in a commonly accessible system directory `filedir`.

Use function `root.scheme` to access the [ROOT](#) scheme file from new R sessions to avoid creating a new [ROOT](#) scheme file for every session.

**Value**

A `SchemeTreeSet` object.

**Warning**

The current version of ‘xps’ should be able to import all Affymetrix exon array annotation files up to September 2011. However, since Affymetrix is still changing the headers and/or columns of the annotation files, future annotation files may require adaptation of the source code, thus the current version of ‘xps’ may not be able to read these files.

**Note**

As mentioned above, use function `root.scheme` to access the [ROOT](#) scheme file from new R sessions to avoid creating a new [ROOT](#) scheme file for every R session.

Do not separate `filename` of [ROOT](#) files with dots, use underscores, e.g. do not use `filename="Scheme.HuEx10stv2r2"` but use `filename="Scheme_HuEx10stv2r2_na32"` instead. Extension “root” is added automatically, so that [ROOT](#) is able to recognize the file as [ROOT](#) file.

Do not set `add.mask=TRUE` unless you know that your computer has sufficient RAM.

Do not add `item control` unless you want to use one of the old annotation files where the probeset annotation file does not contain the AFX controls.

**Author(s)**

Christian Stratowa

**See Also**

[import.expr.scheme](#), [root.scheme](#), [SchemeTreeSet](#)

**Examples**

```
## Not run:
## define paths
smdir <- "/common/path/schemes"
libdir <- "/my/path/Affy/libraryfiles"
anndir <- "/my/path/Affy/Annotation"

## create scheme for HuEx-1_0-st-v2.r2 Exon array
scheme.huex10stv2r2.na32 <- import.exon.scheme("Scheme_HuEx10stv2r2_na32", filedir=smdir,
                                             layoutfile=file.path(libdir, "HuEx-1_0-st-v2_libraryfile", "HuEx-1_0-st-r2/HuEx-1_0-s
```

```

schemefile=file.path(libdir, "HuEx-1_0-st-v2_libraryfile", "HuEx-1_0-st-r2/HuEx-1_0-
probeset=file.path(anndir, "HuEx-1_0-st-v2.na32.hg19.probeset.csv"),
transcript=file.path(anndir, "HuEx-1_0-st-v2.na32.hg19.transcript.csv"))

## access ROOT scheme file from new R session
scheme.exon <- root.scheme(paste(scmdir, "Scheme_HuEx10stv2r2_na32.root", sep="/"))

## create scheme for HuGene-1_0-st-v1.r4 as exon array
scheme.hugene10stv1r4.na32 <- import.exon.scheme("Scheme_HuGene10stv1r4_na32",filedir=scmdir,
layoutfile=file.path(libdir, "HuGene-1_0-st-v1.r4.analysis-lib-files", "HuGene-1_0-
schemefile=file.path(libdir, "HuGene-1_0-st-v1.r4.analysis-lib-files", "HuGene-1_0-
probeset=file.path(anndir, "HuGene-1_0-st-v1.na32.hg19.probeset.csv"),
transcript=file.path(anndir, "HuGene-1_0-st-v1.na32.hg19.transcript.csv"))

## access ROOT scheme file from new R session
scheme.gene <- root.scheme(file.path(scmdir, "Scheme_HuGene10stv1r4_na32.root"))

## create scheme for HuEx-1_0-st-v2.r2 Exon array with the old annotation file
scheme.huex10stv2r2.old <- import.exon.scheme("Scheme_HuEx10stv2r2_old",filedir=scmdir,
layoutfile=file.path(libdir, "HuEx-1_0-st-v2_libraryfile", "HuEx-1_0-st-r2", "HuEx-1_0-
schemefile=file.path(libdir, "HuEx-1_0-st-v2_libraryfile", "HuEx-1_0-st-r2", "HuEx-1_0-
probeset=file.path(anndir, "HuEx-1_0-st-probeset-annot.csv"),
transcript=file.path(anndir, "HuEx-1_0-st-transcript-annot.csv"),
control=file.path(libdir, "HuEx-1_0-st-v2_libraryfile", "HuEx-1_0-st-r2", "HuEx-1_0-st-

## End(Not run)

```

---

```
import.expr.scheme      Import CDF, probe and annotation files into a SchemeTreeSet
```

---

## Description

Import the Affymetrix CDF, probe and annotation files into a ROOT file and create S4 class Scheme-TreeSet

## Usage

```

import.expr.scheme(filename = character(0),
                   filedir  = getwd(),
                   schemefile = character(0),
                   probefile  = character(0),
                   annotfile  = character(0),
                   chipname   = NULL,
                   add.mask   = FALSE,
                   verbose    = TRUE)

```

## Arguments

filename	file name of ROOT scheme file.
filedir	system directory where ROOT scheme file should be stored.
schemefile	name of CDF-file, including full path.

probefile	name of probe-file, including full path.
annotfile	name of annotation-file, including full path.
chipname	optional chip name when using an alternative CDF-file.
add.mask	logical. If TRUE mask information will be included as slot mask.
verbose	logical, if TRUE print status information.

### Details

`import.expr.scheme` is used to import all information for an Affymetrix expression array into a [ROOT](#) scheme file, including CDF-file, the corresponding probe file, and the current Affymetrix annotation file.

Usually, `chipname` is extracted from the name of the CDF-file, however, when using an alternative CDF-file, e.g. from BrainArray or AffyProbeMiner, a `chipname` must be supplied which starts with (or contains) the exact Affymetrix chip name.

An S4 class [SchemeTreeSet](#) will be created, serving as R wrapper to the [ROOT](#) scheme file filename.

Since a new [ROOT](#) scheme file needs only to be created when a new annotation file is available from the Affymetrix website, it is recommended to store all [ROOT](#) scheme files in a commonly accessible system directory `filedir`.

Use function [root.scheme](#) to access the [ROOT](#) scheme file from new R sessions to avoid creating a new [ROOT](#) scheme file for every session.

### Value

A [SchemeTreeSet](#) object.

### Note

As mentioned above, use function [root.scheme](#) to access the [ROOT](#) scheme file from new R sessions to avoid creating a new [ROOT](#) scheme file for every R session.

Do not separate filename of [ROOT](#) files with dots, use underscores, e.g. do not use `filename="Scheme.Test3.na32"` but use `filename="Scheme_Test3_na32"` or simply `filename="SchemeTest3na32"` instead. Extension "root" is added automatically, so that [ROOT](#) is able to recognize the file as [ROOT](#) file.

For a few probesets, parsing the Affymetrix annotation files will provide ambiguous results. Setting `verbose=11` will list these probesets.

### Author(s)

Christian Stratowa

### See Also

[import.exon.scheme](#), [import.genome.scheme](#), [root.scheme](#), [SchemeTreeSet](#)

### Examples

```
## Not run:
## define paths
scmdir <- "/common/path/schemes"
libdir <- "/my/path/Affy/libraryfiles"
anndir <- "/my/path/Affy/Annotation"
```

```
## create scheme for Test3 GeneChip
scheme.test3.na32 <- import.expr.scheme("Scheme_Test3_na32", filedir=scmdir,
                                       schemefile=file.path(libdir, "Test3.CDF"),
                                       probefile=file.path(libdir, "Test3_probe.tab"),
                                       annotfile=file.path(anndir, "Test3.na32.annot.csv"))

## access ROOT scheme file from new R session
scheme.test3 <- root.scheme(file.path(scmdir, "Scheme_Test3_na32.root"))

## create scheme for HG-U133_Plus_2 GeneChip
scheme.hgu133p2.na32 <- import.expr.scheme("Scheme_HGU133p2_na32", filedir=scmdir,
                                           schemefile=file.path(libdir, "HG-U133_Plus_2.cdf"),
                                           probefile=file.path(libdir, "HG-U133-PLUS_probe.tab"),
                                           annotfile=file.path(anndir, "HG-U133_Plus_2.na32.annot.csv"))

## access ROOT scheme file from new R session
scheme.hgu133p2 <- root.scheme(file.path(scmdir, "Scheme_HGU133p2_na32.root"))

## End(Not run)
```

---

import.genome.scheme    *Import CLF, PGF and annotation files into a SchemeTreeSet*

---

## Description

Import the Affymetrix CLF, PGF and transcript annotation files into a ROOT file and create S4 class SchemeTreeSet

## Usage

```
import.genome.scheme(filename = character(0),
                    filedir   = getwd(),
                    layoutfile = character(0),
                    schemefile = character(0),
                    transcript  = character(0),
                    add.mask    = FALSE,
                    verbose     = TRUE)
```

## Arguments

filename	file name of ROOT scheme file.
filedir	system directory where ROOT scheme file should be stored.
layoutfile	name of CLF-file, including full path.
schemefile	name of PGF-file, including full path.
transcript	name of transcript annotation-file, including full path.
add.mask	logical. If TRUE mask information will be included as slot mask.
verbose	logical, if TRUE print status information.

## Details

`import.genome.scheme` is used to import all information for an Affymetrix whole genome array into a [ROOT](#) scheme file, including CLF and PGF-files, and the current Affymetrix transcript annotation files.

An S4 class [SchemeTreeSet](#) will be created, serving as R wrapper to the [ROOT](#) scheme file `filename`.

Since a new [ROOT](#) scheme file needs only to be created when new annotation files are available from the Affymetrix website, it is recommended to store all [ROOT](#) scheme files in a commonly accessible system directory `filedir`.

Use function [root.scheme](#) to access the [ROOT](#) scheme file from new R sessions to avoid creating a new [ROOT](#) scheme file for every session.

## Value

A [SchemeTreeSet](#) object.

## Warning

The current version of 'xps' is able to import all Affymetrix genome array annotation files up to November 2008, i.e. all files of release 3 (r3) and earlier. However, in January 2009 Affymetrix has updated all CLF, PGF and annotation files to release 4 (r4) and added a new probeset annotation file, thus in effect changing the whole genome arrays to exon arrays!

Thus, for release 4 (r4) files, function `import.genome.scheme` can no longer be used, but you must use function [import.exon.scheme](#) instead (see examples).

## Note

As mentioned above, use function [root.scheme](#) to access the [ROOT](#) scheme file from new R sessions to avoid creating a new [ROOT](#) scheme file for every R session.

Do not separate `filename` of [ROOT](#) files with dots, use underscores, e.g. do not use `filename="Scheme.HuGene10stv1"` but use `filename="Scheme_HuGene10stv1_na27"` instead. Extension "root" is added automatically, so that [ROOT](#) is able to recognize the file as [ROOT](#) file.

Do not set `add.mask=TRUE` unless you know that your computer has sufficient RAM.

Do not add `item.control` unless you want to use one of the old annotation files where the probeset annotation file does not contain the AFFX controls.

## Author(s)

Christian Stratowa

## See Also

[import.exon.scheme](#), [root.scheme](#), [SchemeTreeSet](#)

## Examples

```
## Not run:
## define paths
smdir <- "/common/path/schemes"
libdir <- "/my/path/Affy/libraryfiles"
anndir <- "/my/path/Affy/Annotation"

## create scheme for HuGene-1_0-st-v1 whole genome array
```

```

scheme.hugene10stv1r3.na27 <- import.genome.scheme("Scheme_HuEx10stv1r3_na27",
  filedir=scmdir,
  layoutfile=file.path(libdir, "HuGene-1_0-st-v1.r3.analysis_libraryfile", "HuGene-1_0-st-v1.r3.analysis_libraryfile"),
  schemefile=file.path(libdir, "HuGene-1_0-st-v1.r3.analysis_libraryfile", "HuGene-1_0-st-v1.r3.analysis_libraryfile"),
  transcript=file.path(anndir, "HuGene-1_0-st-v1.na27.hg18.transcript.csv"))

## access ROOT scheme file from new R session
scheme.hugene10stv1r3 <- root.scheme(file.path(scmdir, "Scheme_HuEx10stv1r3_na27.root"))

## End(Not run)

```

---

indexUnits-methods      *Unit Locations*

---

## Description

Returns a data.frame or list with locations of the probes in each probe set.

### Usage

```

indexUnits(object, which = "", unitID = NULL, unittype = "transcript", as.list = TRUE, data = NULL)
pmindex(object, unitID = NULL, as.list = TRUE)
mmindex(object, unitID = NULL, as.list = TRUE)

```

## Arguments

object	Object of class "DataTreeSet".
which	type of probes to be used, for details see <a href="#">validData</a> .
unitID	optional vector of UNIT_IDs.
unittype	character vector, "transcript" or "probeset".
as.list	if TRUE a list will be returned (default is data.frame).
data	optional data.frame containing (x,y)-coordinates.

## Details

Function indexUnits returns a data.frame or list with locations of the probes in each probe set.

By default a data.frame for selected unitIDs or all unitIDs (unitID="\*") will be returned with columns <UNIT\_ID, X, Y, XY>. Here "XY" are the selected rows of slot data.

For as.list=TRUE a list of unitIDs will be returned containing the selected rows "XY". The names of the elements in the list returned are the UNIT\_IDs.

For unitID=NULL a vector of data rows "XY" will be returned.

For expression arrays which can be one of "pm", "mm", or "both". Alternatively, functions pmindex and mmindex can be used for PM probes or MM probes, respectively.

For exon arrays which is described in [validData](#). However, in this case slot data must contain the (x,y)-coordinates of the probesetIDs.

## Value

A list or data.frame.



**Author(s)**

Christian Stratowa

**See Also**[unitID2transcriptID](#), [unitID2probesetID](#)**Examples**

```
## first, load ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

## dataXY not attached
id <- indexUnits(data.test3, which="pm", unitID=c(34,36,122))
id

## dataXY attached (only necessary for whole genome and exon arrays)
data.test3 <- attachDataXY(data.test3)
xy <- treeData(data.test3)
id <- indexUnits(data.test3, which="pm", unitID=c(34,36,122), data=xy)
id
id <- indexUnits(data.test3, which="", unitID=c(34,36,122), data=xy)
id
id <- indexUnits(data.test3, which="", unitID=34, as.list=FALSE, data=xy)
id
data.test3 <- removeDataXY(data.test3)

rm(scheme.test3, data.test3)
gc()
```

ini.call

*Informative/Non-Informative Call***Description**

Computes the Informative/Non-Informative Call for the exclusion of non-informative probe sets.

**Usage**

```
ini.call(xps.data,
        filename = character(0),
        filedir  = getwd(),
        tmpdir   = "",
        weight   = 0.5,
        mu       = 0.0,
        scale    = 1.0,
        tol      = 0.00001,
        cyc      = 100,
        alpha1   = 0.4,
        alpha2   = 0.6,
        version  = "1.3.1",
        option   = "transcript",
```

```

    exonlevel = "",
    xps.scheme = NULL,
    add.data = TRUE,
    verbose = TRUE)

```

```
xpsINICall(object, ...)
```

### Arguments

xps.data	object of class DataTreeSet.
filename	file name of ROOT data file.
filedir	system directory where ROOT data file should be stored.
tmpdir	optional temporary directory where temporary ROOT files should be stored.
weight	hyperparameter, usually set to 0.5 for version="1.3.1" and to 8.0 for version="1.3.0".
mu	hyperparameter allowing to correct for potential bias.
scale	scaling parameter, usually set to 1.0 for version="1.3.1" and to 2.0 for version="1.3.0".
tol	termination tolerance for EM algorithm.
cyc	maximum number of cycles of EM algorithm.
alpha1	a significance threshold in (0,alpha2).
alpha2	a significance threshold in (alpha1,1.0).
version	version of original farms package. Currently, version="1.3.1" and version="1.3.0" are implemented. Default is version="1.3.1".
option	option determining the grouping of probes for summarization, one of 'transcript', 'exon', 'probeset'; exon arrays only.
exonlevel	exon annotation level determining which probes should be used for summarization; exon/genome arrays only.
xps.scheme	optional alternative SchemeTreeSet.
add.data	logical. If TRUE call data will be added to slots data and detcall.
verbose	logical, if TRUE print status information.
object	object of class DataTreeSet.
...	the arguments described above.

### Details

In contrast to [mas5.call](#) this function quantifies the signal-to-noise ratio for each probe set, as described in Talloen et al. Thus, the returned p-values and detection calls have a different meaning:

The p-value that is returned estimates the signal-to-noise ratio (SNR):

P-values (SNR) of less than 0.5 indicate that there is more signal than noise and the corresponding genes are considered to be 'informative' for further analysis. In contrast, values greater than 0.5 indicate 'non-informative' genes.

The informative call is computed by thresholding the p-value as in:

```

call "P" if p-value < alpha1
call "M" if alpha1 <= p-value < alpha2
call "A" if alpha2 <= p-value

```

Here "P" should be considered as informative "I", "M" as marginally informative, and "A" as non-informative "NI".

The defaults for  $\alpha_1=0.4$  and  $\alpha_2=0.6$  are set to allow “M” calls. In order to get the same results as package ‘farms\1.3.1’, you need to set  $\alpha_1=0.5$  and  $\alpha_2=0.5$ .

For exon/genome arrays it is necessary to supply option and exonlevel.

Following options are valid for exon arrays only:

transcript: expression levels are computed for transcript clusters, i.e. probe sets containing the same ‘transcript\_cluster\_id’.  
 exon: expression levels are computed for exon clusters, i.e. probe sets containing the same ‘exon\_id’, where each probe set contains only one exon.  
 probeset: expression levels are computed for individual probe sets, i.e. for each ‘probeset\_id’.

Following exonlevel annotations are valid for exon arrays:

core: probesets supported by RefSeq and full-length GenBank transcripts.  
 metacore: core meta-probesets.  
 extended: probesets with other cDNA support.  
 metaextended: extended meta-probesets.  
 full: probesets supported by gene predictions only.  
 metafull: full meta-probesets.  
 ambiguous: ambiguous probesets only.  
 affx: standard AFFX controls.  
 all: combination of above.

Following exonlevel annotations are valid for whole genome arrays:

core: probesets with category ‘unique’ and ‘mixed’.  
 metacore: probesets with category ‘unique’ only.  
 affx: standard AFFX controls.  
 all: combination of above.

Exon levels can also be combined, with following combinations being most useful:

exonlevel="metacore+affx": core meta-probesets plus AFFX controls  
 exonlevel="core+extended": probesets with cDNA support  
 exonlevel="core+extended+full": supported plus predicted probesets

Exon level annotations are described in the Affymetrix whitepaper ‘exon\_probeset\_trans\_clust\_whitepaper.pdf’.

In order to use an alternative [SchemeTreeSet](#) set the corresponding SchemeTreeSet xps.scheme.

xpsINICall is the DataTreeSet method called by function ini.call, containing the same parameters.

## Value

A [CallTreeSet](#)

## Note

Since I/NI-calls distinguish only between informative and non-informative genes, the calls are identical for all samples.

**Author(s)**

Christian Stratowa

**References**

Talloon, W., Clevert D.-A., Hochreiter, S., Amaratunga, D., Bijmens, J., Kass, S., and Gohlmann, H.W.H. (2006), I/NI-calls for the exclusion of non-informative genes: a highly effective filtering tool for microarray data. *Bioinformatics* 23(21):2897-2902

**See Also**

[farms](#), [mas5.call](#)

**Examples**

```
## first, load ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

## I/NI call
call.ini <- ini.call(data.test3, "tmp_Test3INI", verbose=FALSE)

## get data.frames
snr.ini <- pvalData(call.ini)
inf.ini <- presCall(call.ini)
head(snr.ini)
head(inf.ini)

## plot results
if (interactive()) {
  callplot(call.ini)
}

rm(scheme.test3, data.test3)
gc()
```

---

initialize-methods      *Initialize Classes*

---

**Description**

Initialize S4 classes.

**Methods**

Internal method to initialize S4 classes.

**Description**

Get/set data values from/for class `DataTreeSet`.

*Usage*

```
intensity(object)
intensity(object, filename = NULL, verbose = FALSE) <- value
```

**Arguments**

<code>object</code>	object of class <code>DataTreeSet</code> .
<code>filename</code>	character vector containing optional ROOT file name.
<code>verbose</code>	logical, if TRUE print status information.
<code>value</code>	<code>data.frame</code> containing expression values.

**Details**

Get the intensity values from slot data or set slot data to value.

Method `intensity` returns the data values from slot data as `data.frame`, while replacement method `intensity<-` allows to replace slot data with a `data.frame`.

Using replacement method `intensity<-` with default settings will not change the data stored in the ROOT data file, and thus will not have any effect on subsequent processing methods. If you really want to use the replacement data for further processing you must supply a new ROOT filename. This will export each intensity column of value as CEL-file (version 3), which will then be imported into the new ROOT data file filename.

Warning: Do not use replacement method `intensity<-` until you really know what you are doing!

Note: The first two columns of replacement `data.frame` value must be the (X,Y) coordinates, followed by the intensities whereby the number of intensity columns must be identical to the columns to be replaced.

Note: If you do not want to replace your current object, create first a copy of type `DataTreeSet` by simply writing `newobj <- oldobj`, and use `newobj` for replacement. This is important since `intensity<-` does also update slots `rootfile`, `filedir` and `treenames` when a new filename was chosen.

Note: The CEL-files created are fully functional CEL-files (version 3), however some header rows such as `GridCornerUL`, `AlgorithmParameters`, and some of the data in `DatHeader` are placeholders only.

Warning: The CEL-files created WILL REPLACE THE ORIGINAL CEL-files, if they have identical names to the original CEL-files and the original CEL-files are located in the working directory. Thus the original CEL-files should preferable be located in directory `celdir` of function `import.data`.

**Author(s)**

Christian Stratowa

**See Also**[validData](#)**Examples**

```
## Not run:
## load ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

## get intensity values
value <- intensity(data.test3)

## make a copy of your object if you do not want to replace it
newdata.test3 <- data.test3

## replace slot data with value
intensity(newdata.test3, "ReplacementData", FALSE) <- value
str(newdata.test3)

## now you can create an ExprTreeSet using the new intensity data
data.rma <- rma(newdata.test3, "ReplacementRMA", tmpdir="", background="none", normalize=TRUE, verbose=FALSE)

## End(Not run)
```

---

intensity2GCplot-methods

*Boxplot of Probe Intensities Stratified by GC Content.*


---

**Description**

Creates a boxplot of probe intensities stratified by GC content.

*Usage*

```
intensity2GCplot(x, treename, which = "", transfo = log2
```

**Arguments**

x	object of class <a href="#">DataTreeSet</a> .
treename	character vector, tree name containing intensities.
which	type of probes to be used, for details see <a href="#">validData</a> .
transfo	a valid function to transform the data, usually “log2”, or “0”.
range	determines how far the plot whiskers extend out from the box.
col	color pair to be used by function <a href="#">colorRampPalette</a> .
...	optional arguments to be passed to <a href="#">intensity2GCplot</a> .

**Details**

Creates a boxplot of probe intensities for treename stratified by GC content for an object of class [DataTreeSet](#).

**Note**

G/C content must first be attached to class [DataTreeSet](#) using method [attachProbeContentGC](#). It is also recommended to attach the probe mask using method [attachMask](#).

**Author(s)**

Christian Stratowa

**See Also**

[plotIntensity2GC](#)

**Examples**

```
## load existing ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

## need to attach probe G/C content and optionally mask
data.test3 <- attachProbeContentGC(data.test3)
data.test3 <- attachMask(data.test3)

if (interactive()) {
  intensity2GCplot(data.test3, treename = "TestA1.cel", which="mm")
}

## optionally remove probe G/C content and mask to free memory
data.test3 <- removeMask(data.test3)
data.test3 <- removeProbeContentGC(data.test3)
```

---

isROOTFile

*Test for ROOT File*

---

**Description**

Test if a file is a valid ROOT file.

**Usage**

```
isROOTFile(filename)
```

**Arguments**

filename            name of ROOT file, including full path.

**Value**

Return TRUE if file filename is a valid [ROOT](#) file.

**Author(s)**

Christian Stratowa

**See Also**[existsROOTFile](#)**Examples**

```
isROOTFile(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
```

---

lowFilter-methods      *Lower Threshold Filter*

---

**Description**

This method initializes the Lower Threshold Filter. The cutoff value defines the lower threshold for allowed expression levels. If e.g. the number of samples lower than this cutoff value is greater than parameter then the corresponding dataframe row is flagged, i.e. `flag = 0`.

The Lower Threshold Filter flags all rows with: `flag = (sum(expression[i] >= cutoff) >= parameter)`

*Usage*

```
lowFilter(object)
lowFilter(object, value)<-
```

**Arguments**

object	object of class <code>PreFilter</code> .
value	character vector <code>c(cutoff, parameter, condition)</code> .

**Details**

The method `lowFilter` initializes the following parameters:

cutoff:	the lower threshold level for the filter.
parameter:	this value depends on the condition used:
condition:	condition="samples": number of samples (default):
	condition="percent": percent of samples.
	condition="mean": mean value of samples.
	condition="percentile": percentile of samples.

**Value**

An initialized `PreFilter` object.

**Author(s)**

Christian Stratowa

**Examples**

```
prefltr <- PreFilter()
lowFilter(prefltr) <- c(4.0, 3, "samples")
str(prefltr)
```



---

madFilter-methods      *Median Absolute Deviation Filter*


---

**Description**

This method initializes the Median Absolute Deviation Filter.  
The MAD Filter flags all rows with: `flag = (mad >= cutoff)`

*Usage*

```
madFilter(object)
madFilter(object, value)<-
```

**Arguments**

`object`            object of class `PreFilter`.  
`value`              numeric vector `c(cutoff, epsilon)`.

**Details**

The method `madFilter` initializes the following parameters:

`cutoff:`    the cutoff level for the filter.  
`epsilon:`    value to replace mean (default is `epsilon=0.01`).

Note, that `epsilon` has no effect on `mad`.

**Value**

An initialized `PreFilter` object.

**Author(s)**

Christian Stratowa

**Examples**

```
prefltr <- PreFilter()
madFilter(prefltr) <- c(0.5,0.01)
str(prefltr)
```

---

madplot-methods      *Array-Array Expression Level Distance Plot*


---

**Description**

A false color display of between arrays distances, computed as the MAD of the M-values of each pair of arrays.

*Usage*

```
madplot(x,            which    = "UnitName",            transfo   = log2,            col        = NULL,            name
```

**Arguments**

<code>x</code>	object of class <a href="#">ExprTreeSet</a> .
<code>which</code>	type of probes to be used, for details see <a href="#">validData</a> .
<code>transfo</code>	a valid function to transform the data, usually “log2”, or “0”.
<code>col</code>	vector of colors for plot, length is number of samples.
<code>names</code>	optional vector of sample names.
<code>sort</code>	logical, if TRUE the correlation matrix will be sorted decreasingly.
<code>bmar</code>	optional list for bottom margin and axis label magnification <code>cex.axis</code> .
<code>add.legend</code>	logical, if TRUE then a color bar will be drawn.
<code>...</code>	optional arguments to be passed to <code>plot</code> .

**Details**

Produces a false color display, i.e. heatmap, of between array distances for slot data for an object of class [ExprTreeSet](#), computed as the MAD of the M-values of each pair of arrays.

For `names=NULL` full column names of slot data will be displayed while for `names="namepart"` column names will be displayed without name extension. If `names` is a vector of column names, only these columns will displayed as `mdaplot`.

For `bmar=NULL` the default list `bmar = list(b=6, cex.axis=1.0)` will be used initially. However, both bottom margin and axis label magnification will be adjusted depending on the number of label characters and the number of samples.

**Author(s)**

Christian Stratowa

**See Also**

[plotMAD](#), [corplot](#)

---

mas4

*MAS 4.0 Expression Measure*

---

**Description**

This function converts a [DataTreeSet](#) into an [ExprTreeSet](#) using the XPS implementation of Affymetrix’s MAS 4.0 expression measure.

**Usage**

```
mas4(xps.data,
     filename = character(0),
     filedir  = getwd(),
     tmpdir   = "",
     normalize = FALSE,
     sc       = 500,
     option   = "transcript",
     exonlevel = "",
```

```

update      = FALSE,
xps.scheme = NULL,
add.data    = TRUE,
verbose     = TRUE)

```

```
xpsMAS4(object, ...)
```

### Arguments

xps.data	object of class <code>DataTreeSet</code> .
filename	file name of ROOT data file.
filedir	system directory where ROOT data file should be stored.
tmpdir	optional temporary directory where temporary ROOT files should be stored.
normalize	logical. If TRUE scale normalization is used after an <code>ExprTreeSet</code> is obtained.
sc	value at which all arrays will be scaled to.
option	option determining the grouping of probes for summarization, one of ‘transcript’, ‘exon’, ‘probeset’; exon arrays only.
exonlevel	exon annotation level determining which probes should be used for summarization; exon/genome arrays only.
update	logical. If TRUE the existing ROOT data file filename will be updated.
xps.scheme	optional alternative <code>SchemeTreeSet</code> .
add.data	logical. If TRUE expression data will be included as slot data.
verbose	logical, if TRUE print status information.
object	object of class <code>DataTreeSet</code> .
...	arguments filename, filedir, tmpdir, option, exonlevel, xps.scheme.

### Details

This function computes the Affymetrix MAS 4.0 expression measure, i.e. the ‘Average Difference’ expression level, as implemented in XPS.

If `normalize=TRUE` then the expression levels will be scaled to `sc`. For `sc=0` the expression levels will be scaled to the mean expression level.

`xpsMAS4` is the `DataTreeSet` method called by function `mas4`, however, expression levels will not be scaled to a common mean expression level.

For further details see [mas5](#).

### Value

An `ExprTreeSet`

### Note

In contrast to function `mas4`, expression levels computed with `xpsMAS4` will not be scaled to a common mean expression level.

### Author(s)

Christian Stratowa

## References

Affymetrix (1999) GeneChip Expression Analysis Algorithm Tutorial, Affymetrix Inc., Santa Clara, CA.

## See Also

[xpsMAS4](#), [express](#)

## Examples

```
## first, load ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

data.mas4 <- mas4(data.test3, "tmp_Test3MAS4", tmpdir="", normalize=TRUE, sc=500, update=TRUE, verbose=FALSE)

## get data.frame
expr.mas4 <- validData(data.mas4)
head(expr.mas4)

## plot results (negative expression values!)
if (interactive()) {
  boxplot(expr.mas4)
}

rm(scheme.test3, data.test3)
gc()
```

---

mas5

*MAS 5.0 Expression Measure*

---

## Description

This function converts a [DataTreeSet](#) into an [ExprTreeSet](#) using the XPS implementation of Affymetrix's MAS 5.0 expression measure.

## Usage

```
mas5(xps.data,
     filename = character(0),
     filedir  = getwd(),
     tmpdir   = "",
     normalize = FALSE,
     sc       = 500,
     option   = "transcript",
     exonlevel = "",
     update   = FALSE,
     xps.scheme = NULL,
     add.data  = TRUE,
     verbose   = TRUE)
```

```
xpsMAS5(object, ...)
```

**Arguments**

xps.data	object of class DataTreeSet.
filename	file name of ROOT data file.
filedir	system directory where ROOT data file should be stored.
tmpdir	optional temporary directory where temporary ROOT files should be stored.
normalize	logical. If TRUE scale normalization is used after an ExprTreeSet is obtained.
sc	value at which all arrays will be scaled to.
option	option determining the grouping of probes for summarization, one of 'transcript', 'exon', 'probeset'; exon arrays only.
exonlevel	exon annotation level determining which probes should be used for summarization; exon/genome arrays only.
update	logical. If TRUE the existing ROOT data file filename will be updated.
xps.scheme	optional alternative SchemeTreeSet.
add.data	logical. If TRUE expression data will be included as slot data.
verbose	logical, if TRUE print status information.
object	object of class DataTreeSet.
...	arguments filename,filedir,tmpdir,option,exonlevel,xps.scheme.

**Details**

This function computes the Affymetrix MAS 5.0 expression measure as implemented in XPS. Although this implementation is based on the Affymetrix 'sadd\_whitepaper.pdf', it can be used to compute an expression level for both expression arrays and exon arrays. For exon arrays it is necessary to supply the requested option and exonlevel.

Following options are valid for exon arrays:

transcript:	expression levels are computed for transcript clusters, i.e. probe sets containing the same 'transcript_cluster'.
exon:	expression levels are computed for exon clusters, i.e. probe sets containing the same 'exon_id', where each
probeset:	expression levels are computed for individual probe sets, i.e. for each 'probeset_id'.

Following exonlevel annotations are valid for exon arrays:

core:	probesets supported by RefSeq and full-length GenBank transcripts.
metacore:	core meta-probesets.
extended:	probesets with other cDNA support.
metaextended:	extended meta-probesets.
full:	probesets supported by gene predictions only.
metafull:	full meta-probesets.
ambiguous:	ambiguous probesets only.
affx:	standard AFFX controls.
all:	combination of above (including affx).

Following exonlevel annotations are valid for whole genome arrays:

core:	probesets with category 'unique', 'similar' and 'mixed'.
metacore:	probesets with category 'unique' only.

affx: standard AFFX controls.  
 all: combination of above (including affx).

Exon levels can also be combined, with following combinations being most useful:

exonlevel="metacore+affx": core meta-probesets plus AFFX controls  
 exonlevel="core+extended": probesets with cDNA support  
 exonlevel="core+extended+full": supported plus predicted probesets

Exon level annotations are described in the Affymetrix whitepaper 'exon\_probeset\_trans\_clust\_whitepaper.pdf'.

If normalize=TRUE then the expression levels will be scaled to sc. For sc=0 the expression levels will be scaled to the mean expression level.

If update=TRUE then the existing ROOT file filename will be updated, however, this is usually only recommended as option for function [express](#).

In order to use an alternative [SchemeTreeSet](#) set the corresponding SchemeTreeSet xps.scheme. xpsMAS5 is the DataTreeSet method called by function mas5, however, expression levels will not be scaled to a common mean expression level.

### Value

An [ExprTreeSet](#)

### Note

In contrast to function mas5, expression levels computed with xpsMAS5 will not be scaled to a common mean expression level.

### Author(s)

Christian Stratowa

### References

Affymetrix (2002) Statistical Algorithms Description Document, Affymetrix Inc., Santa Clara, CA, whitepaper. [http://www.affymetrix.com/support/technical/whitepapers/sadd\\_whitepaper.pdf](http://www.affymetrix.com/support/technical/whitepapers/sadd_whitepaper.pdf)

Affymetrix (2005) Exon Probeset Annotations and Transcript Cluster Groupings, Affymetrix Inc., Santa Clara, CA, exon\_probeset\_trans\_clust\_whitepaper.pdf.

### See Also

[express](#)

### Examples

```
## first, load ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

data.mas5 <- mas5(data.test3, "tmp_Test3MAS5", tmpdir="", normalize=TRUE, sc=500, update=TRUE, verbose=FALSE)
```

```
## get data.frame
expr.mas5 <- validData(data.mas5)
head(expr.mas5)

## plot results
if (interactive()) {
  boxplot(data.mas5)
  boxplot(log2(expr.mas5))
}

rm(scheme.test3, data.test3)
gc()
```

---

mas5.call

*MAS 5.0 Absolute Detection Call*


---

### Description

Performs the Wilcoxon signed rank-based gene expression presence/absence detection algorithm first implemented in the Affymetrix Microarray Suite version 5.

### Usage

```
mas5.call(xps.data,
          filename = character(0), filedir = getwd(), tmpdir = "",
          tau = 0.015, alpha1 = 0.04, alpha2 = 0.06, ignore.saturated = TRUE, bgcorrect.option = "none",
          option = "transcript", exonlevel = "", xps.scheme = NULL, add.data = TRUE, verbose = TRUE)

xpsMAS5Call(object, ...)
```

### Arguments

xps.data	object of class DataTreeSet.
filename	file name of ROOT data file.
filedir	system directory where ROOT data file should be stored.
tmpdir	optional temporary directory where temporary ROOT files should be stored.
tau	a small positive constant.
alpha1	a significance threshold in (0,alpha2).
alpha2	a significance threshold in (alpha1,0.5).
ignore.saturated	logical. If TRUE do the saturation correction described in the paper, with a saturation level of 46000.
bgcorrect.option	bgcorrect option determining whether to subtract background first, one of 'none' or 'correctbg'.
option	option determining the grouping of probes for summarization, one of 'transcript', 'exon', 'probeset'; exon arrays only.
exonlevel	exon annotation level determining which probes should be used for summarization; exon/genome arrays only.

xps.scheme	optional alternative SchemeTreeSet.
add.data	logical. If TRUE call data will be added to slots data and detcall.
verbose	logical, if TRUE print status information.
object	object of class DataTreeSet.
...	the arguments described above.

## Details

This function performs the hypothesis test:

H0: median(Ri) = tau, corresponding to absence of transcript  
 H1: median(Ri) > tau, corresponding to presence of transcript

where  $R_i = (PM_i - MM_i) / (PM_i + MM_i)$  for each  $i$  a probe-pair in the probe-set represented by data.

The p-value that is returned estimates the usual quantity:

Pr(observing a more "present looking" probe-set than data | data is absent)

Small p-values imply presence while large ones imply absence of transcript. The detection call is computed by thresholding the p-value as in:

call "P" if p-value < alpha1

call "M" if alpha1 <= p-value < alpha2

call "A" if alpha2 <= p-value

The defaults for tau, alpha1 and alpha2 correspond to those in MAS5.0 for expression arrays.

However, when using this function for exon or whole genome arrays, new values for alpha1 and alpha2 must be determined. Furthermore, in these cases it may be better to use `bgcorrect.option = "correctbg"` to get reasonable present calls. Note that the recommended function for exon/genome arrays is [dabg.call](#).

In order to use an alternative [SchemeTreeSet](#) set the corresponding SchemeTreeSet `xps.scheme`.

`xpsMAS5Call` is the DataTreeSet method called by function `mas5.call`, containing the same parameters.

## Value

A [CallTreeSet](#)

## Author(s)

Christian Stratowa

## References

Liu, W. M. and Mei, R. and Di, X. and Ryder, T. B. and Hubbell, E. and Dee, S. and Webster, T. A. and Harrington, C. A. and Ho, M. H. and Baid, J. and Smeekens, S. P. (2002) Analysis of high density expression microarrays with signed-rank call algorithms, *Bioinformatics*, 18(12), pp. 1593-1599.

Liu, W. and Mei, R. and Bartell, D. M. and Di, X. and Webster, T. A. and Ryder, T. (2001) Rank-based algorithms for analysis of microarrays, *Proceedings of SPIE, Microarrays: Optical Technologies and Informatics*, 4266.

Affymetrix (2002) Statistical Algorithms Description Document, Affymetrix Inc., Santa Clara, CA, whitepaper. [http://www.affymetrix.com/support/technical/whitepapers/sadd\\_whitepaper.pdf](http://www.affymetrix.com/support/technical/whitepapers/sadd_whitepaper.pdf)



**See Also**[dabg.call](#)**Examples**

```
## first, load ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

## MAS5 detection call
call.mas5 <- mas5.call(data.test3, "tmp_Test3Call", tmpdir="", verbose=FALSE)

## get data.frames
pval.mas5 <- pvalData(call.mas5)
pres.mas5 <- presCall(call.mas5)
head(pval.mas5)
head(pres.mas5)

## plot results
if (interactive()) {
  callplot(call.mas5, beside=FALSE, ylim=c(0,125))
}

rm(scheme.test3, data.test3)
gc()
```

mboxplot-methods

*Box Plots of Relative M Values***Description**

Produce boxplots of relative M values for the set of arrays.

*Usage*

```
mboxplot(x, which = "", size = 0, transfo = log2, method = "mean", range = 0, ylim = c(-1,1), outl
```

**Arguments**

x	object of class <a href="#">DataTreeSet</a> or <a href="#">ExprTreeSet</a> .
which	type of probes to be used, for details see <a href="#">validData</a> .
size	length of sequence to be generated as subset.
transfo	a valid function to transform the data, usually “log2”, or “0”.
method	method to create the reference data, “mean” or “median”.
range	determines how far the plot whiskers extend out from the box.
ylim	range for the plotted y values.
outline	if outline is not true, the outliers are not drawn.
names	optional vector of sample names.
...	optional arguments to be passed to boxplot.

**Details**

Create boxplots of  $M$  plots, where  $M$  is determined relative to a pseudo-mean reference chip.

For `names=NULL` full column names of slot data will be displayed while for `names="namepart"` column names will be displayed without name extension. If `names` is a vector of column names, only these columns will displayed as boxplot.

**Note**

For a `DataTreeSet` object, data must first be attached using method `attachInten`.

**Author(s)**

Christian Stratowa

**See Also**

`mvaplot`, `boxplot`

**Examples**

```
# load existing ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

# need to attach scheme mask and probe intensities
data.test3 <- attachMask(data.test3)
data.test3 <- attachInten(data.test3)

if (interactive()) {
  mboxplot(data.test3, ylim=c(-6,6))
}

# optionally remove mask and data to free memory
data.test3 <- removeInten(data.test3)
data.test3 <- removeMask(data.test3)
```

---

metaProbesets

*Create MetaProbeset File for APT*

---

**Description**

Create MetaProbeset File for APT function “`apt-probeset-summarize`”.

**Usage**

```
metaProbesets(xps.scheme, infile = character(0), outfile = character(0), exonlevel="metacore")
```

**Arguments**

<code>xps.scheme</code>	exon SchemeTreeSet.
<code>infile</code>	Name of file containing exon transcript\_cluster\_ids.
<code>outfile</code>	Name of resulting file containing meta probeset definitions.
<code>exonlevel</code>	exon annotation level determining which probes should be used.

**Details**

This function allows to create a metaprobeset file for APT function “apt-probeset-summarize” to be used with option “-m”. The infile must contain exon transcript\cluster\ids, one per line, e.g. one can export the rownames(data.rma)

The resulting file may be useful if you want to compare results created with xps to results created with APT function “apt-probeset-summarize”.

**Value**

None.

**Author(s)**

Christian Stratowa

**Examples**

```
## Not run:
## first, load ROOT exon scheme file:
scmdir <- "/Volumes/GigaDrive/CRAN/Workspaces/Schemes"
scheme.exon <- root.scheme(paste(scmdir,"Scheme_HuEx10stv2r2_na25.root",sep="/"))

metaProbesets(scheme.exon,"metacore.txt","metacoreList.mps","metacore")

## End(Not run)
```

---

mvaplot-methods

*M vs A Plot*


---

**Description**

Produce scatter plots of M values vs A values of the samples.

*Usage*

```
mvaplot(x,          which = "UnitName",          transfo = log2,          method = "median",          names
```

**Arguments**

x                    object of class [ExprTreeSet](#).

which                type of probes to be used, for details see [validData](#).

transfo             a valid function to transform the data, usually “log2”, or “0”.

method              method to compute M, “mean” or “median”.

names               optional vector of sample names.

ylim                range for the plotted M values.

xlab                a label for the x axis.

ylab                a label for the y axis.

pch                 an integer specifying a symbol or a character to be used as the default in plotting points.

las                 the style of axis labels.

...                 optional arguments to be passed to plot.

**Details**

Produces mvaplots for slot data for an object of class [ExprTreeSet](#).

For names=NULL full column names of slot data will be displayed while for names="namepart" column names will be displayed without name extension. If names is a vector of column names, only these columns will displayed as mvaplot.

**Author(s)**

Christian Stratowa

**See Also**

[plotMA](#)

---

namePart

*Get Tree Names w/o Extension*

---

**Description**

Get (tree) names w/o their extension.

**Usage**

```
namePart(names)
```

**Arguments**

names            vector of names.

**Details**

Extracts the name part of names, e.g.of tree names of treename . treetype stored in a [ROOT](#) file.

**Value**

A vector of tree names w/o its extension.

**Author(s)**

Christian Stratowa

**See Also**

[extenPart](#)

**Examples**

```
names <- c("TestA1.int", "TestA2.int")
namePart(names)
```

---

normalize	<i>Normalization on Affymetrix Probe Level Data or on Expression Levels</i>
-----------	---

---

### Description

Functions that allow to normalize Affymetrix arrays both at the probe level (“low-level normalization”) and/or at the expression level (“high-level normalization”).

### Usage

```
normalize(xps.data, filename = character(0), filedir = getwd(), tmpdir = "", update = FALSE, select
normalize.constant(xps.data, filename = character(0), filedir = getwd(), tmpdir = "", update = FALSE
normalize.lowess(xps.data, filename = character(0), filedir = getwd(), tmpdir = "", update = FALSE
normalize.quantiles(xps.data, filename = character(0), filedir = getwd(), tmpdir = "", update = FALSE
normalize.supsmu(xps.data, filename = character(0), filedir = getwd(), tmpdir = "", update = FALSE
xpsNormalize(object, ...)
```

### Arguments

xps.data	object of class <code>DataTreeSet</code> or <code>ExprTreeSet</code> .
filename	file name of ROOT data file.
filedir	system directory where ROOT data file should be stored.
tmpdir	optional temporary directory where temporary ROOT files should be stored.
update	logical. If TRUE the existing ROOT data file filename will be updated.
select	type of probes to select for normalization.
method	normalization method to use.
option	option determining the grouping of probes for normalization, and the selection of the probes.
logbase	logarithm base as character, one of ‘0’, ‘log’, ‘log2’, ‘log10’.
exonlevel	exon annotation level determining which probes should be used for summarization; exon/genome arrays only.
refindex	index of reference tree to use, or 0.
refmethod	for refindex=0, either trimmed mean or median of trees.
params	vector of parameters for normalization method.
add.data	logical. If TRUE expression data will be included as slot data.
verbose	logical, if TRUE print status information.
object	object of class <code>DataTreeSet</code> or <code>ExprTreeSet</code> .
...	the arguments described above.

**Details**

Functions that allow to normalize Affymetrix arrays both at the probe level (“low-level normalization”) and/or at the expression level (“high-level normalization”).

Please have a look at vignette “xpsPreprocess.pdf” for details on how to use function `normalize`.

`xpsNormalize` are the `DataTreeSet` or `ExprTreeSet` methods, respectively, called by function `normalize`, containing the same parameters.

**Value**

An object of type `DataTreeSet` or `ExprTreeSet`.

**Warning**

Functions `normalize.lowess` and `normalize.supsmu` have only be tested for objects of type `ExprTreeSet` but not for objects of type `DataTreeSet`, i.e. for probe level intensities.

**Author(s)**

Christian Stratowa

**See Also**

[express](#)

**Examples**

```
## first, load ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

## RMA background
data.bg.rma <- bgcorrect.rma(data.test3, "tmp_Test3NormRMA", filedir=getwd(), tmpdir="", verbose=FALSE)
## normalize quantiles
data.qu.rma <- normalize.quantiles(data.bg.rma, "tmp_Test3NormRMA", filedir=getwd(), tmpdir="", update=TRUE, verbose=FALSE)
## summarize medianpolish
data.mp.rma <- summarize.rma(data.qu.rma, "tmp_Test3NormRMA", filedir=getwd(), tmpdir="", update=TRUE, verbose=FALSE)
```

---

NUSE-methods

*Normalized Unscaled Standard Errors (NUSE)*

---

**Description**

Produce boxplot of Normalized Unscaled Standard Errors (NUSE) for the set of arrays. Alternatively, summary statistics or NUSE values can be extracted as `data.frame`.

*Usage*

```
NUSE(x, treename = "*", type = c("plot", "stats", "values"), qualopt = NULL,
```

**Arguments**

x	object of class <a href="#">QualTreeSet</a> .
treename	vector of tree names to export.
type	type of output, plot, stats or values.
qualopt	quality control option, i.e. 'raw', 'adjusted', 'normalized' or 'all'.
...	optional arguments to be passed to nuseplot.

**Details**

Create boxplots of Normalized Unscaled Standard Errors (NUSE) for the set of arrays.

Alternatively it is possible to extract either the summary statistics as `data.frame (type="stats")` or all NUSE values as `data.frame (type="values")`.

If an object of class [QualTreeSet](#) was created by fitting a probe level model with `qualopt="all"` then NUSE will plot or extract NUSE for "all" quality options. If you want to plot or extract NUSE for a certain quality option only, e.g. "normalized" data only, then you can use parameter `qualopt` with `qualopt="<qualopt>"`.

**Author(s)**

Christian Stratowa

**See Also**

[plotNUSE](#), [nuseplot](#)

**Examples**

```
## Not run:
## load existing ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

## qualification - rlm
rlm.all <- rmaPLM(data.test3, "tmp_Test3RLMall", filedir=getwd(), tmpdir="", qualopt="all", option="transcrip

## plot expression levels
if (interactive()) {
  NUSE(rlm.all)
  NUSE(rlm.all, qualopt="normalized")
  qcNUSE <- NUSE(rlm.all, type="stats")
  qcNUSE <- NUSE(rlm.all, type="values")
  qcNUSE <- NUSE(rlm.all, treename="TestA1_normalized.rlm", type="stats")
  qcNUSE <- NUSE(rlm.all, treename="TestA1_normalized.rlm", type="values")
}

## End(Not run)
```

---

nuseplot-methods      *Box Plots of Normalized Unscaled Standard Errors (NUSE)*

---

### Description

Produce boxplot of Normalized Unscaled Standard Errors (NUSE) for the set of arrays.

#### Usage

```
nuseplot(x, which = "UnitName", size = 0, range = 0, names = )
```

### Arguments

x	object of class or <a href="#">QualTreeSet</a> .
which	type of probes to be used, for details see <a href="#">validData</a> (only ExprTreeSet).
size	length of sequence to be generated as subset (only ExprTreeSet).
range	determines how far the plot whiskers extend out from the box.
names	optional vector of sample names.
main	the main title for the plot.
ylim	range for the plotted y values.
las	the style of axis labels.
add.line	logical, if TRUE a horizontal line is drawn.
outline	if outline is not true, the outliers are not drawn (only ExprTreeSet).
...	optional arguments to be passed to boxplot.

### Details

Create boxplots of Normalized Unscaled Standard Errors (NUSE) for the set of arrays.

For names=NULL full column names of slot data will be displayed while for names="namepart" column names will be displayed without name extension. If names is a vector of column names, only these columns will displayed as boxplot.

If an object of class [QualTreeSet](#) was created by fitting a probe level model with qualopt="all" then nuseplot will plot NUSE for "all" quality options. If you want to plot NUSE for a certain quality option only, e.g. "normalized" data only, then you can use parameter names with names="namepart:<qualopt>", e.g. names="namepart:normalized".

### Author(s)

Christian Stratowa

### See Also

[NUSE](#), [plotNUSE](#), [rleplot](#)



**Examples**

```
# load existing ROOT scheme file and ROOT expression file for rma
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.rma <- root.expr(scheme.test3, paste(path.package("xps"), "rootdata/tmp_Test3RMA.root", sep="/"), "mdp")

if (interactive()) {
  nuseplot(data.rma)
}
```

pcaplot-methods

*PCA Plot***Description**

This function produces a PCA plot of the first two principle components.

*Usage*

```
pcaplot(x, which = "UnitName", transfo = log2, method = "none", g
```

**Arguments**

x	object of class <a href="#">ExprTreeSet</a> .
which	type of probes to be used, for details see <a href="#">validData</a> .
transfo	a valid function to transform the data, usually "log2", or "0".
method	a character string indicating which correlation coefficient is to be computed. One of "pearson", "spearman", "kendall", or "none".
groups	character vector listing the group names in order of the names.
screepplot	logical, if TRUE plot a <a href="#">screepplot</a> instead of a PCA plot.
squarepca	logical, if TRUE make the y-axis of the PCA plot comparable to the x-axis.
pcs	a character vector of length two indicating which principal components to plot.
add.labels	logical, if TRUE then name labels will be added to the points.
add.legend	logical, if TRUE and groups are supplied then a legend indicating the groups will be drawn. Optionally, a character indicating the position of the legend, default is "topleft".
col	vector of colors for plot, length is number of samples.
names	optional vector of sample names.
as.list	logical, if TRUE then a list will be returned in addition to the plot.
...	optional arguments to be passed to plot.

**Details**

Function pcaplot produces a PCA plot of the first two principle components for slot data or the correlations between the columns of slot data, respectively, of an object of class [ExprTreeSet](#).

For method="none" function [stats]prcomp will be applied to slot data directly, otherwise prcomp will be applied to (1 - cor(data)) with the respective method.

For screepplot=TRUE a [screepplot](#) will be plotted instead of a PCA plot.

For names=NULL full column names of slot data will be displayed while for names="namepart" column names will be displayed without name extension. If names is a vector of column names, only these columns will displayed as mvaplot.

**Value**

None by default.

Optionally, for `as.list=TRUE` a list will be returned with the components `sdev` and `rotation`, see `[stats]prcomp`.

**Author(s)**

Christian Stratowa, partly adapted from function `plotPCA()` of package `affycoretools`

**See Also**

[plotPCA](#), [corplot](#) [madplot](#)

---

plotBorder

*Plots of Border Elements for Device*

---

**Description**

Produce box-and-whisker plot(s) of the positive and negative feature intensities for the selected device.

**Usage**

```
plotBorder(x,
           type = c("pos", "neg"),
           qualopt = "raw",
           transfo = log2,
           range = 0,
           names = "namepart",
           ylim = NULL,
           bmar = NULL,
           las = 2,
           dev = "screen",
           outfile = "BorderPlot",
           w = 800,
           h = 540,
           ...)
```

**Arguments**

<code>x</code>	object of class <a href="#">QualTreeSet</a> .
<code>type</code>	type of border elements to be used, one of “pos”, “neg”, or both.
<code>qualopt</code>	character string specifying whether to draw boxplots for “raw”, “adjusted”, or “normalized” border intensities.
<code>transfo</code>	a valid function to transform the data, usually “log2”, or “0”.
<code>range</code>	determines how far the plot whiskers extend out from the box.
<code>names</code>	optional vector of sample names.
<code>ylim</code>	the y limits of the plot.
<code>bmar</code>	optional list for bottom margin and axis label magnification <code>cex.axis</code> .

las	the style of axis labels.
dev	graphics device to plot to, i.e. one of "screen", "jpeg", "png", "pdf" or "ps".
outfile	the name of the output file.
w	the width of the device in pixels.
h	the height of the device in pixels.
...	optional arguments to be passed to borderplot.

### Details

Creates a boxplot of the positive and negative feature intensities for an object of class [QualTreeSet](#).

For names=NULL full tree names will be displayed while for names="namepart" tree names will be displayed without name extension. If names is a vector of tree names, only these columns will be displayed as boxplot.

For bmar=NULL the default list `bmar = list(b=6, cex.axis=1.0)` will be used initially. However, both bottom margin `b` and axis label magnification `cex.axis` will be adjusted depending on the number of label characters and the number of samples.

### Author(s)

Christian Stratowa

### See Also

[borderplot](#)

---

plotBoxplot	<i>Box Plots for Device</i>
-------------	-----------------------------

---

### Description

Produce box-and-whisker plot(s) of the samples for the selected device.

### Usage

```
plotBoxplot(x,
            which = "",
            size  = 0,
            transfo = log2,
            range = 0,
            names = "namepart",
            mar   = NULL,
            las   = 2,
            cex   = 1.0,
            dev   = "screen",
            outfile = "BoxPlot",
            w     = 800,
            h     = 540,
            ...)
```

**Arguments**

x	object of class <a href="#">DataTreeSet</a> or <a href="#">ExprTreeSet</a> .
which	type of probes to be used, for details see <a href="#">validData</a> .
size	length of sequence to be generated as subset.
transfo	a valid function to transform the data, usually <code>log2</code> , or 0.
range	determines how far the plot whiskers extend out from the box.
names	optional vector of sample names.
mar	plot margin.
las	style of axis labels.
cex	amount by which plotting text and symbols should be magnified.
dev	graphics device to plot to, i.e. one of "screen", "jpeg", "png", "pdf" or "ps".
outfile	the name of the output file.
w	the width of the device in pixels.
h	the height of the device in pixels.
...	optional arguments to be passed to <code>boxplot</code> .

**Details**

Produces a boxplot for slot data for an object of class [DataTreeSet](#), [ExprTreeSet](#) or [QualTreeSet](#) for the selected graphics device.

**Author(s)**

Christian Stratowa

**See Also**

[boxplot](#), [plotBorder](#), [plotNUSE](#), [plotRLE](#)

---

plotCall

*Barplot of Percent Present and Absent Calls for Device*

---

**Description**

Creates a barplot of percent Present/Marginal/Absent calls for the selected device.

**Usage**

```
plotCall(x,
         beside = TRUE,
         names = "namepart",
         col = c("red", "green", "blue"),
         legend = c("P", "M", "A"),
         ylim = c(0, 100),
         ylab = "detection call [%]",
         las = 2,
         dev = "screen",
```

```

outfile = "CallPlot",
w       = 800,
h       = 540,
...)
```

### Arguments

x	object of class <a href="#">CallTreeSet</a> .
beside	logical. If FALSE, the columns of height are portrayed as stacked bars, and if TRUE the columns are portrayed as juxtaposed bars.
names	optional vector of sample names.
col	color for P/M/A bars
legend	legend for the plot, defaults to P/M/A.
ylim	the y limits of the plot.
ylab	a label for the y axis.
las	the style of axis labels.
dev	graphics device to plot to, i.e. one of "screen", "jpeg", "png", "pdf" or "ps".
outfile	the name of the output file.
w	the width of the device in pixels.
h	the height of the device in pixels.
...	optional arguments to be passed to <code>barplot</code> .

### Details

Creates a barplot of percent Present/Marginal/Absent calls.

For `names=NULL` full column names of slot data will be displayed while for `names="namepart"` column names will be displayed without name extension. If `names` is a vector of column names, only these columns will displayed as callplot.

### Author(s)

Christian Stratowa

### See Also

[callplot](#)

---

plotCOI

*Center-Of-Intensity QC Plots for Device*

---

### Description

Produce Center-Of-Intensity plot(s) of the positive and negative feature intensities for the selected device.

**Usage**

```
plotCOI(x,
        type    = c("pos", "neg"),
        qualopt = "raw",
        radius  = 0.5,
        linecol = "gray70",
        visible = TRUE,
        dev     = "screen",
        outfile = "CenterOfIntensityPlot",
        w      = 540,
        h      = 540,
        ...)
```

**Arguments**

x	object of class <a href="#">QualTreeSet</a> .
type	type of border elements to be used, one of “pos”, “neg”, or both.
qualopt	character string specifying whether to draw boxplots for “raw”, “adjusted”, or “normalized” border intensities.
radius	determines the radius within which the COI for each array should be located.
linecol	the color of the ablines and the circle to be drawn.
visible	logical, if TRUE then arrays outside the circle with radius will be flagged by labeling the data point with the array name.
dev	graphics device to plot to, i.e. one of “screen”, “jpeg”, “png”, “pdf” or “ps”.
outfile	the name of the output file.
w	the width of the device in pixels.
h	the height of the device in pixels.
...	optional arguments to be passed to <code>coiplot</code> .

**Details**

Produces Center-Of-Intensity (COI) plot(s) of the positive and negative feature intensities for an object of class [QualTreeSet](#). This plot is useful for detecting spatial biases in intensities on an array.

Mean intensities for the left, right, top and bottom border elements are calculated, separated into positive and negative controls, and the “center of intensity” is calculated on a relative scale [-1,1]. Arrays with a COI outside a range with radius are considered to be outliers. If `visible = TRUE` then outlier arrays will be flagged by labeling the data point(s) with the array name(s).

**Author(s)**

Christian Stratowa

**See Also**

[coiplot](#)

---

plotCorr

*Array-Array Expression Level Correlation Plot for Device*


---

### Description

A heat map of the array-array Spearman rank correlation coefficients for the selected device.

### Usage

```
plotCorr(x,
         which      = "UnitName",
         transfo    = log2,
         method     = "spearman",
         col        = NULL,
         names      = "namepart",
         sort       = FALSE,
         reverse    = TRUE,
         bmar       = NULL,
         add.legend = FALSE,
         dev        = "screen",
         outfile    = "CorrelationPlot",
         w         = 540,
         h         = 540,
         ...)
```

### Arguments

x	object of class <a href="#">ExprTreeSet</a> .
which	type of probes to be used, for details see <a href="#">validData</a> .
transfo	a valid function to transform the data, usually "log2", or "0".
method	a character string indicating which correlation coefficient is to be computed.
col	vector of colors for plot, length is number of samples.
names	optional vector of sample names.
sort	logical, if TRUE the correlation matrix will be sorted decreasingly.
reverse	logical, if TRUE the correlation matrix will be replaced by $1 - \text{cor}()$ .
bmar	optional list for bottom margin and axis label magnification <code>cex.axis</code> .
add.legend	logical, if TRUE then a color bar will be drawn.
dev	graphics device to plot to, i.e. one of "screen", "jpeg", "png", "pdf" or "ps".
outfile	the name of the output file.
w	the width of the device in pixels.
h	the height of the device in pixels.
...	optional arguments to be passed to plot.

**Details**

Produces a heat map of the array-array Spearman rank correlation coefficients for slot data for an object of class [ExprTreeSet](#).

For names=NULL full column names of slot data will be displayed while for names="namepart" column names will be displayed without name extension. If names is a vector of column names, only these columns will displayed as corplot.

For bmar=NULL the default list `bmar = list(b=6, cex.axis=1.0)` will be used initially. However, both bottom margin and axis label magnification will be adjusted depending on the number of label characters and the number of samples.

**Author(s)**

Christian Stratowa

**See Also**

[corplot](#)

---

plotDensity

*Plot Density Estimate for Device*

---

**Description**

Plot the density estimates for each sample for the selected device.

**Usage**

```
plotDensity(x,
            which = "",
            size = 0,
            transfo = log2,
            ylab = "density",
            xlab = "log intensity",
            names = "namepart",
            type = "l",
            col = 1:6,
            lty = 1:5,
            add.legend = FALSE,
            dev = "screen",
            outfile = "DensityPlot",
            w = 540,
            h = 540,
            verbose = TRUE,
            ...)
```

**Arguments**

x                    object of class [DataTreeSet](#) or [ExprTreeSet](#).  
 which                type of probes to be used, for details see [validData](#).  
 size                  length of sequence to be generated as subset.



transfo	a valid function to transform the data, usually “log2”, or “0”.
xlab	a title for the x axis.
ylab	a title for the y axis.
names	optional vector of sample names.
type	type for the plot.
col	colors to use for the different arrays.
lty	line types to use for the different arrays.
add.legend	logical, if TRUE then a legend will be drawn.
dev	graphics device to plot to, i.e. one of “screen”, “jpeg”, “png”, “pdf” or “ps”.
outfile	the name of the output file.
w	the width of the device in pixels.
h	the height of the device in pixels.
verbose	logical, if TRUE print status information.
...	optional arguments to be passed to plot.

### Details

Plots the non-parametric density estimates for each sample.

For names=NULL full column names of slot data will be displayed while for names="namepart" column names will be displayed without name extension. If names is a vector of column names, only these columns will displayed as callplot.

### Author(s)

Christian Stratowa

### See Also

[hist](#)

---

plotImage	<i>Plot Image(s) for Device</i>
-----------	---------------------------------

---

### Description

Creates an image for each sample for the selected device.

### Usage

```
plotImage(x,
          type      = character(),
          qualopt   = c("raw", "adjusted", "normalized"),
          transfo   = log2,
          col       = NULL,
          names     = character(),
          dev       = "screen",
          outfile   = "Image",
          w         = 800,
          h         = 800,
          verbose   = TRUE,
          ...)
```

**Arguments**

x	object of class <a href="#">DataTreeSet</a> or <a href="#">QualTreeSet</a> .
type	character string specifying the type of image.
qualopt	character string specifying whether to draw residual image for “raw”, “adjusted”, or “normalized” intensities.
transfo	a valid function to transform the data, usually “log2”, or “0”.
col	color range for intensities.
names	vector of sample names.
dev	graphics device to plot to, i.e. one of “screen”, “jpeg”, “png”, “pdf” or “ps”.
outfile	the name of the output file.
w	the width of the device in pixels.
h	the height of the device in pixels.
verbose	logical, if TRUE print status information.
...	optional arguments to be passed to <code>image</code> .

**Details**

Creates intensity image(s) or residual image(s), respectively, for each array for the selected graphics device, see [image](#) for more details.

For intensity image(s) type must be one of “intensity”.

For residual image(s) type must be one of “resids”, “pos.resids”, “neg.resids”, “sign.resids”, or “weights”. Furthermore, qualopt determines if images should be drawn for “raw”, “adjusted”, or “normalized” data.

For names="\*" names of all samples will be displayed as images. If names is a vector of column names, only these samples will displayed as image(s).

**Author(s)**

Christian Stratowa

**See Also**

[image-methods](#), [image](#)

**Examples**

```
## Not run:
## first, load ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

## qualification - rlm
rlm.all <- rmaPLM(data.test3, "tmp_Test3RLMall", filedir=getwd(), tmpdir="", qualopt="all", option="transcrip

if (interactive()) {
## image(s) of raw data
plotImage(data.test3, type="intensity", names="*")
plotImage(data.test3, type="intensity", names="TestA2.cel")

## image(s) of residuals/weights
```

```

plotImage(rlm.all, type="weights", names="*")
plotImage(rlm.all, type="weights", qualopt="adjusted", names="*")
plotImage(rlm.all, type="resids", names="TestA2_raw.res")
}

## function image.dev() will be deprecated since it needs attachInten!!
## need to attach scheme mask and data
data.test3 <- attachMask(data.test3)
data.test3 <- attachInten(data.test3)
if (interactive()) {
  image.dev(data.test3)
}
## to avoid memory consumption of R remove data:
data.test3 <- removeInten(data.test3)
data.test3 <- removeMask(data.test3)

## End(Not run)

```

---

plotIntensity2GC

*Boxplot of Probe Intensities Stratified by GC Content for Device.*


---

## Description

Creates a boxplot of probe intensities stratified by GC content for the selected device.

## Usage

```

plotIntensity2GC(x,
                 treename,
                 which = "",
                 transfo = log2,
                 range = 0,
                 col = c("lightblue", "darkblue"),
                 dev = "screen",
                 outfile = "Intensity2GCPlot",
                 w = 540,
                 h = 540,
                 ...)

```

## Arguments

x	object of class <a href="#">DataTreeSet</a> .
treename	character vector, tree name used for intensities.
which	type of probes to be used, for details see <a href="#">validData</a> .
transfo	a valid function to transform the data, usually "log2", or "0".
range	determines how far the plot whiskers extend out from the box.
col	color pair to be used by function <a href="#">colorRampPalette</a> .
dev	graphics device to plot to, i.e. one of "screen", "jpeg", "png", "pdf" or "ps".
outfile	the name of the output file.
w	the width of the device in pixels.
h	the height of the device in pixels.
...	optional arguments to be passed to <a href="#">plotIntensity2GC</a> .

**Details**

Creates a boxplot of probe intensities for treename stratified by GC content for an object of class [DataTreeSet](#).

**Note**

G/C content must first be attached to class [DataTreeSet](#) using method [attachProbeContentGC](#). It is also recommended to attach the probe mask using method [attachMask](#).

**Author(s)**

Christian Stratowa

**See Also**

[intensity2GCplot](#)

---

plotMA

*MvA Scatter Plot for Device*

---

**Description**

Produce scatter plots of M values vs A values of the samples for the selected device.

**Usage**

```
plotMA(x,
       transfo = log2,
       method  = "median",
       names   = "namepart",
       ylim    = c(-6, 6),
       xlab    = "A",
       ylab    = "M",
       pch     = ".",
       mar     = c(3, 3, 2, 1),
       dev     = "screen",
       outfile = "MvAPlot",
       w       = 540,
       h       = 540,
       ...)
```

**Arguments**

x	object of class <a href="#">ExprTreeSet</a> .
transfo	a valid function to transform the data, usually "log2", or "0".
method	method to compute M, "mean" or "median".
names	optional vector of sample names.
ylim	range for the plotted M values.
xlab	a title for the x axis.

ylab	a title for the y axis.
pch	either an integer specifying a symbol or a single character to be used in plotting points.
mar	plot margin.
dev	graphics device to plot to, i.e. one of "screen", "jpeg", "png", "pdf" or "ps".
outfile	the name of the output file.
w	the width of the device in pixels.
h	the height of the device in pixels.
...	optional arguments to be passed to plot.

### Details

Produces M vs A plots for slot data for an object of class [ExprTreeSet](#) for the selected graphics device.

For names=NULL full column names of slot data will be displayed while for names="namepart" column names will be displayed without name extension. If names is a vector of column names, only these columns will displayed as M vs A plot.

### Author(s)

Christian Stratowa

### See Also

[mvaplot](#)

---

plotMAD

*Array-Array Expression Level Distance Plot for Device*

---

### Description

A false color display of between arrays distances, computed as the MAD of the M-values of each pair of arrays for the selected device.

### Usage

```
plotMAD(x,
        which      = "UnitName",
        transfo    = log2,
        col        = NULL,
        names      = "namepart",
        sort       = FALSE,
        bmar       = NULL,
        add.legend = FALSE,
        dev        = "screen",
        outfile    = "MADPlot",
        w         = 540,
        h         = 540,
        ...)
```

**Arguments**

x	object of class <a href="#">ExprTreeSet</a> .
which	type of probes to be used, for details see <a href="#">validData</a> .
transfo	a valid function to transform the data, usually “log2”, or “0”.
col	vector of colors for plot, length is number of samples.
names	optional vector of sample names.
sort	logical, if TRUE the correlation matrix will be sorted decreasingly.
bmar	optional list for bottom margin and axis label magnification <code>cex.axis</code> .
add.legend	logical, if TRUE then a color bar will be drawn.
dev	graphics device to plot to, i.e. one of “screen”, “jpeg”, “png”, “pdf” or “ps”.
outfile	the name of the output file.
w	the width of the device in pixels.
h	the height of the device in pixels.
...	optional arguments to be passed to plot.

**Details**

Produces a false color display, i.e. heatmap, of between array distances for slot data for an object of class [ExprTreeSet](#), computed as the MAD of the M-values of each pair of arrays.

For `names=NULL` full column names of slot data will be displayed while for `names="namepart"` column names will be displayed without name extension. If `names` is a vector of column names, only these columns will displayed as `mdaplot`.

For `bmar=NULL` the default list `bmar = list(b=6, cex.axis=1.0)` will be used initially. However, both bottom margin and axis label magnification will be adjusted depending on the number of label characters and the number of samples.

**Author(s)**

Christian Stratowa

**See Also**

[mdaplot](#)

---

plotNUSE

*Box Plots of Normalized Unscaled Standard Errors (NUSE) for Device*

---

**Description**

Produce boxplot of Normalized Unscaled Standard Errors (NUSE) for the set of arrays and the selected device.

## Usage

```
plotNUSE(x,  
         which = "UnitName",  
         size  = 0,  
         range = 0,  
         names = "namepart",  
         main  = "NUSE Plot",  
         ylim  = c(0.8,1.2),  
         las   = 2,  
         add.line = TRUE,  
         outline = FALSE,  
         dev    = "screen",  
         outfile = "NUSEPlot",  
         w      = 800,  
         h      = 540,  
         ...)
```

## Arguments

x	object of class <a href="#">ExprTreeSet</a> or <a href="#">QualTreeSet</a> .
which	type of probes to be used, for details see <a href="#">validData</a> .
size	length of sequence to be generated as subset.
range	determines how far the plot whiskers extend out from the box.
names	optional vector of sample names.
main	the main title for the plot.
ylim	range for the plotted y values.
las	the style of axis labels.
add.line	logical, if TRUE a horizontal line is drawn.
outline	if outline is not true, the outliers are not drawn.
dev	graphics device to plot to, i.e. one of "screen", "jpeg", "png", "pdf" or "ps".
outfile	the name of the output file.
w	the width of the device in pixels.
h	the height of the device in pixels.
...	optional arguments to be passed to <code>boxplot</code> .

## Details

Create boxplots of Normalized Unscaled Standard Errors (NUSE) for the set of arrays.

For `names=NULL` full column names of slot data will be displayed while for `names="namepart"` column names will be displayed without name extension. If `names` is a vector of column names, only these columns will be displayed as boxplot.

## Author(s)

Christian Stratowa

## See Also

[nuseplot](#)

plotPCA

*PCA Plot for Device***Description**

This function produces a PCA plot of the first two principle components for the selected device.

**Usage**

```
plotPCA(x,
        which      = "UnitName",
        transfo    = log2,
        method     = "none",
        groups     = NULL,
        screeplot  = FALSE,
        squarepca  = FALSE,
        pcs        = c(1,2),
        add.labels = FALSE,
        add.legend  = FALSE,
        col        = NULL,
        names      = "namepart",
        as.list    = FALSE,
        dev        = "screen",
        outfile    = "PCAPlot",
        w          = 540,
        h          = 540,
        ...)
```

**Arguments**

x	object of class <a href="#">ExprTreeSet</a> .
which	type of probes to be used, for details see <a href="#">validData</a> .
transfo	a valid function to transform the data, usually “log2”, or “0”.
method	a character string indicating which correlation coefficient is to be computed. One of “pearson”, “spearman”, “kendall”, or “none”.
groups	character vector listing the group names in order of the names.
screeplot	logical, if TRUE plot a <a href="#">screeplot</a> instead of a PCA plot.
squarepca	logical, if TRUE make the y-axis of the PCA plot comparable to the x-axis.
pcs	a character vector of length two indicating which principal components to plot.
add.labels	logical, if TRUE then name labels will be added to the points.
add.legend	logical, if TRUE and groups are supplied then a legend indicating the groups will be drawn. Optionally, a character indicating the position of the legend, default is “topleft”.
col	vector of colors for plot, length is number of samples.
names	optional vector of sample names.
as.list	logical, if TRUE then a list will be returned in addition to the plot.
dev	graphics device to plot to, i.e. one of “screen”, “jpeg”, “png”, “pdf” or “ps”.



outfile        the name of the output file.  
 w              the width of the device in pixels.  
 h              the height of the device in pixels.  
 ...            optional arguments to be passed to plot.

### Details

Function plotPCA produces a PCA plot of the first two principle components for slot data or the correlations between the columns of slot data, respectively, of an object of class [ExprTreeSet](#).

For method="none" function [stats]prcomp will be applied to slot data directly, otherwise prcomp will be applied to (1 - cor(data)) with the respective method.

For screeplot=TRUE a [screeplot](#) will be plotted instead of a PCA plot.

For names=NULL full column names of slot data will be displayed while for names="namepart" column names will be displayed without name extension. If names is a vector of column names, only these columns will displayed as mvaplot.

### Author(s)

Christian Stratowa

### See Also

[pcaplot](#)

---

plotPM

*Barplot of PM and MM Intensities for Device*

---

### Description

Creates a barplot of mean perfect match and mismatch intensities for the selected device.

### Usage

```

plotPM(x,
       which = "",
       size  = 0,
       transfo = NULL,
       method = mean,
       names  = "namepart",
       beside = TRUE,
       col    = c("red", "blue"),
       legend = c("PM", "MM"),
       las    = 2,
       ylab   = "mean intensities",
       dev    = "screen",
       outfile = "PMPlot",
       w      = 540,
       h      = 540,
       ...)
```

**Arguments**

x	object of class <a href="#">DataTreeSet</a> .
which	type of probes to be used, for details see <a href="#">validData</a> .
size	length of sequence to be generated as subset.
transfo	a valid function to transform the data, usually “log2”, or “0”.
method	method to compute average intensities, “mean” or “median”.
names	optional vector of sample names.
beside	logical. If FALSE, mean intensities are portrayed as stacked bars, and if TRUE the columns are portrayed as juxtaposed bars.
col	color of PM, MM bars.
legend	a vector of text used to construct a legend for the plot, or a logical indicating whether a legend should be included.
las	the style of axis labels.
ylab	a title for the y axis.
dev	graphics device to plot to, i.e. one of “screen”, “jpeg”, “png”, “pdf” or “ps”.
outfile	the name of the output file.
w	the width of the device in pixels.
h	the height of the device in pixels.
...	optional arguments to be passed to barplot.

**Details**

Produces barplots of mean perfect match and mismatch intensities for slot data for an object of class [DataTreeSet](#).

For names=NULL full column names of slot data will be displayed while for names="namepart" column names will be displayed without name extension. If names is a vector of column names, only these columns will displayed as pmplot.

**Author(s)**

Christian Stratowa

**See Also**

[pmplot](#)

---

plotProbeset

*Plot of Probe Intensities for a Probeset for Device.*

---

**Description**

Creates a line plot of probe intensities for a probeset for the selected device.

**Usage**

```
plotProbeset(x,
             unitID,
             unittype = "transcript",
             which     = "pm",
             transfo   = log2,
             names     = "namepart",
             ylim      = NULL,
             col       = 1:6,
             lty       = 1:5,
             add.legend = FALSE,
             dev       = "screen",
             outfile   = "ProbesetPlot",
             w         = 540,
             h         = 540,
             ...)
```

**Arguments**

x	object of class <a href="#">DataTreeSet</a> .
unitID	unit ID of probeset with type of ID determined by parameter <code>unittype</code> .
unittype	character vector, one of "unit", "transcript", "probeset".
which	type of probes to be used, for details see <a href="#">validData</a> .
transfo	a valid function to transform the data, usually "log2", or "0".
names	optional vector of sample names.
ylim	range for the plotted y values.
col	color to use for the different samples.
lty	line types to use for the different samples.
add.legend	logical, if TRUE a legend of sample names will be drawn. Optionally, a character indicating the position of the legend, default is "topleft".
dev	graphics device to plot to, i.e. one of "screen", "jpeg", "png", "pdf" or "ps".
outfile	the name of the output file.
w	the width of the device in pixels.
h	the height of the device in pixels.
...	optional arguments to be passed to <code>plotProbeset</code> .

**Details**

Produces line plots of the probe intensities for probeset `unitID`. Probe intensities are taken from slot data.

For `names=NULL` full column names of slot data will be displayed while for `names="namepart"` column names will be displayed without name extension. If `names` is a vector of column names, line plots of probe intensities will only be drawn for these columns.

**Note**

Data must first be attached to class [DataTreeSet](#) using method [attachInten](#). Furthermore, unit names must be attached using method [attachUnitNames](#).

**Author(s)**

Christian Stratowa

**See Also**[probesetplot](#)

plotRLE

*Box Plots of Relative Log Expression (RLE) for Device***Description**

Produce boxplot of Relative Log Expression (RLE) for the set of arrays and the selected device.

**Usage**

```
plotRLE(x,
        which = "UnitName",
        size = 0,
        range = 0,
        names = "namepart",
        main = "RLE Plot",
        ylim = c(-1.0, 1.0),
        las = 2,
        add.line = TRUE,
        outline = FALSE,
        dev = "screen",
        outfile = "RLEPlot",
        w = 800,
        h = 540,
        verbose = TRUE,
        ...)
```

**Arguments**

x	object of class <a href="#">ExprTreeSet</a> or <a href="#">QualTreeSet</a> .
which	type of probes to be used, for details see <a href="#">validData</a> .
size	length of sequence to be generated as subset.
range	determines how far the plot whiskers extend out from the box.
names	optional vector of sample names.
main	the main title for the plot.
ylim	range for the plotted y values.
las	the style of axis labels.
add.line	logical, if TRUE a horizontal line is drawn.
outline	if outline is not true, the outliers are not drawn.
dev	graphics device to plot to, i.e. one of "screen", "jpeg", "png", "pdf" or "ps".
outfile	the name of the output file.

w	the width of the device in pixels.
h	the height of the device in pixels.
verbose	logical, if TRUE print status information.
...	optional arguments to be passed to boxplot.

### Details

Create boxplots of Relative Log Expression (RLE) values for the set of arrays, i.e. of M plots, where M is determined relative to a pseudo-median reference chip.

For names=NULL full column names of slot data will be displayed while for names="namepart" column names will be displayed without name extension. If names is a vector of column names, only these columns will displayed as boxplot.

### Author(s)

Christian Stratowa

### See Also

[rleplot](#)

---

plotVolcano

*Volcano Plot*

---

### Description

Produce a scatter plot of fold-change values vs p-values, called volcano plot.

### Usage

```
plotVolcano(x,  
            labels      = "",  
            p.value     = "pval",  
            mask        = FALSE,  
            show.cutoff = TRUE,  
            cex.text    = 0.7,  
            col.text    = "blue",  
            col.cutoff  = "grey",  
            xlim        = NULL,  
            xlab        = "Log2(Fold-Change)",  
            ylab        = "-Log10(P-Value)",  
            pch         = '.',  
            dev         = "screen",  
            outfile     = "VolcanoPlot",  
            w           = 540,  
            h           = 540,  
            ...)
```

**Arguments**

x	object of class <a href="#">AnalysisTreeSet</a> .
labels	optional transcript labels to be drawn at plotting points.
p.value	type of p-value, 'pval' for p-value, 'padj' for adjusted p-value, or 'pcha' for p-chance.
mask	logical, if TRUE draw only points for transcripts satisfying the univariate test.
show.cutoff	logical, if TRUE draw lines indicating cutoff.
cex.text	magnification to be used for optional labels.
col.text	color to be used for optional labels.
col.cutoff	color to be used for lines indicating cutoff, if show.cutoff=TRUE.
xlim	optional range for the plotted fold-change values.
xlab	label of x-axis.
ylab	label of y-axis.
pch	either an integer specifying a symbol or a single character to be used as the default in plotting points.
dev	graphics device to plot to, i.e. one of "screen", "jpeg", "png", "pdf" or "ps".
outfile	the name of the output file.
w	the width of the device in pixels.
h	the height of the device in pixels.
...	optional arguments to be passed to <code>barplot</code> .

**Details**

Produces a volcano plot for slot data for an object of class [AnalysisTreeSet](#).

It is possible to label the points of the volcano plot, whereby the following labels parameters are valid:

fUnitName:	unit name (probeset ID).
fName:	gene name.
fSymbol:	gene symbol.
fChromosome:	chromosome.
fCytoBand:	cytoband.

**Author(s)**

Christian Stratowa

**See Also**

[volcanoplot](#)

**Description**

Methods for accessing perfect match (PM) and mismatch (MM) probes.

*Usage*

```
pm(object, which = "pm", unitID = NULL, unittype = "transcript")
mm(object, which = "mm", unitID = NULL, unittype = "transcript")
```

**Arguments**

object	object of class DataTreeSet.
which	type of perfect match or mismatch probes to be returned.
unitID	optional vector of UNIT_IDs.
unittype	character vector, "transcript" or "probeset".

**Details**

For expression arrays all the perfect match (pm) or mismatch (mm) probes on the arrays the object represents are returned as data.frame.

For exon arrays, pm returns the probes of the different exon levels as data.frame, i.e. which can have one of the following values:

core:	probesets supported by RefSeq and full-length GenBank transcripts.
metacore:	core meta-probesets.
extended:	probesets with other cDNA support.
metaextended:	extended meta-probesets.
full:	probesets supported by gene predictions only.
metafull:	full meta-probesets.
affx:	standard AFFX controls.

For whole genome arrays, pm returns the probes of the different exon levels as data.frame, i.e. which can have one of the following values:

core:	probesets with category 'unique' and 'mixed'.
metacore:	probesets with category 'unique' only.
affx:	standard AFFX controls.

For exon/genome arrays, mm returns the background probes as data.frame, i.e. which is either "genomic" or "antigenomic".

**Value**

A [data.frame](#).

**Author(s)**

Christian Stratowa

**See Also**

[validData](#)

**Examples**

```
## load existing ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

## need to attach scheme mask and probe intensities
data.test3 <- attachMask(data.test3)
data.test3 <- attachInten(data.test3)

pm <- pm(data.test3)
mm <- mm(data.test3)
head(pm)
head(mm)

## need to convert Affy ID to UNIT_ID first
id <- transcriptID2unitID(schemeSet(data.test3), transcriptID="100084_at", as.list=FALSE)
pm <- pm(data.test3, unitID=id)
mm <- mm(data.test3, unitID=id)
head(pm)
head(mm)

## optionally remove mask and data to free memory
data.test3 <- removeInten(data.test3)
data.test3 <- removeMask(data.test3)
```

pmplot-methods

*Barplot of PM and MM Intensities.***Description**

Creates a barplot of mean perfect match and mismatch intensities.

*Usage*

```
pmplot(x, which = "", size = 0, transfo = NULL, method = "mean", name
```

**Arguments**

x	object of class <a href="#">DataTreeSet</a> .
which	type of probes to be used, for details see <a href="#">validData</a> .
size	length of sequence to be generated as subset.
transfo	a valid function to transform the data, usually “log2”, or “0”.
method	method to compute average intensities, “mean” or “median”.
names	optional vector of sample names.
beside	logical. If FALSE, mean intensities are portrayed as stacked bars, and if TRUE the columns are portrayed as juxtaposed bars.
col	color of PM, MM bars.
legend	a vector of text used to construct a legend for the plot, or a logical indicating whether a legend should be included.
las	the style of axis labels.
ylab	a title for the y axis.
...	optional arguments to be passed to barplot.



## Details

Produces barplots of mean perfect match and mismatch intensities for slot data for an object of class `DataTreeSet`.

For `names=NULL` full column names of slot data will be displayed while for `names="namepart"` column names will be displayed without name extension. If `names` is a vector of column names, only these columns will displayed as `pmplot`.

## Note

Data must first be attached to class `DataTreeSet` using method `attachInten`.

## Author(s)

Christian Stratowa

## See Also

[plotPM](#), [boxplot](#), [barplot](#)

## Examples

```
## load existing ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

## need to attach scheme mask and probe intensities
data.test3 <- attachMask(data.test3)
data.test3 <- attachInten(data.test3)

if (interactive()) {
  pmplot(data.test3)
}

## optionally remove mask and data to free memory
data.test3 <- removeInten(data.test3)
data.test3 <- removeMask(data.test3)
```

---

prefilter

*Function for Applying a PreFilter to an ExprTreeSet*

---

## Description

This function applies a `PreFilter` to an `ExprTreeSet`.

## Usage

```
prefilter(xps.expr,
          filename = character(0),
          filedir  = getwd(),
          filter   = NULL,
          minfilters = 999,
          logbase  = "log2",
```

```

treename = "PreFilter",
xps.call = NULL,
verbose = TRUE)

```

```
xpsPreFilter(object, ...)
```

### Arguments

xps.expr	object of class ExprTreeSet.
filename	file name of ROOT filter file.
filedir	system directory where ROOT filter file should be stored.
filter	object of class PreFilter.
minfilters	minimum number of initialized filter methods to satisfy (default is all filters).
logbase	convert data to logarithm of base: "0", "log", "log2" (default), "log10"
treename	tree name to be used in ROOT filter file.
xps.call	optional object of class CallTreeSet.
verbose	logical, if TRUE print status information.
object	object of class ExprTreeSet.
...	same arguments as function prefilter.

### Details

This function applies the different filters initialized with constructor [PreFilter](#) to the [ExprTreeSet](#) `xps.expr`.

Slot `minfilters` determines the minimum number of initialized filters, which must be satisfied so that the mask is set to `flag=1`. For `minfilters=1` at least one filter must be satisfied, equivalent to logical 'OR'; for `minfilters=999` all filters must be satisfied, equivalent to logical 'AND'.

If method [callFilter](#) was initialized with constructor [PreFilter](#) then [CallTreeSet](#) `xps.call` must be supplied, usually created with function [mas5.call](#).

### Value

A [FilterTreeSet](#)

### Author(s)

Christian Stratowa

### See Also

[PreFilter](#), [unifilter](#)

### Examples

```

## Not run:
## first, load ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

## second, create an ExprTreeSet

```

```

data.rma <- rma(data.test3,"tmp_TestRMA",tmpdir="",background="pmonly",normalize=TRUE,verbose=FALSE)
## note: do not copy/paste this code, it is necessary only because R CMD check fails since it does not find tmp
data.rma@rootfile <- paste(path.package("xps"),"rootdata/tmp_Test3RMA.root",sep="/")
data.rma@filedir <- paste(path.package("xps"),"rootdata",sep="/")

## third, construct a PreFilter
prefltr <- PreFilter(mad=c(0.5,0.01),lothreshold=c(6.0,0.02,"mean"),hihreshold=c(10.5,80.0,"percent"))

## finally, create a FilterTreeSet
rma.pfr <- prefilter(data.rma,"tmp_Test3Prefilter",getwd(),prefltr,2,verbose=FALSE)
str(rma.pfr)

## End(Not run)

```

---

PreFilter-class	<i>Class PreFilter</i>
-----------------	------------------------

---

## Description

Class `PreFilter` allows to apply different filters to class `ExprTreeSet`, i.e. to the expression level data.frame data.

## Objects from the Class

Objects can be created by calls of the form `new("PreFilter", ...)`. Alternatively, the constructor `PreFilter` can be used.

## Slots

`mad`: Object of class "list" describing parameters for `madFilter`.  
`cv`: Object of class "list" describing parameters for `cvFilter`.  
`variance`: Object of class "list" describing parameters for `varFilter`.  
`difference`: Object of class "list" describing parameters for `diffFilter`.  
`ratio`: Object of class "list" describing parameters for `ratioFilter`.  
`gap`: Object of class "list" describing parameters for `gapFilter`.  
`hihreshold`: Object of class "list" describing parameters for `highFilter`.  
`lothreshold`: Object of class "list" describing parameters for `lowFilter`.  
`quantile`: Object of class "list" describing parameters for `quantileFilter`.  
`prescall`: Object of class "list" describing parameters for `callFilter`.  
`numfilters`: Object of class "numeric" giving the number of filters applied.

## Extends

Class `Filter`, directly.

**Methods**

**callFilter** signature(object = "PreFilter"): extracts slot prescall.

**callFilter<-** signature(object = "PreFilter", value = "character"): replaces slot prescall with character vector c(cutoff, samples, condition).

**cvFilter** signature(object = "PreFilter"): extracts slot cv.

**cvFilter<-** signature(object = "PreFilter", value = "numeric"): replaces slot cv with numeric vector c(cutoff, trim, epsilon).

**diffFilter** signature(object = "PreFilter"): extracts slot difference.

**diffFilter<-** signature(object = "PreFilter", value = "numeric"): replaces slot difference with numeric vector c(cutoff, trim, epsilon).

**gapFilter** signature(object = "PreFilter"): extracts slot gap.

**gapFilter<-** signature(object = "PreFilter", value = "numeric"): replaces slot gap with numeric vector c(cutoff, window, trim, epsilon).

**highFilter** signature(object = "PreFilter"): extracts slot hithreshold.

**highFilter<-** signature(object = "PreFilter", value = "character"): replaces slot hithreshold with character vector c(cutoff, parameter, condition).

**lowFilter** signature(object = "PreFilter"): extracts slot lothreshold.

**lowFilter<-** signature(object = "PreFilter", value = "character"): replaces slot lothreshold with character vector c(cutoff, parameter, condition).

**madFilter** signature(object = "PreFilter"): extracts slot mad.

**madFilter<-** signature(object = "PreFilter", value = "numeric"): replaces slot mad with numeric vector c(cutoff, epsilon).

**quantileFilter** signature(object = "PreFilter"): extracts slot quantile.

**quantileFilter<-** signature(object = "PreFilter", value = "numeric"): replaces slot quantile with numeric vector c(cutoff, loquantile, hiquantile).

**ratioFilter** signature(object = "PreFilter"): extracts slot ratio.

**ratioFilter<-** signature(object = "PreFilter", value = "numeric"): replaces slot ratio with numeric vector c(cutoff).

**varFilter** signature(object = "PreFilter"): extracts slot variance.

**varFilter<-** signature(object = "PreFilter", value = "numeric"): replaces slot variance with numeric vector c(cutoff, trim, epsilon).

**Author(s)**

Christian Stratowa

**See Also**

related classes [Filter](#), [UniFilter](#).

**Examples**

```
## for demonstration purposes only: initialize all pre-filters
prefltr <- new("PreFilter")
madFilter(prefltr) <- c(0.5,0.01)
cvFilter(prefltr) <- c(0.3,0.0,0.01)
varFilter(prefltr) <- c(0.6,0.02,0.01)
```

```
diffFilter(prefltr) <- c(2.2,0.0,0.01)
ratioFilter(prefltr) <- c(1.5)
gapFilter(prefltr) <- c(0.3,0.05,0.0,0.01)
lowFilter(prefltr) <- c(4.0,3,"samples")
highFilter(prefltr) <- c(14.5,75.0,"percent")
quantileFilter(prefltr) <- c(3.0, 0.05, 0.95)
callFilter(prefltr) <- c(0.02,80.0,"percent")
str(prefltr)
```

---

PreFilter-constructor *Constructor for Class PreFilter*

---

## Description

Constructor for class PreFilter allows to apply different filters to class [ExprTreeSet](#), i.e. to the expression level data.frame data.

## Usage

```
PreFilter(mad          = character(),
          cv           = character(),
          variance     = character(),
          difference   = character(),
          ratio        = character(),
          gap          = character(),
          lothreshold  = character(),
          hithreshold  = character(),
          quantile     = character(),
          prescall     = character())
```

## Arguments

mad	"character" vector describing parameters for <a href="#">madFilter</a> .
cv	"character" vector describing parameters for <a href="#">cvFilter</a> .
variance	"character" vector describing parameters for <a href="#">varFilter</a> .
difference	"character" vector describing parameters for <a href="#">diffFilter</a> .
ratio	"character" vector describing parameters for <a href="#">ratioFilter</a> .
gap	"character" vector describing parameters for <a href="#">gapFilter</a> .
lothreshold	"character" vector describing parameters for <a href="#">lowFilter</a> .
hithreshold	"character" vector describing parameters for <a href="#">highFilter</a> .
quantile	"character" vector describing parameters for <a href="#">quantileFilter</a> .
prescall	"character" vector describing parameters for <a href="#">callFilter</a> .

**Details**

The PreFilter constructor allows to apply the following filters to class [ExprTreeSet](#):

mad:	character vector c(cutoff,epsilon).
cv:	character vector c(cutoff,trim,epsilon).
variance:	character vector c(cutoff,trim,epsilon).
difference:	character vector c(cutoff,trim,epsilon).
ratio:	character vector c(cutoff).
gap:	character vector c(cutoff>window,trim,epsilon).
lothreshold:	character vector c(cutoff,parameter,condition).
hithreshold:	character vector c(cutoff,parameter,condition).
quantile:	character vector c(cutoff,loquantile,hiquantile).
prescall:	character vector c(cutoff,samples,condition).

**Value**

An object of type "[PreFilter](#)"

**Note**

Function [PreFilter](#) is used as constructor for class [PreFilter](#) so that the user need not know details for creating S4 classes.

**Author(s)**

Christian Stratowa

**See Also**

[Filter](#), [UniFilter](#)

**Examples**

```
## fill character vectors within constructor
prefltr <- PreFilter(mad=c(0.5,0.01), prescall=c(0.002, 6,"samples"),
                    lothreshold=c(6.0,0.02,"mean"), hithreshold=c(10.5,80.0,"percent"))
str(prefltr)

## alternatively add character vectors as methods after creation of constructor
prefltr <- PreFilter()
madFilter(prefltr) <- c(0.5,0.01)
gapFilter(prefltr) <- c(0.3,0.05,0.0,0.01)
lowFilter(prefltr) <- c(4.0,3,"samples")
highFilter(prefltr) <- c(14.5,75.0,"percent")
str(prefltr)
```

---

presCall-methods	<i>Get/Set Present Call Values</i>
------------------	------------------------------------

---

### Description

Get/set present call values from/for class `CallTreeSet`.

#### *Usage*

```
presCall(object)
presCall(object, treenames = NULL) <- value

pvalData(object)
pvalData(object, treenames = NULL) <- value
```

### Arguments

<code>object</code>	object of class <code>CallTreeSet</code> .
<code>treenames</code>	character vector containing optional tree names to be used as subset.
<code>value</code>	<code>data.frame</code> containing present call values.

### Details

Get the p-values from slot data or present calls from slot `detcall`, or set slot data or `detcall`, respectively, to value.

Method `presCall` returns the present calls from slot `detcall` as `data.frame`, while replacement method `presCall<-` allows to replace slot `detcall` with a `data.frame`.

Method `pvalData` returns the p-values from slot data as `data.frame`, while replacement method `pvalData<-` allows to replace slot data with a `data.frame`.

In order to create an `CallTreeSet` containing only a subset of e.g. slot data, first export slot data using method `pvalData`, create a character vector containing only `treenames` to be used in the subset, and then use replacement method `pvalData<-` to replace slot data with the subset. Slots `treenames` and `numtrees` will be updated automatically for `pvalData<-` but not for `presCall<-`.

Note: When creating character vector `treenames` it is sufficient to use the name part of the tree name w/o the extension.

Note: If you do not want to replace your current object, create first a copy of type `CallTreeSet` by simply writing `newobj <- oldobj`, and use `newobj` for replacement.

### Author(s)

Christian Stratowa

### See Also

[exprs](#)

**Examples**

```
## Not run:
## load ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

## create an CallTreeSet
call.mas5 <- mas5.call(data.test3, "tmp_TestMAS5Call", tmpdir="", verbose=FALSE)

## get p-values
value <- pvalData(call.mas5)

## selected treenames only
treenames <- c("TestA2", "TestB1")

## make a copy of your object if you do not want to replace it
subset.call <- call.mas5

## replace slot data with subset
exprs(subset.call, treenames) <- value
str(subset.call)

## End(Not run)
```

---

probeContentGC-methods

*Get G/C Content for Probes*


---

**Description**

Get G/C content for all or selected UNIT\_IDs.

*Usage*

```
probeContentGC(object, which = "", unitID = NULL, unittype = "transcript")
```

**Arguments**

object	Object of class "SchemeTreeSet" or "DataTreeSet".
which	type of probes to be used, for details see <a href="#">validData</a> .
unitID	optional vector of UNIT_IDs.
unittype	character vector, one of "transcript", "probeset".

**Details**

Function probeContentGC returns a data.frame containing columns "Mask" and "ContentGC" for all or selected the UNIT\_ID(s).

For exon arrays the type of UNIT\_ID(s) depends on unittype.

**Value**

A data.frame.



**Author(s)**

Christian Stratowa

**See Also**[probeSequence](#)**Examples**

```
## load ROOT scheme file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
scheme.test3 <- attachProbeContentGC(scheme.test3)

## get UNIT_ID for probeset IDs
id <- probesetID2unitID(scheme.test3, c("PA1178_oprH_at", "AFFX-Bt_eIF-4E_3_at", "100084_at"))

## get GC content
gc <- probeContentGC(scheme.test3, unitID=id)
head(gc)

scheme.test3 <- removeProbeContentGC(scheme.test3)

rm(scheme.test3)
gc()
```

---

probeSequence-methods *Get Probe Sequence*

---

**Description**

Get probe sequences for all or selected UNIT\_IDs.

*Usage*

```
probeSequence(object, unitID = NULL)
```

**Arguments**

object	Object of class "SchemeTreeSet" or "DataTreeSet".
unitID	optional vector of UNIT_IDs.

**Details**

Function probeSequence returns a data.frame containing column "ProbeSequence" for all or selected the UNIT\_ID(s).

**Value**

A data.frame.

**Author(s)**

Christian Stratowa

**See Also**[probeContentGC](#)**Examples**

```
## load ROOT scheme file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
scheme.test3 <- attachProbeSequence(scheme.test3)

## get UNIT_ID for probeset ID
id <- probesetID2unitID(scheme.test3, "100084_at")

## get GC content
seq <- probeSequence(scheme.test3, unitID=id)
head(seq)

scheme.test3 <- removeProbeSequence(scheme.test3)

rm(scheme.test3)
gc()
```

---

 probesetID2unitID-methods

*Conversion between Probeset IDs and UnitIDs*


---

**Description**

Convert probeset IDs and transcript IDs to internal UNIT\_IDs and vice verse.

*Usage*

```
probesetID2unitID(object, probesetID = NULL, as.list = TRUE)
transcriptID2unitID(object, transcriptID = NULL, as.list = TRUE)
unitID2probesetID(object, unitID = NULL, as.list = TRUE)
unitID2transcriptID(object, unitID = NULL, as.list = TRUE)
```

**Arguments**

object	Object of class "SchemeTreeSet" or "DataTreeSet".
probesetID	optional vector of probeset IDs.
transcriptID	optional vector of transcript IDs.
unitID	optional vector of UNIT_IDs.
as.list	if TRUE a list will be returned (default is data.frame).

**Details**

Functions probesetID2unitID and transcriptID2unitID return the UNIT\_ID(s) for all or selected probeset IDs or transcript IDs, respectively.

Conversely, functions unitID2probesetID and unitID2transcriptID return the probeset IDs or transcript IDs, respectively, for all or selected UNIT\_IDs. . For expression arrays the functions for probeset IDs and transcript IDs return identical IDs. . For exon arrays the functions for probeset IDs and transcript IDs return the probeset\_id(s) or transcript\_cluster\_id(s), respectively.

By default a list is returned, however for as.list=FALSE a character vector of IDs is returned.

**Value**

A list or character vector.

**Author(s)**

Christian Stratowa

**See Also**

[unitID2transcriptID](#), [unitID2probesetID](#)

**Examples**

```
## load ROOT scheme file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))

## unitNames not attached
id <- unitID2probesetID(scheme.test3, c(2,34,229))
id

## unitNames attached
scheme.test3 <- attachUnitNames(scheme.test3)
id <- probesetID2unitID(scheme.test3, c("PA1178_oprH_at", "AFFX-Bt_eIF-4E_3_at", "100084_at"))
id
scheme.test3 <- removeUnitNames(scheme.test3)

rm(scheme.test3)
gc()
```

---

probesetplot-methods *Plot of Probe Intensities for a Probeset.*

---

**Description**

Creates a line plot of probe intensities for a probeset.

*Usage*

```
probesetplot(x, unitID, unittype = "transcript", which = "pm",
```

**Arguments**

x	object of class <a href="#">DataTreeSet</a> .
unitID	unit ID of probeset with type of ID determined by parameter unittype.
unittype	character vector, one of "unit", "transcript", "probeset".
which	type of probes to be used, for details see <a href="#">validData</a> .
transfo	a valid function to transform the data, usually "log2", or "0".
names	optional vector of sample names.
ylim	range for the plotted y values.
col	color to use for the different samples.
lty	line types to use for the different samples.
add.legend	logical, if TRUE a legend of sample names will be drawn. Optionally, a character indicating the position of the legend, default is "topleft".
...	optional arguments to be passed to probesetplot.

**Details**

Produces line plots of the probe intensities for probeset unitID. Probe intensities are taken from slot data.

For names=NULL full column names of slot data will be displayed while for names="namepart" column names will be displayed without name extension. If names is a vector of column names, line plots of probe intensities will only be drawn for these columns.

**Note**

Data must first be attached to class [DataTreeSet](#) using method [attachInten](#). Furthermore, unit names must be attached using method [attachUnitNames](#).

**Author(s)**

Christian Stratowa

**See Also**

[plotPM](#), [boxplot](#), [barplot](#)

**Examples**

```
## load existing ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

## need to attach probe intensities and optionally unit names
data.test3 <- attachUnitNames(data.test3)
data.test3 <- attachInten(data.test3)

if (interactive()) {
  probesetplot(data.test3, unitID="100084_at", unittype="transcript", add.legend=TRUE)
}

## optionally remove unit names and data to free memory
data.test3 <- removeInten(data.test3)
data.test3 <- removeUnitNames(data.test3)
```

---

ProcesSet-class

*Class ProcesSet*

---

**Description**

This class provides access to class [SchemeTreeSet](#) for the derived classes [DataTreeSet](#), [ExprTreeSet](#) and [CallTreeSet](#). It extends class [TreeSet](#).

**Objects from the Class**

Usually, no objects are created from it.

**Slots**

**scheme:** Object of class "SchemeTreeSet" providing access to [ROOT](#) scheme file.

**data:** Object of class "data.frame". The data.frame can contain the data stored in [ROOT](#) data trees.

**params:** Object of class "list" representing relevant parameters.

**setname:** Object of class "character" representing the name to the [ROOT](#) file subdirectory where the [ROOT](#) trees are stored, usually one of 'DataTreeSet', 'PreprocesSet', 'CallTreeSet'.

**settype:** Object of class "character" describing the type of treeset stored in setname, usually one of 'rawdata', 'preprocess'.

**rootfile:** Object of class "character" representing the name of the [ROOT](#) file, including full path.

**filedir:** Object of class "character" describing the full path to the system directory where rootfile is stored.

**numtrees:** Object of class "numeric" representing the number of [ROOT](#) trees stored in subdirectory setname.

**treenames:** Object of class "list" representing the names of the [ROOT](#) trees stored in subdirectory setname.

**Extends**

Class "[TreeSet](#)", directly.

**Methods**

**attachData** signature(object = "ProcesSet"): exports data from [ROOT](#) data file and saves as data.frame data.

**boxplot** signature(x = "ProcesSet"): creates a [boxplot](#) of the data from data.frame data.

**chipName** signature(object = "ProcesSet"): extracts slot chipname from slot scheme.

**chipType** signature(object = "ProcesSet"): extracts slot chiptype from slot scheme.

**export** signature(object = "ProcesSet"): exports [ROOT](#) trees as text file, see [export-methods](#).

**getTreeData** signature(object = "ProcesSet"): exports tree data from [ROOT](#) file rootfile, and saves as data.frame data.

**hist** signature(x = "ProcesSet"): creates a plot showing the histograms for data.frame data.

**image** signature(x = "ProcesSet"): creates an image for each column from data.frame data or bgrd, respectively.

**mboxplot** signature(x = "ProcesSet"): creates an M-boxplot of the data from data.frame data.

**removeData** signature(object = "ProcesSet"): replaces data.frame data with an empty data.frame of dim(0,0).

**schemeFile** signature(object = "ProcesSet"): extracts the [ROOT](#) scheme file from slot scheme.

**schemeFile<-** signature(object = "ProcesSet"), value = "character"): replaces the [ROOT](#) scheme file from slot scheme.

**schemeSet** signature(object = "ProcesSet"): extracts slot scheme.

**schemeSet<-** signature(object = "ProcesSet"), value = "SchemeTreeSet"): replaces slot scheme with a different SchemeTreeSet.

**treeData** signature(object = "ProcesSet"): extracts all columns from data.frame data.

**validData** signature(object = "ProcesSet"): extracts a subset of columns from data.frame data.

**Author(s)**

Christian Stratowa

**See Also**

derived classes [DataTreeSet](#), [ExprTreeSet](#), [CallTreeSet](#), [QualTreeSet](#).

**Examples**

```
showClass("ProcesSet")
```

---

ProjectInfo-class	<i>Class ProjectInfo</i>
-------------------	--------------------------

---

**Description**

This class allows to save the relevant project information in the [ROOT](#) data file and in class [DataTreeSet](#).

**Objects from the Class**

Objects can be created by calls of the form  
`new("ProjectInfo", submitter=[character], laboratory=[character], contact=[character], ...)`.  
 Alternatively, the constructor [ProjectInfo](#) can be used.

**Slots**

**submitter:** Object of class "character" representing the name of the submitter.  
**laboratory:** Object of class "character" representing the laboratory of the submitter.  
**contact:** Object of class "character" representing the contact address of the submitter.  
**project:** Object of class "list" representing the project information.  
**author:** Object of class "list" representing the author information.  
**dataset:** Object of class "list" representing the dataset information.  
**source:** Object of class "list" representing the sample source information.  
**sample:** Object of class "list" representing the sample information.  
**celline:** Object of class "list" representing the sample information for cell lines.  
**primarycell:** Object of class "list" representing the sample information for primary cells.  
**tissue:** Object of class "list" representing the sample information for tissues.  
**biopsy:** Object of class "list" representing the sample information for biopsies.  
**arraytype:** Object of class "list" representing the array information.  
**hybridizations:** Object of class "data.frame" representing the hybridization information for each hybridization.  
**treatments:** Object of class "data.frame" representing the treatment information for each hybridization.

**Methods**

**projectInfo** signature(object = "ProjectInfo"): extracts slot project.

**projectInfo<-** signature(object = "ProjectInfo", value = "character"): replaces slot project with character vector c(name,date,type,description,comments).

**authorInfo** signature(object = "ProjectInfo"): extracts slot author.

**authorInfo<-** signature(object = "ProjectInfo", value = "character"): replaces slot author with character vector c(lastname,firstname,type,company,department,email, phone,comments).

**datasetInfo** signature(object = "ProjectInfo"): extracts slot dataset.

**datasetInfo<-** signature(object = "ProjectInfo", value = "character"): replaces slot dataset with character vector c(name,type,sample,submitter,date,description,comments).

**sourceInfo** signature(object = "ProjectInfo"): extracts slot source.

**sourceInfo<-** signature(object = "ProjectInfo", value = "character"): replaces slot source with character vector c(name,type,species,subspecies,description,comments).

**sampleInfo** signature(object = "ProjectInfo"): extracts slot sample.

**sampleInfo<-** signature(object = "ProjectInfo", value = "character"): replaces slot sample with character vector c(name,type,sex,phenotype,genotype,extraction, isxenograft,xenostain,xenosex,xenoage).

**celllineInfo** signature(object = "ProjectInfo"): extracts slot cellline.

**celllineInfo<-** signature(object = "ProjectInfo", value = "character"): replaces slot cellline with character vector c(name,type,parent,atcc,modification,sex,phenotype, genotype,extraction,isxenograft,xenostain,xenosex,xenoage).

**primcellInfo** signature(object = "ProjectInfo"): extracts slot primarycell.

**primcellInfo<-** signature(object = "ProjectInfo", value = "character"): replaces slot primarycell with character vector c(name,type,date,description,sex,phenotype, genotype,extraction,isxenograft,xenostain,xenosex,xenoage,xenoageunit,comments).

**tissueInfo** signature(object = "ProjectInfo"): extracts slot tissue.

**tissueInfo<-** signature(object = "ProjectInfo", value = "character"): replaces slot tissue with character vector c(name,type,development,morphology,disease,stage, donorage,ageunit,status,sex,phenotype,xenoage,xenoageunit,comments).

**biopsyInfo** signature(object = "ProjectInfo"): extracts slot biopsy.

**biopsyInfo<-** signature(object = "ProjectInfo", value = "character"): replaces slot biopsy with character vector c(name,type,morphology,disease,stage,donorage,ageunit, status,sex,phenotype,genotype,extraction,isxenograft,xenostain,xenosex,xenoage,xenoageunit,comments).

**arrayInfo** signature(object = "ProjectInfo"): extracts slot arraytype.

**arrayInfo<-** signature(object = "ProjectInfo", value = "character"): replaces slot arraytype with character vector c(chipname,chiptype,description,comments).

**hybridizInfo** signature(object = "ProjectInfo"): extracts slot hybridizations.

**hybridizInfo<-** signature(object = "ProjectInfo", value = "character"): replaces slot hybridizations with vector of character vectors with each containing c(name,type,inputname,date,preparation,proportion).

**treatmentInfo** signature(object = "ProjectInfo"): extracts slot treatments.

**treatmentInfo<-** signature(object = "ProjectInfo", value = "character"): replaces slot treatments with vector of character vectors with each containing c(name,type,concentration,concentrationunit,timepoint).

**show** signature(object = "ProjectInfo"): shows the content of ProjectInfo.

**Author(s)**

Christian Stratowa

**Examples**

```

project <- new("ProjectInfo",submitter="Christian", laboratory="home",contact="email")
projectInfo(project) <- c("TestProject", "20060106", "Project Type", "use Test3 data for testing", "my comment")
authorInfo(project) <- c("Stratowa", "Christian", "Project Leader", "Company", "Dept", "cstrato.at.aon.at", "+")
datasetInfo(project) <- c("Test3Set", "MC", "Tissue", "Stratowa", "20060106", "description", "my comment")
sourceInfo(project) <- c("Unknown", "source type", "Homo sapiens", "caucasian", "description", "my comment")
primcellInfo(project) <- c("Mel131", "primary cell", "20071123", "extracted from patient", "male", "my pheno", "my")
arrayInfo(project) <- c("Test3", "GeneChip", "description", "my comment")
hybridizInfo(project) <- c(c("TestA1", "hyb type", "TestA1.CEL", "20071117", "my prep1", "standard protocol", "A1", "1", "mM", "1.0", "hours", "intravenous", "my comment"),
c("TestA2", "hyb type", "TestA2.CEL", "20071117", "my prep2", "standard protocol", "A2", "1", "mM", "1.0", "hours", "intravenous", "my comment"),
c("TestB1", "hyb type", "TestB1.CEL", "20071117", "my prep1", "standard protocol", "B1", "2", "mM", "1.0", "hours", "intravenous", "my comment"),
c("TestB2", "hyb type", "TestB2.CEL", "20071117", "my prep2", "standard protocol", "B2", "2", "mM", "1.0", "hours", "intravenous", "my comment"))
treatmentInfo(project) <- c(c("TestA1", "DMSO", "4.3", "mM", "1.0", "hours", "intravenous", "my comment"),
c("TestA2", "DMSO", "4.3", "mM", "8.0", "hours", "intravenous", "my comment"),
c("TestB1", "DrugA2", "4.3", "mM", "1.0", "hours", "intravenous", "my comment"),
c("TestB2", "DrugA2", "4.3", "mM", "8.0", "hours", "intravenous", "my comment"))

show(project)

```

---

ProjectInfo-constructor

*Constructor for Class ProjectInfo*


---

**Description**

Constructor for class ProjectInfo class allows to save the relevant project information in the [ROOT](#) data file and in class [DataTreeSet](#).

**Usage**

```

ProjectInfo(submitter      = character(),
            laboratory     = character(),
            contact        = character(),
            project        = character(),
            author         = character(),
            dataset        = character(),
            source         = character(),
            sample         = character(),
            celline        = character(),
            primarycell    = character(),
            tissue         = character(),
            biopsy         = character(),
            arraytype      = character(),
            hybridizations = character(),
            treatments     = character())

```

**Arguments**

submitter	"character" representing the name of the submitter.
laboratory	"character" representing the laboratory of the submitter.
contact	"character" representing the contact address of the submitter.
project	"character" vector representing the project information.



author	"character" vector representing the author information.
dataset	"character" vector representing the dataset information.
source	"character" vector representing the sample source information.
sample	"character" vector representing the sample information.
celline	"character" vector representing the sample information for cell lines.
primarycell	"character" vector representing the sample information for primary cells.
tissue	"character" vector representing the sample information for tissues.
biopsy	"character" vector representing the sample information for biopsies.
arraytype	"character" vector representing the array information.
hybridizations	"character" vector representing the hybridization information for each hybridization.
treatments	"character" vector representing the treatment information for each hybridization.

### Details

The ProjectInfo constructor allows to save the following project information in the [ROOT](#) data file and in class [DataTreeSet](#):

submitter:	name of the submitter.
laboratory:	laboratory of the submitter.
contact:	contact address of the submitter.
project:	character vector c(name,date,type,description,comments).
author:	character vector c(lastname,firstname,type,company,department,email, phone,comments)..
dataset:	character vector c(name,type,sample,submitter,date,description,comments).
source:	character vector c(name,type,species,subspecies,description,comments).
sample:	character vector c(name,type,sex,phenotype,genotype,extraction, isxenograft,xenostain,xenosex,xenostrain).
celline:	character vector c(name,type,parent,atcc,modification,sex,phenotype, genotype,extraction,isxenograft,xenostain).
primarycell:	character vector c(name,type,date,description,sex,phenotype, genotype,extraction,isxenograft,xenostain).
tissue:	character vector c(name,type,development,morphology,disease,stage, donorage,ageunit,status,sex,phenotype,genotype).
biopsy:	character vector c(name,type,morphology,disease,stage,donorage,ageunit, status,sex,phenotype,genotype).
arraytype:	character vector c(chipname,chiptype,description,comments).
hybridizations:	vector of character vectors with each containing c(name,type,inputname,date,preparation,protocol,comments).
treatments:	vector of character vectors with each containing c(name,type,concentration,concentrationunit,time,comments).

### Value

An object of type "[ProjectInfo](#)"

### Note

Function [ProjectInfo](#) is used as constructor for class [ProjectInfo](#) so that the user need not know details for creating S4 classes.

### Author(s)

Christian Stratowa

### See Also

[ProjectInfo](#)

**Examples**

```
## fill character vectors within constructor
project <- ProjectInfo(submitter="Christian", laboratory="home",contact="email",
  project=c("TestProject","20060106","Project Type","use Test3 data for testing","my commen
  hybridizations=c(c("TestA1","hyb type","TestA1.CEL",20071117,"my prep1","standard protoc
  c("TestA2","hyb type","TestA2.CEL",20071117,"my prep2","standard protocol",
  c("TestB1","hyb type","TestB1.CEL",20071117,"my prep1","standard protocol",
  c("TestB2","hyb type","TestB2.CEL",20071117,"my prep2","standard protocol",

str(project)

## alternatively add character vectors as methods after creation of constructor
authorInfo(project) <- c("Stratowa","Christian","Project Leader","Company","Dept","cstrato.at.aon.at","+
datasetInfo(project) <- c("Test3Set","MC","Tissue","Stratowa","20060106","description","my comment")
treatmentInfo(project) <- c(c("TestA1","DMSO",4.3,"mM",1.0,"hours","intravenous","my comment"),
  c("TestA2","DMSO",4.3,"mM",8.0,"hours","intravenous","my comment"),
  c("TestB1","DrugA2",4.3,"mM",1.0,"hours","intravenous","my comment"),
  c("TestB2","DrugA2",4.3,"mM",8.0,"hours","intravenous","my comment"))

str(project)
```

qualify

*Probe Set Quality Control Functions***Description**

Converts Affymetrix probe level data to expression levels by fitting a multichip model.

**Usage**

```
qualify(xps.data,
  filename = character(),
  filedir  = getwd(),
  tmpdir   = "",
  update   = FALSE,
  select   = "none",
  method   = character(),
  option    = "transcript",
  logbase  = "log2",
  exonlevel = "",
  params    = list(),
  xps.scheme = NULL,
  add.data  = TRUE,
  verbose   = TRUE)

qualify.rlm(xps.data,
  filename = character(),
  filedir  = getwd(),
  tmpdir   = "",
  update   = FALSE,
  option    = "transcript",
  exonlevel = "",
  xps.scheme = NULL,
  add.data  = TRUE,
```

```
verbose = TRUE)
```

```
xpsQualify(object, ...)
```

### Arguments

xps.data	object of class <code>DataTreeSet</code> .
filename	file name of ROOT data file.
filedir	system directory where ROOT data file should be stored.
tmpdir	optional temporary directory where temporary ROOT files should be stored.
update	logical. If TRUE the existing ROOT data file filename will be updated.
select	type of probes to select for summarization.
method	qualification method to use, currently <code>rlm</code> .
option	option determining the grouping of probes for summarization, one of 'transcript', 'exon', 'probeset'; exon/genome arrays only.
logbase	logarithm base as character, one of '0', 'log', 'log2', 'log10'.
exonlevel	exon annotation level determining which probes should be used for summarization; exon/genome arrays only.
params	vector of parameters for summarization method.
xps.scheme	optional alternative <code>SchemeTreeSet</code> .
add.data	logical. If TRUE expression data will be included as slot data.
verbose	logical, if TRUE print status information.
object	object of class <code>DataTreeSet</code> .
...	the arguments described above.

### Details

Converts Affymetrix probe level data to expression levels by fitting a multichip model.

This function stores three types of ROOT trees in filename:

- quality trees containing expression levels, normalized unscaled standard errors (NUSE), relative log expressions (RLE)
- residual trees containing the residual SE and the model fit weights
- border trees containing the border intensities, mean border intensities and center of intensities (COI)

`xpsQualify` is the `DataTreeSet` method called by function `qualify`, containing the same parameters.

### Value

An `QualTreeSet`.

### Note

This function takes any `DataTreeSet` and computes expression levels by summarizing the probe set values into one expression measure. It does NOT do any further preprocessing such as background correction or (quantile) normalization. If you want to do background correction and/or normalization first then you need to use function `fitQC`.

**Author(s)**

Christian Stratowa

**See Also**[fitQC](#)**Examples**

```
## Not run:
## first, load ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

## compute RMA stepwise

## background correction
data.bg.rma <- bgcorrect.rma(data.test3, "tmp_Test3RMABgrd", filedir=getwd())

## normalize quantiles
data.qu.rma <- normalize.quantiles(data.bg.rma, "tmp_Test3RMANorm", filedir=getwd())

## summarize medianpolish
data.mp.rma <- summarize.rma(data.qu.rma, "tmp_Test3RMAExpr", filedir=getwd(), tmpdir="")

## qualification - rlm

## fit model on raw data
data.raw.rlm <- qualify.rlm(data.test3, "tmp_Test3RawQual", filedir=getwd(), tmpdir="", option="transcript",

## fit model on background adjusted data
data.adj.rlm <- qualify.rlm(data.bg.rma, "tmp_Test3AdjQual", filedir=getwd(), tmpdir="", option="transcript",

## fit model on normalized data
data.nrm.rlm <- qualify.rlm(data.qu.rma, "tmp_Test3NormQual", filedir=getwd(), tmpdir="", option="transcript",

## get expression levels
expr.raw.rlm <- validData(data.raw.rlm)
expr.adj.rlm <- validData(data.adj.rlm)
expr.nrm.rlm <- validData(data.nrm.rlm)

## get borders
brd.raw <- borders(data.raw.rlm)
brd.adj <- borders(data.adj.rlm)

## get residuals
res.raw <- residuals(data.raw.rlm)
res.adj <- residuals(data.adj.rlm)

## get weights
w.raw <- weights(data.raw.rlm)
w.adj <- weights(data.adj.rlm)

## End(Not run)
```

---

QualTreeSet-class	<i>Class QualTreeSet</i>
-------------------	--------------------------

---

### Description

This class provides the link to the [ROOT](#) quality control file and the [ROOT](#) trees contained therein. It extends class [ProcesSet](#).

### Objects from the Class

Objects are created using functions [qualify](#), [fitQC](#), or the specialized functions [qualify.rlm](#), [fitRLM](#) or [rmaPLM](#).

### Slots

**qualopt:** Object of class "character" representing the quality control option, i.e. 'raw', 'adjusted', 'normalized' or 'all'.

**qualtype:** Object of class "character" representing the quality control type, i.e. 'rlm'.

**scheme:** Object of class "SchemeTreeSet" providing access to [ROOT](#) scheme file.

**data:** Object of class "data.frame". The data.frame can contain the data (e.g. expression levels) stored in [ROOT](#) data trees.

**params:** Object of class "list" representing relevant parameters.

**setname:** Object of class "character" representing the name to the [ROOT](#) file subdirectory where the [ROOT](#) data trees are stored, usually 'PreprocesSet'.

**settype:** Object of class "character" describing the type of treeset stored in setname, usually 'preprocess'.

**rootfile:** Object of class "character" representing the name of the [ROOT](#) data file, including full path.

**filedir:** Object of class "character" describing the full path to the system directory where rootfile is stored.

**numtrees:** Object of class "numeric" representing the number of [ROOT](#) trees stored in subdirectory setname.

**treenames:** Object of class "list" representing the names of the [ROOT](#) trees stored in subdirectory setname.

### Extends

Class "[ProcesSet](#)", directly. Class "[TreeSet](#)", by class "[ProcesSet](#)", distance 2.

### Methods

**borderplot** signature(x = "QualTreeSet"): creates a boxplot of positive and negative border elements.

**borders** signature(object = "QualTreeSet"): exports border trees from [ROOT](#) quality control file as data.frame data.

**coiplot** signature(x = "QualTreeSet"): creates a Center-of-Intensity-plot for positive and negative feature intensities.

- image** signature(x = "QualTreeSet"): creates a pseudo image for each quality control tree, i.e. residual images.
- NUSE** signature(x = "QualTreeSet"): plot Normalized Unscaled Standard Errors, or return stats, values.
- nuseplot** signature(x = "QualTreeSet"): creates a NUSE-plot.
- qualOption** signature(object = "QualTreeSet"): extracts slot qualopt.
- qualOption<-** signature(object = "QualTreeSet", value = "character"): replaces slot qualopt.
- qualType** signature(object = "QualTreeSet"): extracts slot qualtype.
- qualType<-** signature(object = "QualTreeSet", value = "character"): replaces slot qualtype.
- residuals** signature(object = "QualTreeSet"): exports residuals from the residuals trees of the **ROOT** quality control file as data.frame data.
- RLE** signature(x = "QualTreeSet"): plot Relative Log Expression, or return stats, values.
- rleplot** signature(x = "QualTreeSet"): creates a RLE-plot.
- weights** signature(object = "QualTreeSet"): exports weights from the residuals trees of the **ROOT** quality control file as data.frame data.
- xpsRNAdeg** signature(x = "QualTreeSet"): list with parameters for RNA degradation.

**Author(s)**

Christian Stratowa

**See Also**

related classes [DataTreeSet](#), [CallTreeSet](#), [ExprTreeSet](#).

**Examples**

```
showClass("QualTreeSet")
```

---

quantileFilter-methods

*Quantile Filter*

---

**Description**

This method initializes the Quantile Filter.

The Quantile Filter flags all rows with:  $\text{flag} = (\text{quantile}[\text{high}]/\text{quantile}[\text{low}] \geq \text{cutoff})$

*Usage*

```
quantileFilter(object)
quantileFilter(object, value)<-
```

**Arguments**

**object**                    object of class PreFilter.

**value**                    numeric vector c(cutoff, loquantile, hiquantile).

**Details**

The method `quantileFilter` initializes the following parameters:

cutoff: the cutoff level for the filter.  
loquantile: value for low quantile (default is loquantile=0.05).  
hiquantile: value for high quantile (default is hiquantile=0.95).

**Value**

An initialized `PreFilter` object.

**Author(s)**

Christian Stratowa

**Examples**

```
prefltr <- PreFilter()
quantileFilter(prefltr) <- c(3.0, 0.05, 0.95)
str(prefltr)
```

---

ratioFilter-methods    *Ratio Filter*

---

**Description**

This method initializes the Ratio Filter. The ratio is the maximum value divided by minimum value for each row of the expression dataframe.

The Ratio Filter flags all rows with: `flag = (max/min >= cutoff)`

*Usage*

```
ratioFilter(object)
ratioFilter(object, value)<-
```

**Arguments**

object            object of class `PreFilter`.  
value            numeric value `c(cutoff)`.

**Details**

The method `ratioFilter` initializes the following parameters:

cutoff: the cutoff level for the filter.

**Value**

An initialized `PreFilter` object.

**Author(s)**

Christian Stratowa



**Examples**

```
prefltr <- PreFilter()
ratioFilter(prefltr) <- c(1.5)
str(prefltr)
```

---

rawCELName-methods      *Method for getting names of the raw CEL-files*

---

**Description**

Method for getting names (and full path) of the original CEL-files.

*Usage*

```
rawCELName(object, treename = "*", fullpath = TRUE)
```

**Arguments**

object	object of class DataTreeSet.
treename	treename, for which the name of the original CEL-file should be returned.
fullpath	logical, if TRUE return full path.

**Details**

Since CEL-files can be imported with `import.data` using alternative `celnames`, method `rawCELName` allows to return the original name and optionally the full path for each CEL-file.

**Value**

A character vector.

**Author(s)**

Christian Stratowa

**See Also**

[import.data](#)

**Examples**

```
## load existing ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

rawCELName(data.test3)
rawCELName(data.test3, treename = "TestA2.cel", fullpath = FALSE)
```

RLE-methods

*Relative Log Expression (RLE)***Description**

Produce boxplots of Relative Log Expression (RLE) values for the set of arrays. Alternatively, summary statistics or RLE values can be extracted as `data.frame`.

*Usage*

```
RLE(x, treename = "*", type = c("plot", "stats", "values"), qualopt = NULL, ...)
```

**Arguments**

<code>x</code>	object of class <code>QualTreeSet</code> .
<code>treename</code>	vector of tree names to export.
<code>type</code>	type of output, plot, stats or values.
<code>qualopt</code>	quality control option, i.e. 'raw', 'adjusted', 'normalized' or 'all'.
<code>...</code>	optional arguments to be passed to <code>rleplot</code> .

**Details**

Create boxplots of Relative Log Expression (RLE) values for the set of arrays, i.e. of `M` plots, where `M` is determined relative to a pseudo-median reference chip.

Alternatively it is possible to extract either the summary statistics as `data.frame` (`type="stats"`) or all RLE values as `data.frame` (`type="values"`).

If an object of class `QualTreeSet` was created by fitting a probe level model with `qualopt="all"` then RLE will plot or extract RLE for "all" quality options. If you want to plot or extract RLE for a certain quality option only, e.g. "normalized" data only, then you can use parameter `qualopt` with `qualopt="<qualopt>"`.

**Author(s)**

Christian Stratowa

**See Also**

`plotRLE`, `rleplot`

**Examples**

```
## Not run:
## load existing ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

## qualification - rlm
rlm.all <- rmaPLM(data.test3, "tmp_Test3RLMall", filedir=getwd(), tmpdir="", qualopt="all", option="transcrip

## plot expression levels
if (interactive()) {
  RLE(rlm.all)
```

```

RLE(rlm.all, qualopt="normalized")
qcRLE <- RLE(rlm.all, type="stats")
qcRLE <- RLE(rlm.all, type="values")
qcRLE <- RLE(rlm.all, treename="TestA1_normalized.rlm", type="stats")
qcRLE <- RLE(rlm.all, treename="TestA1_normalized.rlm", type="values")
}

## End(Not run)

```

rleplot-methods

*Box Plots of Relative Log Expression (RLE)***Description**

Produce boxplots of Relative Log Expression (RLE) values for the set of arrays.

*Usage*

```
rleplot(x, which = "UnitName", size = 0, range = 0, names = "name")
```

**Arguments**

x	object of class <a href="#">ExprTreeSet</a> or <a href="#">QualTreeSet</a> .
which	type of probes to be used, for details see <a href="#">validData</a> .
size	length of sequence to be generated as subset.
range	determines how far the plot whiskers extend out from the box.
names	optional vector of sample names.
main	the main title for the plot.
ylim	range for the plotted y values.
las	the style of axis labels.
add.line	logical, if TRUE a horizontal line is drawn.
outline	if outline is not true, the outliers are not drawn.
...	optional arguments to be passed to <code>boxplot</code> .

**Details**

Create boxplots of Relative Log Expression (RLE) values for the set of arrays, i.e. of M plots, where M is determined relative to a pseudo-median reference chip.

For names=NULL full column names of slot data will be displayed while for names="namepart" column names will be displayed without name extension. If names is a vector of column names, only these columns will displayed as boxplot.

If an object of class [QualTreeSet](#) was created by fitting a probe level model with qualopt="all" then rleplot will plot RLE for "all" quality options. If you want to plot RLE for a certain quality option only, e.g. "normalized" data only, then you can use parameter names with names="namepart:<qualopt>", e.g. names="namepart:normalized".

**Author(s)**

Christian Stratowa

**See Also**

[RLE](#), [plotRLE](#), [mboxplot](#), [nuseplot](#)

**Examples**

```
# load existing ROOT scheme file and ROOT expression file for rma
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.rma <- root.expr(scheme.test3, paste(path.package("xps"), "rootdata/tmp_Test3RMA.root", sep="/"), "mdp")

if (interactive()) {
  rleplot(data.rma)
}
```

---

rma

*Robust Multi-Array Average Expression Measure*


---

**Description**

This function converts a [DataTreeSet](#) into an [ExprTreeSet](#) using the robust multi-array average (RMA) expression measure.

**Usage**

```
rma(xps.data,
    filename = character(0),
    filedir  = getwd(),
    tmpdir   = "",
    background = "pmonly",
    normalize = TRUE,
    option    = "transcript",
    exonlevel = "",
    params    = list(16384, 0.0, 1.0, 10, 0.01, 1),
    xps.scheme = NULL,
    add.data   = TRUE,
    verbose    = TRUE)
```

```
xpsRMA(object, ...)
```

**Arguments**

xps.data	object of class <a href="#">DataTreeSet</a> .
filename	file name of ROOT data file.
filedir	system directory where ROOT data file should be stored.
tmpdir	optional temporary directory where temporary ROOT files should be stored.
background	probes used to compute background, one of 'pmonly', 'mmonly', 'both'; for genome/exon arrays one of 'genomic', 'antigenomic'
normalize	logical. If TRUE normalize data using quantile normalization.
option	option determining the grouping of probes for summarization, one of 'transcript', 'exon', 'probeset'; exon arrays only.

exonlevel	exon annotation level determining which probes should be used for summarization; exon/genome arrays only.
params	list of (default) parameters for rma.
xps.scheme	optional alternative SchemeTreeSet.
add.data	logical. If TRUE expression data will be included as slot data.
verbose	logical, if TRUE print status information.
object	object of class DataTreeSet.
...	the arguments described above.

## Details

This function computes the RMA (Robust Multichip Average) expression measure described in Irizarry et al. for both expression arrays and exon arrays. For exon arrays it is necessary to supply the requested option and exonlevel.

Following options are valid for exon arrays:

transcript:	expression levels are computed for transcript clusters, i.e. probe sets containing the same 'transcript_cluster'.
exon:	expression levels are computed for exon clusters, i.e. probe sets containing the same 'exon_id', where each
probeset:	expression levels are computed for individual probe sets, i.e. for each 'probeset_id'.

Following exonlevel annotations are valid for exon arrays:

core:	probesets supported by RefSeq and full-length GenBank transcripts.
metacore:	core meta-probesets.
extended:	probesets with other cDNA support.
metaextended:	extended meta-probesets.
full:	probesets supported by gene predictions only.
metafull:	full meta-probesets.
ambiguous:	ambiguous probesets only.
affx:	standard AFFX controls.
all:	combination of above (including affx).

Following exonlevel annotations are valid for whole genome arrays:

core:	probesets with category 'unique', 'similar' and 'mixed'.
metacore:	probesets with category 'unique' only.
affx:	standard AFFX controls.
all:	combination of above (including affx).

Exon levels can also be combined, with following combinations being most useful:

exonlevel="metacore+affx":	core meta-probesets plus AFFX controls
exonlevel="core+extended":	probesets with cDNA support
exonlevel="core+extended+full":	supported plus predicted probesets

Exon level annotations are described in the Affymetrix whitepaper [exon\\_probeset\\_trans\\_clust\\_whitepaper.pdf](#): "Exon Probeset Annotations and Transcript Cluster Groupings".

In order to use an alternative [SchemeTreeSet](#) set the corresponding `SchemeSet` `xps.scheme`. `xpsRMA` is the `DataTreeSet` method called by function `rma`, containing the same parameters.

**Value**

An [ExprTreeSet](#)

**Note**

In contrary to other implementations of RMA the expression measure is given to you in linear scale, analogously to the expression measures computed with [mas5](#) and [mas4](#).

Please note that the default settings of `params` gives results which are identical to the results obtained with APT (Affymetrix Power Tools) and with package `affy_1.14.2` or earlier. If you want to obtain results which are identical to the results obtained with `affy_1.16.0` or later then you need to set `params = list(16384, 0.0, 0.4, 10, 0.01, 1)`.

By setting parameter `background="none"` it is possible to skip background correction .

For the analysis of many exon arrays it may be better to define a `tmpdir`, since this will store only the results in the main file and not e.g. background and normalized intensities, and thus will reduce the file size of the main file. For quantile normalization memory should not be an issue, however medianpolish depends on RAM unless you are using a temporary file.

Parameter `exonlevel` determines not only which probes are used for medianpolish, but also the probes used for background calculation and for quantile normalization. If you want to use separate probes for background calculation, quantile normalization and medianpolish summarization, you can pass a numeric vector containing three integer values corresponding to the respective `exonlevel`, e.g. you can use `exonlevel=c(16316, 8252, 8252)`, see function [exonLevel](#) for more details.

**Author(s)**

Christian Stratowa

**References**

Rafael. A. Irizarry, Benjamin M. Bolstad, Francois Collin, Leslie M. Cope, Bridget Hobbs and Terence P. Speed (2003), Summaries of Affymetrix GeneChip probe level data *Nucleic Acids Research* 31(4):e15

Bolstad, B.M., Irizarry R. A., Astrand M., and Speed, T.P. (2003), A Comparison of Normalization Methods for High Density Oligonucleotide Array Data Based on Bias and Variance. *Bioinformatics* 19(2):185-193

Irizarry, RA, Hobbs, B, Collin, F, Beazer-Barclay, YD, Antonellis, KJ, Scherf, U, Speed, TP (2003) Exploration, Normalization, and Summaries of High Density Oligonucleotide Array Probe Level Data. *Biostatistics* .Vol. 4, Number 2: 249-264

**See Also**

[express](#)

**Examples**

```

## first, load ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

data.rma <- rma(data.test3, "tmp_Test3RMA", tmpdir="", background="pmonly", normalize=TRUE, verbose=FALSE)

## get data.frame
expr.rma <- validData(data.rma)
head(expr.rma)

## plot results
if (interactive()) {
  boxplot(data.rma)
  boxplot(log2(expr.rma))
}

rm(scheme.test3, data.test3)
gc()

## Not run:
## examples using Affymetrix human tissue dataset (see also xps/examples/script4exon.R)
## first, load ROOT scheme file and ROOT data file from e.g.:
scmdir <- "/Volumes/GigaDrive/CRAN/Workspaces/Schemes"
datdir <- "/Volumes/GigaDrive/CRAN/Workspaces/ROOTData"

## 1. example - expression array, e.g. HG-U133_Plus_2:
scheme.u133p2 <- root.scheme(paste(scmdir, "Scheme_HGU133p2_na25.root", sep="/"))
data.u133p2 <- root.data(scheme.u133p2, paste(datdir, "HuTissuesU133P2_cel.root", sep="/"))

workdir <- "/Volumes/GigaDrive/CRAN/Workspaces/Exon/hutissues/u133p2"
data.rma <- rma(data.u133p2, "MixU133P2RMA", filedir=workdir, tmpdir="",
  background="pmonly", normalize=TRUE)

## 2. example - whole genome array, e.g. HuGene-1_0-st-v1:
scheme.genome <- root.scheme(paste(scmdir, "Scheme_HuGene10stv1r3_na25.root", sep="/"))
data.genome <- root.data(scheme.genome, paste(datdir, "HuTissuesGenome_cel.root", sep="/"))

workdir <- "/Volumes/GigaDrive/CRAN/Workspaces/Exon/hutissues/hugene"
data.g.rma <- rma(data.genome, "HuGeneMixRMAMetacore", filedir=workdir, tmpdir="",
  background="antigenomic", normalize=T, exonlevel="metacore+affx")

## 3. example - exon array, e.g. HuEx-1_0-st-v2:
scheme.exon <- root.scheme(paste(scmdir, "Scheme_HuEx10stv2r2_na25.root", sep="/"))
data.exon <- root.data(scheme.exon, paste(datdir, "HuTissuesExon_cel.root", sep="/"))

workdir <- "/Volumes/GigaDrive/CRAN/Workspaces/Exon/hutissues/exon"
data.x.rma <- rma(data.exon, "MixRMAMetacore", filedir=workdir, tmpdir="", background="antigenomic",
  normalize=T, option="transcript", exonlevel="metacore")

## End(Not run)

```

## Description

ROOT system overview

## Details

ROOT is a modular object-oriented framework aimed at solving the data analysis challenges of high-energy physics. The relevant features of ROOT are as follows:

**Architecture:** The ROOT architecture is a layered class hierarchy with over 500 classes divided into different categories. Most of the classes inherit from a common base class TObject, which provides the default behavior and protocol for all objects.

**ROOT Files:** Object input/output is handled by class TFile, which has a UNIX-like directory structure and provides a hierarchical sequential and direct access persistent object store. ROOT files store information in a machine independent format and support on-the-fly data compression. Furthermore, ROOT files are self-describing: for every object stored in TFile, a dictionary describing the corresponding class is written to the file. A dictionary generator, called ROOTCINT, parses the class header files and generates a dictionary. Note: TFile can be considered to be the ROOT analogon to an R environment.

**Data Trees:** Any object derived from TObject can be written to a file with an associated key TKey. However, each key has an overhead in the directory structure in memory. To reduce this overhead, a novel concept, called Trees (class TTree) has been developed. Trees are designed to support very large numbers of complex objects in a large number of files. A Tree consists of branches (TBranch) with each branch described by its leaves (TLeaf). Trees allow direct and random access to any entry of a selected subset of branches. Thus, Trees extend and replace the usual data tables. The concept of Tree friends allows the joining of many trees as one virtual tree. However, unlike table joins in an RDBMS, the processing time is independent of the number of tree friends. Note: TTree can be considered to be the ROOT analogon to an R data.frame.

**CINT:** CINT is an interactive C/C++ interpreter, which is aimed at processing C/C++ scripts, called macros. Currently, CINT covers 99% of ANSI C and 95% of ANSI C++. CINT offers a gdb-like debugger for interpreted programs and allows the automatic compilation of scripts using ACLiC, the automatic compiler of libraries for CINT. Although available as independent program, CINT is embedded in ROOT as command line interpreter and macro processor, as well as dictionary generator.

**User interaction:** The ROOT system can be accessed from the command line, by writing macros, or via a graphic user interface (e.g. RootBrowser). Furthermore, it is possible to write libraries and applications. The ROOT GUI classes allow the development of full-featured standalone applications. Note: A macro can be considered to be the ROOT analogon of an R script. The RootBrowser can be opened using function `root.browser`

**Platform independence:** The ROOT system is available for most platforms and operating systems, including Linux, MacOS X, and the major flavors of UNIX and Windows. ROOT and ROOT-derived applications can be compiled for any supported platform.

## Author(s)

The ROOT team <http://root.cern.ch/root/Authors.html>

## References

ROOT User Guide <http://root.cern.ch/root/doc/RootDoc.html>

ROOT publications <http://root.cern.ch/root/Publications.html>



Christian Stratowa (2003), Distributed Storage and Analysis of Microarray Data in the Terabyte Range: An Alternative to BioConductor <http://www.ci.tuwien.ac.at/Conferences/DSC-2003/Proceedings/Stratowa.pdf>

---

root.browser-methods    *Open the ROOT object browser*

---

### Description

Open the [ROOT](#) object browser to see all objects stored in a [ROOT](#) file including [ROOT](#) trees.

### Usage

```
root.browser(object)
```

### Arguments

object                    an object of type [SchemeTreeSet](#), [DataTreeSet](#), [ExprTreeSet](#), or [CallTreeSet](#)

### Note

Always select menu item “Quit ROOT” from menu “File” to close the ROOT browser, otherwise you are in the CINT C/C++ interpreter from [ROOT](#). To exit CINT, you need to type “.q”.

### Author(s)

Christian Stratowa

---

root.call                    *Create class CallTreeSet accessing ROOT detection call file*

---

### Description

Create class [CallTreeSet](#) accessing [ROOT](#) detection call file.

### Usage

```
root.call(xps.scheme, rootfile = character(0), treetype = character(0), treenames = "*")
```

### Arguments

xps.scheme                A [SchemeTreeSet](#) containing the correct scheme for the [ROOT](#) data file.  
rootfile                    name of [ROOT](#) data file, including full path.  
treetype                    tree type.  
treenames                  optional character vector of tree names to get only subset of trees.

**Details**

An S4 class `CallTreeSet` will be created, serving as R wrapper to the existing `ROOT` detection call file `rootfile`.

Parameter `treetype` must be supplied to identify the `ROOT` trees for slots `data` and `detcall`. Valid tree types are listed in `validTreetype`.

To get the names of all trees with their extensions `treetype`, which are stored in `rootfile`, you can call function `getTreeNames` first.

If the `CallTreeSet` should only handle a subset of the trees stored in `rootfile`, the tree names must be supplied as vector `treenames`.

**Value**

A `CallTreeSet` object.

**Author(s)**

Christian Stratowa

**See Also**

`root.data`, `root.expr`

**Examples**

```
## first, load ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

## MAS5 detection call
detcall.mas5 <- mas5.call(data.test3, "tmp_Test3CallAll", tmpdir="", verbose=FALSE)

## use subset of trees
sub.call <- root.call(scheme.test3, "tmp_Test3CallAll.root", "dc5", c("TestA2", "TestB1"))
```

---

`root.data`

*Create class `DataTreeSet` accessing `ROOT` data file*

---

**Description**

Create class `DataTreeSet` accessing `ROOT` data file.

**Usage**

```
root.data(xps.scheme, rootfile = character(0), celnames = "*")
```

**Arguments**

<code>xps.scheme</code>	A <code>SchemeTreeSet</code> containing the correct scheme for the <code>ROOT</code> data file.
<code>rootfile</code>	name of <code>ROOT</code> data file, including full path.
<code>celnames</code>	optional character vector of tree names to get only subset of trees.

**Details**

An S4 class `DataTreeSet` will be created, serving as R wrapper to the existing `ROOT` data file `rootfile`.

If the `DataTreeSet` should only handle a subset of the trees stored in `rootfile`, the tree names must be supplied as vector `celnames`.

To get the names of all trees stored in `rootfile` you can call function `getTreeNames` first.

**Value**

A `DataTreeSet` object.

**Note**

Use `root.data` to access the `ROOT` data file from new R sessions to avoid creating a new `ROOT` data file for every R session.

**Author(s)**

Christian Stratowa

**See Also**

`import.data`, `DataTreeSet`

**Examples**

```
## get scheme and import CEL-files from package
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- import.data(scheme.test3, "tmp_datatest3", celdir=paste(path.package("xps"), "raw", sep="/"), verb=1)

## use subset of CEL-files
subdata.test3 <- root.data(scheme.test3, "tmp_datatest3_cel.root", celnames=c("TestA1.cel", "TestB2.cel"))
```

---

root.density

*ROOT Density Plot*

---

**Description**

Creates a `ROOT` density plot for one or all `ROOT` tree(s).

**Usage**

```
root.density(x, treename = "*", logbase = "log2", canvasname = "DensityPlot", save.as = "", w = 540)
```

**Arguments**

x	object of class <a href="#">DataTreeSet</a> or <a href="#">ExprTreeSet</a> .
treename	name of tree, must be present in rootfile of object x.
logbase	usually “log2”, or “0”, determines if leaf data should be converted to log.
canvasname	name of ROOT canvas
save.as	graphics type for saving canvas, one of “ps”, “eps”, “pdf”, “jpg”, “gif”, “png”, “tiff”
w	the width of the canvas in pixels.
h	the height of the canvas in pixels.

**Details**

Creates a ROOT density plot for one or all tree(s) present in rootfile.

By selecting menu “File->Save->canvasname.xxx” you can save the figure as e.g. \*.gif, \*.jpg, \*.pdf, \*.ps or even as C++ macro.

Alternatively, you can save the plot by setting save.as. However, this will close the canvas immediately after opening it.

**Note**

Always select menu item “Quit ROOT” from menu “File” to close the ROOT canvas, otherwise you are in the CINT C/C++ interpreter from [ROOT](#). To exit CINT, you need to type “.q”.

**Author(s)**

Christian Stratowa

**See Also**

[root.hist1D](#)

**Examples**

```
## Not run:
## first, load ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

root.density(data.test3, "*")
root.density(data.test3, "TestA1.cel")
root.density(data.test3, "TestA1.cel", save.as="png")

## End(Not run)
```

---

root.expr	<i>Create class ExprTreeSet accessing ROOT expression file</i>
-----------	--

---

### Description

Create class ExprTreeSet accessing ROOT expression file.

### Usage

```
root.expr(xps.scheme, rootfile = character(0), treetype = character(0), treenames = "x")
```

### Arguments

xps.scheme	A <a href="#">SchemeTreeSet</a> containing the correct scheme for the ROOT data file.
rootfile	name of ROOT data file, including full path.
treetype	tree type.
treenames	optional character vector of tree names to get only subset of trees.

### Details

An S4 class [ExprTreeSet](#) will be created, serving as R wrapper to the existing [ROOT](#) expression file rootfile.

Parameter treetype must be supplied to identify the ROOT trees for slot data. Valid tree types are listed in [validTreetype](#).

To get the names of all trees with their extensions treetype, which are stored in rootfile, you can call function [getTreeNames](#) first.

If the [ExprTreeSet](#) should only handle a subset of the trees stored in rootfile, the tree names must be supplied as vector treenames.

### Value

A ExprTreeSet object.

### Author(s)

Christian Stratowa

### See Also

[root.data](#), [root.call](#)

### Examples

```
## first, load ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

# rma
all.rma <- rma(data.test3, "tmp_Test3RMAA11", tmpdir="", background="pmonly", normalize=TRUE, verbose=FALSE)

## use subset of trees
sub.rma <- root.expr(scheme.test3, "tmp_Test3RMAA11.root", "mdp", c("TestA2.mdp", "TestB1"))
```

---

root.graph1D	<i>ROOT 1D-Graph</i>
--------------	----------------------

---

### Description

Creates a ROOT 1D-graph for a ROOT tree.

### Usage

```
root.graph1D(x, treename = character(0), logbase = "log2", option = "P", canvasname = "Graph1D", save.as = "tiff", w = 400, h = 400)
```

### Arguments

x	object of class <a href="#">DataTreeSet</a> or <a href="#">ExprTreeSet</a> .
treename	name of tree, must be present in rootfile of object x.
logbase	usually "log2", or "0", determines if leaf data should be converted to log.
option	ROOT TGraph::PaintGraph option, usually one of "P", "*", "L".
canvasname	name of ROOT canvas
save.as	graphics type for saving canvas, one of "ps", "eps", "pdf", "jpg", "gif", "png", "tiff"
w	the width of the canvas in pixels.
h	the height of the canvas in pixels.

### Details

Creates a ROOT 1D-graph for tree treename present in rootfile.

By selecting menu "File->Save->canvasname.xxx" you can save the figure as e.g. \*gif, \*.jpg, \*.pdf, \*.ps or even as C++ macro.

Alternatively, you can save the plot by setting save.as. However, this will close the canvas immediately after opening it.

### Note

Always select menu item "Quit ROOT" from menu "File" to close the ROOT canvas, otherwise you are in the CINT C/C++ interpreter from [ROOT](#). To exit CINT, you need to type ".q".

### Author(s)

Christian Stratowa

### See Also

[root.graph2D](#)

**Examples**

```
## Not run:
## first, load ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

root.graph1D(data.test3, "TestA1.cel")

## End(Not run)
```

---

root.graph2D

*ROOT 2D-Graph*


---

**Description**

Creates a ROOT 2D-graph for a ROOT tree.

**Usage**

```
root.graph2D(x, treename1 = character(0), treename2 = character(0), logbase = "log2", option = "P",
```

**Arguments**

x	object of class <a href="#">DataTreeSet</a> or <a href="#">ExprTreeSet</a> .
treename1	name of first tree, must be present in rootfile of object x.
treename2	name of second tree, must be present in rootfile of object x.
logbase	usually "log2", or "0", determines if leaf data should be converted to log.
option	ROOT TGraph::PaintGraph option, usually one of "P", "*", "L".
canvasname	name of ROOT canvas
save.as	graphics type for saving canvas, one of "ps", "eps", "pdf", "jpg", "gif", "png", "tiff"
w	the width of the canvas in pixels.
h	the height of the canvas in pixels.

**Details**

Creates a ROOT 2D-graph for trees treename1 and treename2 present in rootfile.

By selecting menu "File->Save->canvasname.xxx" you can save the figure as e.g. \*.gif, \*.jpg, \*.pdf, \*.ps or even as C++ macro.

Alternatively, you can save the plot by setting save.as. However, this will close the canvas immediately after opening it.

**Note**

Always select menu item "Quit ROOT" from menu "File" to close the ROOT canvas, otherwise you are in the CINT C/C++ interpreter from [ROOT](#). To exit CINT, you need to type ".q".

**Author(s)**

Christian Stratowa

**See Also**

[root.graph1D](#), [root.mvaplot](#)

**Examples**

```
## Not run:
## first, load ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

root.graph2D(data.test3, "TestA1.cel", "TestB1.cel")

## End(Not run)
```

---

root.hist1D	<i>ROOT 1D-Histogram</i>
-------------	--------------------------

---

**Description**

Creates a ROOT 1D-histogram for a ROOT tree.

**Usage**

```
root.hist1D(x, treename = character(0), logbase = "log2", type = "hist", option = "HIST", canvasname = character(0))
```

**Arguments**

x	object of class <a href="#">DataTreeSet</a> or <a href="#">ExprTreeSet</a> .
treename	name of tree, must be present in rootfile of object x.
logbase	usually "log2", or "0", determines if leaf data should be converted to log.
type	ROOT 1D-hist or density, i.e. "hist" or "density".
option	ROOT 1D-hist option only, usually one of "HIST", "B", "C", "E".
canvasname	name of ROOT canvas
save.as	graphics type for saving canvas, one of "ps", "eps", "pdf", "jpg", "gif", "png", "tiff"
w	the width of the canvas in pixels.
h	the height of the canvas in pixels.

**Details**

Creates a ROOT 1D-histogram for tree `treename` present in `rootfile`.

By selecting menu "File->Save->canvasname.xxx" you can save the figure as e.g. \*.gif, \*.jpg, \*.pdf, \*.ps or even as C++ macro.

Alternatively, you can save the plot by setting `save.as`. However, this will close the canvas immediately after opening it.

**Note**

Always select menu item "Quit ROOT" from menu "File" to close the ROOT canvas, otherwise you are in the CINT C/C++ interpreter from [ROOT](#). To exit CINT, you need to type ".q".



**Author(s)**

Christian Stratowa

**See Also**[root.hist2D](#), [root.hist3D](#)**Examples**

```
## Not run:
## first, load ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

root.hist1D(data.test3, "TestA1.cel")
root.hist1D(data.test3, "TestA1.cel", type="density")

## End(Not run)
```

root.hist2D

*ROOT 2D-Histogram***Description**

Creates a ROOT 2D-histogram for a ROOT tree.

**Usage**

```
root.hist2D(x, treename1 = character(0), treename2 = character(0), logbase = "log2", option = "COLZ")
```

**Arguments**

x	object of class <a href="#">DataTreeSet</a> or <a href="#">ExprTreeSet</a> .
treename1	name of first tree, must be present in rootfile of object x.
treename2	name of second tree, must be present in rootfile of object x.
logbase	usually "log2", or "0", determines if leaf data should be converted to log.
option	ROOT hist TH2 option, usually one of "SCAT", "COLZ", "BOX", "SURF2", "SURF3".
canvasname	name of ROOT canvas
save.as	graphics type for saving canvas, one of "ps", "eps", "pdf", "jpg", "gif", "png", "tiff"
w	the width of the canvas in pixels.
h	the height of the canvas in pixels.

**Details**

Creates a ROOT 2D-histogram for trees treename1 and treename2 present in rootfile.

By selecting menu "File->Save->canvasname.xxx" you can save the figure as e.g. \*.gif, \*.jpg, \*.pdf, \*.ps or even as C++ macro.

Alternatively, you can save the plot by setting save.as. However, this will close the canvas immediately after opening it.

**Note**

Always select menu item “Quit ROOT” from menu “File” to close the ROOT canvas, otherwise you are in the CINT C/C++ interpreter from ROOT. To exit CINT, you need to type “.q”.

**Author(s)**

Christian Stratowa

**See Also**

[root.hist1D](#), [root.hist3D](#)

**Examples**

```
## Not run:
## first, load ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

root.hist2D(data.test3, "TestA1.cel", "TestB1.cel", option="COLZ")

## End(Not run)
```

---

root.hist3D

*ROOT 3D-Histogram*

---

**Description**

Creates a ROOT 3D-histogram for a ROOT tree.

**Usage**

```
root.hist3D(x, treename1 = character(0), treename2 = character(0), treename3 = character(0), logbase
```

**Arguments**

x	object of class <a href="#">DataTreeSet</a> or <a href="#">ExprTreeSet</a> .
treename1	name of first tree, must be present in rootfile of object x.
treename2	name of second tree, must be present in rootfile of object x.
treename3	name of third tree, must be present in rootfile of object x.
logbase	usually “log2”, or “0”, determines if leaf data should be converted to log.
option	ROOT hist TH3 option, usually one of “HIST”, “SCAT”, “BOX”.
canvasname	name of ROOT canvas
save.as	graphics type for saving canvas, one of “ps”, “eps”, “pdf”, “jpg”, “gif”, “png”, “tiff”
w	the width of the canvas in pixels.
h	the height of the canvas in pixels.

## Details

Creates a ROOT 3D-histogram for trees `treename1`, `treename2` and `treename3` present in `rootfile`. By selecting menu “File->Save->canvasname.xxx” you can save the figure as e.g. \*.gif, \*.jpg, \*.pdf, \*.ps or even as C++ macro.

By moving the mouse into the middle of the canvas, the cursor changes and you can rotate the 3D-histogram. By selecting menu “View->View With->OpenGL” the OpenGL viewer opens, where you can rotate the 3D-histogram interactively.

Alternatively, you can save the plot by setting `save.as`. However, this will close the canvas immediately after opening it.

## Note

Always select menu item “Quit ROOT” from menu “File” to close the ROOT canvas, otherwise you are in the CINT C/C++ interpreter from [ROOT](#). To exit CINT, you need to type “.q”.

## Author(s)

Christian Stratowa

## See Also

[root.hist1D](#), [root.hist2D](#)

## Examples

```
## Not run:
## first, load ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

root.hist3D(data.test3, "TestA1.cel", "TestB2.cel", "TestB1.cel", option="BOX")

## End(Not run)
```

---

root.image

*ROOT Image*

---

## Description

Creates a ROOT image for a ROOT tree.

## Usage

```
root.image(x, treename = character(0), leafname = "fInten", logbase = "log2", option = "COLZ", zlin
```

**Arguments**

x	object of class <a href="#">DataTreeSet</a> .
treename	name of tree, must be present in rootfile of object x.
leafname	leaf name of tree, usual "fInten" or "fBg".
logbase	usually "log2", or "0", determines if leaf data should be converted to log.
option	ROOT graph option, usually. one of "COL", "COLZ".
zlim	size limits c(min,max) of leafname.
canvasname	name of ROOT canvas
save.as	graphics type for saving canvas, one of "ps", "eps", "pdf", "jpg", "gif", "png", "tiff"
w	the width of the device in pixels.
h	the height of the device in pixels.

**Details**

Creates a ROOT image for tree `treename` present in `rootfile`.

To zoom-in move the mouse cursor to the x-axis (y-axis) until it changes to a hand and click-drag to select an axis-range. To unzoom move the mouse cursor to the x-axis (y-axis) until it changes to a hand and right-click to select "Unzoom".

By selecting menu "File->Save->canvasname.xxx" you can save the figure as e.g. \*.gif, \*.jpg, \*.pdf, \*.ps or even as C++ macro.

Alternatively, you can save the plot by setting `save.as`. However, this will close the canvas immediately after opening it.

**Note**

Always select menu item "Quit ROOT" from menu "File" to close the ROOT canvas, otherwise you are in the CINT C/C++ interpreter from ROOT. To exit CINT, you need to type ".q".

**Author(s)**

Christian Stratowa

**See Also**

[image-methods](#), [image](#)

**Examples**

```
## Not run:
## first, load ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

root.image(data.test3, "TestA1.cel")
root.image(data.test3, "TestA1.cel", save.as="png")

## End(Not run)
```

---

root.merge.data	Create class <i>DataTreeSet</i> by merging <i>ROOT</i> data files
-----------------	---

---

### Description

Create class *DataTreeSet* by merging different *ROOT* data files.

### Usage

```
root.merge.data(xps.scheme, rootfiles = list(), celnames = "*")
```

### Arguments

xps.scheme	A <a href="#">SchemeTreeSet</a> containing the correct scheme for the <i>ROOT</i> data file.
rootfiles	list of <i>ROOT</i> data file(s), including full path.
celnames	optional character vector of tree names to get only subset of trees.

### Details

This function allows to merge data trees from different existing *ROOT* data files.

An S4 class [DataTreeSet](#) will be created, serving as R wrapper to the existing *ROOT* data file(s) *rootfiles*.

If the [DataTreeSet](#) should only handle a subset of the trees stored in *rootfiles*, the tree names must be supplied as vector *celnames*.

To get the names of all trees stored in separate *rootfiles* you can call function [getTreeNames](#) first.

### Value

A *DataTreeSet* object.

### Author(s)

Christian Stratowa

### See Also

[root.data](#), [DataTreeSet](#)

### Examples

```
## get scheme and import CEL-files from package
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- import.data(scheme.test3, "tmp_datatest3", celdir=paste(path.package("xps"), "raw", sep="/"), verb=0)

## get subset of CEL-files
subdataA <- root.data(scheme.test3, "tmp_datatest3_cel.root", celnames=c("TestA1.cel", "TestA2.cel"))
subdataB <- root.data(scheme.test3, "tmp_datatest3_cel.root", celnames=c("TestB1.cel", "TestB2.cel"))

## merge data
dataAB <- root.merge.data(scheme.test3, c(rootFile(subdataA), rootFile(subdataB)), celnames=c("TestB1.cel", "TestB2.cel"))
```

---

root.mvaplot	<i>ROOT M vs A Plot</i>
--------------	-------------------------

---

### Description

Creates a ROOT M vs A plot for a ROOT tree.

### Usage

```
root.mvaplot(x, treename1 = character(0), treename2 = character(0), logbase = "log2", option = "P",
```

### Arguments

x	object of class <a href="#">ExprTreeSet</a> or <a href="#">DataTreeSet</a> .
treename1	name of first tree, must be present in rootfile of object x.
treename2	name of second tree, must be present in rootfile of object x.
logbase	usually "log2", or "0", determines if leaf data should be converted to log.
option	ROOT TGraph::PaintGraph option, usually one of "P", "*".
canvasname	name of ROOT canvas
save.as	graphics type for saving canvas, one of "ps", "eps", "pdf", "jpg", "gif", "png", "tiff"
w	the width of the canvas in pixels.
h	the height of the canvas in pixels.

### Details

Creates a ROOT M vs A plot for trees treename1 and treename2 present in rootfile.

By selecting menu "File->Save->canvasname.xxx" you can save the figure as e.g. \*gif, \*.jpg, \*.pdf, \*.ps or even as C++ macro.

Alternatively, you can save the plot by setting save.as. However, this will close the canvas immediately after opening it.

### Note

Always select menu item "Quit ROOT" from menu "File" to close the ROOT canvas, otherwise you are in the CINT C/C++ interpreter from [ROOT](#). To exit CINT, you need to type ".q".

### Author(s)

Christian Stratowa

### See Also

[root.graph1D](#)

**Examples**

```
## Not run:
## first, load ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

# compute RMA
data.rma <- rma(data.test3, "Test3RMA", tmpdir="", background="pmonly", normalize=TRUE)

root.mvplot(data.rma, "TestA1.mdp", "TestB1.mdp")

## End(Not run)
```

---

root.profile	<i>ROOT Profile Plot</i>
--------------	--------------------------

---

**Description**

Creates a ROOT profile plot, i.e. a plot of parallel coordinates

**Usage**

```
root.profile(x, treename = "*", varlist = NULL, as.log = TRUE, globalscale = TRUE, boxes = TRUE, ylim
```

**Arguments**

x	S4 object, usually of class <a href="#">DataTreeSet</a> or <a href="#">ExprTreeSet</a> .
treename	name of tree, usually all trees present in rootfile of object x.
varlist	leaf name of tree, usual "fInten" or "fLevel".
as.log	logical indicating if varlist should be drawn as logarithmic data.
globalscale	logical indicating if all axes should be drawn at the same scale.
boxes	logical indicating if box-and-whisker plots should be drawn.
ylim	size limits c(min,max) of varlist.
canvasname	name of ROOT canvas
save.as	graphics type for saving canvas, one of "ps", "eps", "pdf", "jpg", "gif", "png", "tiff"
w	the width of the device in pixels.
h	the height of the device in pixels.

**Details**

Creates a ROOT profile plot for all trees treename="\*" present in rootfile, or for a subset of trees. In this case varlist must be the name of one tree leaf only; for varlist=NULL leaf "fInten" will be used for class [DataTreeSet](#) and leaf "fLevel" will be used for class [ExprTreeSet](#).

If treename is the name of one tree only then varlist can contain up to all leaves of the tree, separated by colons, e.g. varlist="fLevel:fStdev".

For boxes=TRUE the profile plot draws box-and-whisker plots and can thus be considered the equivalent of the usual boxplot.

A ROOT profile plot, i.e. a plot of parallel coordinates, is drawn in a “TreeView”, a graphic user interface designed to handle ROOT trees. You can activate context menus by right-clicking on items or inside the right panel.

The “TreeView” is explained in <http://root.cern.ch/root/html/TTreeView.html>.

By selecting menu “File->Save->canvasname.xxx” you can save the figure as e.g. \*.gif, \*.jpg, \*.pdf, \*.ps or even as C++ macro.

Alternatively, you can save the plot by setting save.as. However, this will close the canvas immediately after opening it.

### Note

Always select menu item “Quit ROOT” from menu “File” to close the ROOT tree viewer, otherwise you are in the CINT C/C++ interpreter from ROOT. To exit CINT, you need to type “.q”.

### Author(s)

Christian Stratowa

### Examples

```
## Not run:
## first, load ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

root.profile(data.test3)

## End(Not run)
```

---

root.scheme

*Create class SchemeTreeSet accessing ROOT scheme file*

---

### Description

Create class SchemeTreeSet accessing ROOT scheme file.

### Usage

```
root.scheme(rootfile = character(0), add.mask = FALSE)
```

### Arguments

rootfile	name of ROOT scheme file, including full path.
add.mask	if TRUE mask information will be included as slot mask.

### Details

An S4 class `SchemeTreeSet` will be created, serving as R wrapper to the ROOT scheme file rootfile.

### Value

A SchemeTreeSet object.



**Note**

Use this function to access the [ROOT](#) scheme file from new R sessions to avoid creating a new [ROOT](#) scheme file for every R session.

Do not set `add.mask=TRUE` for exon arrays unless you know that your computer has sufficient RAM.

**Author(s)**

Christian Stratowa

**See Also**

[import.expr.scheme](#), [import.exon.scheme](#), [SchemeTreeSet](#)

**Examples**

```
## create class SchemeSet to access the ROOT scheme file for the Test3 GeneChip
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
str(scheme.test3)

## Not run:
## scheme set for existing human root exon scheme file
scheme.huex10stv2r2.na22 <- root.scheme("/my/path/schemes/Scheme_HuEx10stv2r2_na22.root")

## End(Not run)
```

---

SchemeTreeSet-class    *Class SchemeTreeSet*

---

**Description**

This class provides the link to the [ROOT](#) scheme file and the [ROOT](#) trees contained therein. It extends class [TreeSet](#).

**Objects from the Class**

Objects can be created using the functions [import.expr.scheme](#), [import.exon.scheme](#), [import.genome.scheme](#) or [root.scheme](#).

**Slots**

**chipname:** Object of class "character" representing the Affymetrix chip name.

**chiptype:** Object of class "character" representing the chip type, either 'GeneChip', 'GenomeChip' or 'ExonChip'.

**probeinfo:** Object of class "list" representing chip information, including nrows, ncols, number of probes, etc.

**unitname:** Object of class "data.frame". The data.frame can contain the mapping between the internal UNIT\_IDs and the UnitNames, i.e. the probeset IDs.

**mask:** Object of class "data.frame". The data.frame can contain the mask used to identify the probes as e.g. PM, MM or control probes.

**probe:** Object of class "data.frame". The data.frame can contain the probe info for the oligos as e.g. probe sequence, G/C content.

**setname:** Object of class "character" representing the name to the **ROOT** file subdirectory where the **ROOT** scheme trees are stored; it is identical to chipname.

**settype:** Object of class "character" describing the type of treeset stored in setname, i.e. 'scheme'.

**rootfile:** Object of class "character" representing the name of the **ROOT** scheme file, including full path.

**filedir:** Object of class "character" describing the full path to the system directory where rootfile is stored.

**numtrees:** Object of class "numeric" representing the number of **ROOT** trees stored in subdirectory setname.

**treenames:** Object of class "list" representing the names of the **ROOT** trees stored in subdirectory setname.

### Extends

Class "**TreeSet**", directly.

### Methods

**attachMask** signature(object = "SchemeTreeSet"): exports scheme tree from **ROOT** scheme file and and saves as data.frame mask.

**attachProbe** signature(object = "SchemeTreeSet"): exports probe tree from **ROOT** scheme file and and saves varlist as data.frame probe.

**attachProbeContentGC** signature(object = "SchemeTreeSet"): exports probe tree from **ROOT** scheme file and and saves fNumberGC as data.frame probe.

**attachProbeSequence** signature(object = "SchemeTreeSet"): exports probe tree from **ROOT** scheme file and and saves fSequence as data.frame probe.

**attachUnitNames** signature(object = "SchemeTreeSet"): exports unit tree from **ROOT** scheme file and and saves as data.frame unitname.

**chipMask** signature(object = "SchemeTreeSet"): extracts data.frame mask.

**chipMask<-** signature(object = "SchemeTreeSet", value = "data.frame"): replaces data.frame mask.

**chipName** signature(object = "SchemeTreeSet"): extracts slot chipname.

**chipProbe** signature(object = "SchemeTreeSet"): extracts data.frame probe.

**chipProbe<-** signature(object = "SchemeTreeSet", value = "data.frame"): replaces data.frame probe.

**chipType** signature(object = "SchemeTreeSet"): extracts slot chiptype.

**chipType<-** signature(object = "SchemeTreeSet", value = "character"): replaces slot chiptype.

**export** signature(object = "SchemeTreeSet"): exports **ROOT** trees as text file, see [export-methods](#).

**ncols** signature(object = "SchemeTreeSet"): extracts the physical number of array columns from slot probeinfo.

**nrows** signature(object = "SchemeTreeSet"): extracts the physical number of array rows from slot probeinfo.

**probeContentGC** signature(object = "SchemeTreeSet"): extracts all or selected GC contents from data.frame probe.

**probeInfo** signature(object = "SchemeTreeSet"): extracts slot probeinfo.

**probeSequence** signature(object = "SchemeTreeSet"): extracts all or selected probe sequences from data.frame probe.

**probesetID2unitID** signature(object = "SchemeTreeSet"): extracts all or selected probesetIDs from data.frame unitname with UnitName, i.e. probeset ID, as (row)names.

**removeMask** signature(object = "SchemeTreeSet"): replaces data.frame mask with an empty data.frame of dim(0,0).

**removeProbe** signature(object = "SchemeTreeSet"): replaces data.frame probe with an empty data.frame of dim(0,0).

**removeProbeContentGC** signature(object = "SchemeTreeSet"): replaces data.frame probe with an empty data.frame of dim(0,0).

**removeProbeSequence** signature(object = "SchemeTreeSet"): replaces data.frame probe with an empty data.frame of dim(0,0).

**removeUnitNames** signature(object = "SchemeTreeSet"): replaces data.frame unitname with an empty data.frame of dim(0,0).

**symbol2unitID** signature(object = "SchemeTreeSet"): extracts internal UNIT\_ID(s) for one or more gene symbols.

**transcriptID2unitID** signature(object = "SchemeTreeSet"): extracts all or selected transcriptIDs from data.frame unitname with UnitName, i.e. transcript ID, as (row)names.

**unitID2probesetID** signature(object = "SchemeTreeSet"): extracts all or selected unitIDs from data.frame unitname with UNIT\_ID as (row)names.

**symbol2unitID** signature(object = "SchemeTreeSet"): extracts gene symbols for one or more internal UNIT\_ID(s).

**unitID2transcriptID** signature(object = "SchemeTreeSet"): extracts all or selected unitIDs from data.frame unitname with UNIT\_ID as (row)names.

**unitNames** signature(object = "SchemeTreeSet"): extracts data.frame unitname.

**unitNames<-** signature(object = "SchemeTreeSet", value = "data.frame"): replaces data.frame unitname.

**Author(s)**

Christian Stratowa

**Examples**

```
showClass("SchemeTreeSet")
```

---

summarize

*Probe Set Summarizing Functions*

---

**Description**

Converts Affymetrix probe level data to expression levels by summarizing the probe set values into one expression measure and a standard error for this summary.

**Usage**

```

summarize(xps.data, filename = character(0), filedir = getwd(), tmpdir = "", update = FALSE, select,
summarize.mas4(xps.data, filename = character(0), filedir = getwd(), tmpdir = "", update = FALSE,
summarize.mas5(xps.data, filename = character(0), filedir = getwd(), tmpdir = "", update = FALSE,
summarize.rma(xps.data, filename = character(0), filedir = getwd(), tmpdir = "", update = FALSE,
xpsSummarize(object, ...)

```

**Arguments**

xps.data	object of class <code>DataTreeSet</code> .
filename	file name of ROOT data file.
filedir	system directory where ROOT data file should be stored.
tmpdir	optional temporary directory where temporary ROOT files should be stored.
update	logical. If TRUE the existing ROOT data file filename will be updated.
select	type of probes to select for summarization.
method	summarization method to use.
option	option determining the grouping of probes for summarization, one of 'transcript', 'exon', 'probeset'; exon arrays only.
logbase	logarithm base as character, one of '0', 'log', 'log2', 'log10'.
exonlevel	exon annotation level determining which probes should be used for summarization; exon/genome arrays only.
params	vector of parameters for summarization method.
xps.scheme	optional alternative <code>SchemeTreeSet</code> .
add.data	logical. If TRUE expression data will be included as slot data.
verbose	logical, if TRUE print status information.
object	object of class <code>DataTreeSet</code> .
...	the arguments described above.

**Details**

Converts Affymetrix probe level data to expression levels by summarizing the probe set values into one expression measure and a standard error for this summary.

`xpsSummarize` is the `DataTreeSet` method called by function `summarize`, containing the same parameters.

**Value**

An [ExprTreeSet](#).

**Author(s)**

Christian Stratowa

**See Also**[express](#)**Examples**

```
## first, load ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

## RMA background
data.bg.rma <- bgcorrect.rma(data.test3, "tmp_Test3RMA", filedir=getwd(), tmpdir="", verbose=FALSE)
## normalize quantiles
data.qu.rma <- normalize.quantiles(data.bg.rma, "tmp_Test3RMA", filedir=getwd(), tmpdir="", update=TRUE, verbose=FALSE)
## summarize medianpolish
data.mp.rma <- summarize.rma(data.qu.rma, "tmp_Test3RMA", filedir=getwd(), tmpdir="", update=TRUE, verbose=FALSE)

## get expression data.frame
expr.rma <- exprs(data.mp.rma)
head(expr.rma)

## plot expression levels
if (interactive()) {
  boxplot(data.mp.rma)
  boxplot(log2(expr.rma[,3:6]))
}
```

---

symbol2unitID-methods *Conversion between Gene Symbols and UnitIDs*

---

**Description**

Convert gene symbols to internal UNIT\_IDs and vice verse.

*Usage*

```
symbol2unitID(object, symbol, unittype = "transcript", as.list = TRUE)
unitID2symbol(object, unitID, unittype = "transcript", as.list = TRUE)
```

**Arguments**

object	Object of class "SchemeTreeSet" or "DataTreeSet".
symbol	character vector of gene symbol(s).
unitID	vector of UNIT_IDs.
unittype	character vector, "transcript" or "probeset".
as.list	if TRUE a list will be returned (default is data.frame).

**Details**

Functions symbol2unitID and unitID2symbol returns the UNIT\_ID(s) for selected gene symbols and vice verse.

For exon arrays the internal UNIT\_ID(s) depend on unittype.

By default a list is returned, however for as.list=FALSE a character vector of IDs is returned.

**Value**

A list or character vector.

**Author(s)**

Christian Stratowa

**See Also**

[transcriptID2unitID](#), [probesetID2unitID](#)

**Examples**

```
## load ROOT scheme file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))

## unitnames not attached
id <- symbol2unitID(scheme.test3, symbol="ACTB", as.list=TRUE)
id
id <- unitID2symbol(scheme.test3, unitID=274, as.list=TRUE)
id

## unitnames attached
scheme.test3 <- attachUnitNames(scheme.test3)
id <- symbol2unitID(scheme.test3, symbol="ACTB", as.list=TRUE)
id
id <- unitID2symbol(scheme.test3, unitID=274, as.list=TRUE)
id
scheme.test3 <- removeUnitNames(scheme.test3)

rm(scheme.test3)
gc()
```

---

treeInfo-methods

*Get UserInfo from ROOT Trees*

---

**Description**

Extract the UserInfo from [ROOT](#) trees, i.e. quality control information.

*Usage*

```
treeInfo(object,          treename = "*",          treetype = character(0),          varlist = "*",
```

**Arguments**

object	Object of class "TreeSet".
treename	Object of class "list" representing the names of the <a href="#">ROOT</a> trees.
treetype	type of tree to export, see <a href="#">validTreetype</a>
varlist	names of tree leaves to export.
qualopt	option determining the data to which to apply qualification, one of 'raw', 'adjusted', 'normalized', 'all'.

## Details

**ROOT** trees have a pointer to a list `fUserInfo` where it is possible to store data which do not fit into the usual tree structure. Taking advantage of this feature `xps` stores certain pre-processed results of the tree(s) in this list. For example, data trees store the minimal/maximal intensities and the number of oligos with minimal/maximal intensities of the CEL-files in list `fUserInfo`, while call trees store the number and percentage of P/M/A calls.

Function `treeInfo` allows to export this user information as a `data.frame`, whereby the parameters of `varlist` depend on the `treetype`:

Parameters for data trees with extensions "cel", "int", and background trees:

`fMinInten`: minimal intensity.  
`fMaxInten`: maximal intensity.  
`fNMinInten`: number of probes with minimal intensity.  
`fNMaxInten`: number of probes with maximal intensity.  
`fMaxNPixels`: maximal number of pixels.  
`fNQuantiles`: number of precalculated quantiles.  
`fQuantiles`: quantiles.  
`fIntenQuant`: intensities at quantiles.

Parameters for expression trees:

`fNUnits`: number of units, i.e. probesets.  
`fMinLevel`: minimal expression level.  
`fMaxLevel`: maximal expression level.  
`fNQuantiles`: number of precalculated quantiles.  
`fQuantiles`: quantiles.  
`fLevelQuant`: expression levels at quantiles.

Parameters for call trees:

`fNUnits`: number of units, i.e. probesets.  
`fNAbsent`: number of units with absent call.  
`fNMarginal`: number of units with marginal call.  
`fNPresent`: number of units with present call.  
`fPcAbsent`: percentage of units with absent call.  
`fPcMarginal`: percentage of units with marginal call.  
`fPcPresent`: percentage of units with present call.  
`fMinPValue`: minimal p-value.  
`fMaxPValue`: maximal p-value.

Parameters for border trees with extension "brd":

`fMeanLeft`: mean intensity of left border.  
`fMeanRight`: mean intensity of right border.  
`fMeanTop`: mean intensity of top border.  
`fMeanBottom`: mean intensity of bottom border.  
`fCOIXhi`: x-location of COI for the positive elements.  
`fCOIYhi`: y-location of COI for the positive elements.  
`fCOIXlo`: x-location of COI for the negative elements.  
`fCOIYlo`: y-location of COI for the negative elements.

Parameters for quality trees with extension "rlm":

`fNUnits`: number of units, i.e. probesets.  
`fMinLevel`: minimal expression level.  
`fMaxLevel`: maximal expression level.  
`fNQuantiles`: number of precalculated quantiles.  
`fQuantiles`: quantiles.  
`fLevelQuant`: expression levels at quantiles.

fNUSEQuant: NUSE at quantiles.  
 fRLEQuant: RLE at quantiles.  
 fQualOption: value of qualopt.

Parameters for residual trees with extension "res":

fNQuantiles: number of precalculated quantiles.  
 fQuantiles: quantiles.  
 fResiduQuant: residual at quantiles.  
 fWeightQuant: weight at quantiles.  
 fQualOption: value of qualopt.

### Value

A data.frame.

### Note

Taking advantage of function treeInfo plotting methods [boxplot](#), [callplot](#), [coiplot](#), [nuseplot](#) and [rleplot](#) are able to display their results much faster, which is especially useful for large datasets.

### Author(s)

Christian Stratowa

### See Also

[validTreetype](#)

### Examples

```
## load existing ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

userinfo <- treeInfo(data.test3, treetype="cel", varlist="*")
userinfo

userinfo <- treeInfo(data.test3, treename="TestB1", treetype="cel", varlist = "fNQuantiles:fIntenQuant")
userinfo

## Not run:
userinfo <- treeInfo(rlm.all, treetype="rlm", varlist = "fNQuantiles:fNUSEQuant:fRLEQuant", qualopt = "raw")
userinfo

userinfo <- treeInfo(rlm.all, treetype="brd")
userinfo

userinfo <- treeInfo(rlm.all, treetype="res", qualopt = "raw")
userinfo

userinfo <- treeInfo(rlm.all, treetype="res", varlist = "fResiduQuant", qualopt = "raw")
userinfo

## End(Not run)
```



---

TreeSet-class	<i>Class TreeSet</i>
---------------	----------------------

---

### Description

This is the virtual base class for all other classes providing the link to a [ROOT](#) file and the [ROOT](#) trees contained therein.

### Objects from the Class

A virtual Class: No objects may be created from it.

### Slots

**setname:** Object of class "character" representing the name to the [ROOT](#) file subdirectory where the [ROOT](#) trees are stored, usually one of 'DataTreeSet', 'PreprocesSet', 'CallTreeSet'.

**settype:** Object of class "character" describing the type of treeset stored in setname, usually one of 'scheme', 'rawdata', 'preprocess'.

**rootfile:** Object of class "character" representing the name of the [ROOT](#) file, including full path.

**filedir:** Object of class "character" describing the full path to the system directory where rootfile is stored.

**numtrees:** Object of class "numeric" representing the number of [ROOT](#) trees stored in subdirectory setname.

**treenames:** Object of class "list" representing the names of the [ROOT](#) trees stored in subdirectory setname.

### Methods

**export** signature(object = "TreeSet"): exports [ROOT](#) trees as text file, see [export-methods](#).

**fileDir** signature(object = "TreeSet"): extracts slot filedir.

**fileDir<-** signature(object = "TreeSet", value = "character"): replaces slot filedir.

**root.browser** signature(object = "TreeSet"): opens the [ROOT](#) file browser.

**rootFile** signature(object = "TreeSet"): extracts slot rootfile.

**rootFile<-** signature(object = "TreeSet", value = "character"): replaces slot rootfile.

**setName** signature(object = "TreeSet"): extracts slot setname.

**setName<-** signature(object = "TreeSet", value = "character"): replaces slot setname.

**setType** signature(object = "TreeSet"): extracts slot settype.

**setType<-** signature(object = "TreeSet", value = "character"): replaces slot settype.

**treeInfo** signature(object = "TreeSet"): extracts UserInfo from [ROOT](#) trees.

**treeNames** signature(object = "TreeSet"): extracts slot treenames.

### Author(s)

Christian Stratowa

### See Also

derived classes [SchemeTreeSet](#), [DataTreeSet](#), [ExprTreeSet](#), [CallTreeSet](#).

**Examples**

```
showClass("TreeSet")
```

---

trma

*transposed Robust Multi-Array Average Expression Measure*


---

**Description**

This function converts a [DataTreeSet](#) into an [ExprTreeSet](#) using the transposed robust multi-array average (RMA) expression measure.

**Usage**

```
trma(xps.data,
     filename = character(0),
     filedir  = getwd(),
     tmpdir   = "",
     background = "pmonly",
     normalize = TRUE,
     option    = "transcript",
     exonlevel = "",
     params    = list(16384, 0.0, 1.0, 10, 0.01, 2),
     xps.scheme = NULL,
     add.data  = TRUE,
     verbose   = TRUE)
```

**Arguments**

xps.data	object of class <a href="#">DataTreeSet</a> .
filename	file name of ROOT data file.
filedir	system directory where ROOT data file should be stored.
tmpdir	optional temporary directory where temporary ROOT files should be stored.
background	probes used to compute background, one of 'pmonly', 'mmonly', 'both'; for genome/exon arrays one of 'genomic', 'antigenomic'
normalize	logical. If TRUE normalize data using quantile normalization.
option	option determining the grouping of probes for summarization, one of 'transcript', 'exon', 'probeset'; exon arrays only.
exonlevel	exon annotation level determining which probes should be used for summarization; exon/genome arrays only.
params	list of (default) parameters for rma.
xps.scheme	optional alternative SchemeTreeSet.
add.data	logical. If TRUE expression data will be included as slot data.
verbose	logical, if TRUE print status information.

**Details**

This function computes the tRMA (transposed Robust Multichip Average) expression measure described in Giorgi et al. for both expression arrays and exon arrays.

To use method xpsRMA or function express to compute trma you need to set `params = list(16384, 0.0, 1.0, 10, 0.0)`

For further details please see [rma](#)

**Value**

An [ExprTreeSet](#)

**Author(s)**

Christian Stratowa

**References**

Federico M. Giorgi, Anthony M. Bolger, Marc Lohse and Bjoern Usadel (2010), Algorithm-driven Artifacts in median polish summarization of Microarray data. BMC Bioinformatics 11:553

**See Also**

[rma](#), [xpsRMA](#), [express](#)

**Examples**

```
## first, load ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

data.trma <- trma(data.test3, "tmp_Test3tRMA", tmpdir="", background="pmonly", normalize=TRUE, verbose=FALSE)

## get data.frame
expr.trma <- validData(data.trma)
head(expr.trma)

rm(scheme.test3, data.test3)
gc()
```

---

type2Exten

*Convert Method Type to Tree Extension*


---

**Description**

Convert Method Type to Tree Extension.

**Usage**

```
type2Exten(type, datatype)
```

**Arguments**

type	method type.
datatype	data type.

**Details**

For every datatype different methods, i.e. algorithms exist which can be applied. Valid datatypes are 'preprocess' and 'normation'.

For datatype 'preprocess' the following methods can be applied:

mean:	trimmed mean
median:	median
quantile:	quantile
tukeybiweight:	tukey biweight
medianpolish:	median polish

For datatype 'normation' the following methods can be applied:

mean:	trimmed mean
median:	median
quantile:	quantile
lowess:	lowess
supsmu:	supsmu

The tree extensions are described in [validTreetype](#).

**Value**

A character with the correct tree extension.

**Author(s)**

Christian Stratowa

**See Also**

[getDatatype](#), [validTreetype](#)

**Examples**

```
type2Exten("quantile", "preprocess")
type2Exten("medianpolish", "preprocess")
type2Exten("supsmu", "normation")
```

---

unifilter

*Function for Applying an UniFilter to an ExprTreeSet*


---

**Description**

This function applies an [UniFilter](#) to an [ExprTreeSet](#).

**Usage**

```
unifilter(xps.expr,
         filename = character(0),
         filedir  = getwd(),
         filter   = NULL,
         minfilters = 999,
         logbase  = "log2",
         group    = character(0),
         treename = "UniTest",
         xps.fltr = NULL,
         xps.call = NULL,
         update   = FALSE,
         verbose  = TRUE)
```

```
xpsUniFilter(object, ...)
```

**Arguments**

xps.expr	object of class ExprTreeSet.
filename	file name of ROOT filter file.
filedir	system directory where ROOT filter file should be stored.
filter	object of class UniFilter.
minfilters	minimum number of initialized filter methods to satisfy (default is all filters).
logbase	convert data to logarithm of base: "0", "log", "log2" (default), "log10"
group	a character vector assigning the trees of xps.expr to one of two groups.
treename	tree name to be used in ROOT filter file.
xps.fltr	optional object of class FilterTreeSet.
xps.call	optional object of class CallTreeSet.
update	logical. If TRUE the existing ROOT filter file filename will be updated.
verbose	logical, if TRUE print status information.
object	object of class ExprTreeSet.
...	same arguments as function unifilter.

**Details**

This function applies the different filters initialized with constructor [UniFilter](#) to the [ExprTreeSet](#) xps.expr.

Slot minfilters determines the minimum number of initialized filters, which must be satisfied so that the mask is set to flag=1. For minfilters=1 at least one filter must be satisfied, equivalent to logical 'OR'; for minfilters=999 all filters must be satisfied, equivalent to logical 'AND'.

If pre-filtering should be done before applying function unifilter then a [FilterTreeSet](#) xps.fltr must be supplied, created with function [prefilter](#).

If method [callFilter](#) was initialized with constructor [UniFilter](#) then [CallTreeSet](#) xps.call must be supplied, usually created with function [mas5.call](#).

**Value**

An [AnalysisTreeSet](#)

**Note**

Internally, slot group will be converted to integer values using `as.integer(as.factor(group))`, thus `group=c("GrpA", "GrpA", "GrpB", "GrpB")` will result in a fold-change of `fc=mean(GrpB)/mean(GrpA)`.

**Author(s)**

Christian Stratowa

**See Also**

[UniFilter](#), [prefilter](#)

**Examples**

```
## Not run:
## first, load ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

## second, create an ExprTreeSet
data.rma <- rma(data.test3, "tmp_Test3_RMA", tmpdir="", background="pmonly", normalize=TRUE, verbose=FALSE)
## note: do not copy/paste this code, it is necessary only because R CMD check fails since it does not find tmp
data.rma@rootfile <- paste(path.package("xps"), "rootdata/tmp_Test3RMA.root", sep="/")
data.rma@filedir <- paste(path.package("xps"), "rootdata", sep="/")

## third, construct an UniFilter
unifltr <- UniFilter(unitest=c("t.test", "two.sided", "none", 0, 0.0, FALSE, 0.95, TRUE), foldchange=c(1.3, "both"),

## finally, create an AnalysisTreeSet
rma.ufr <- unifilter(data.rma, "tmp_Test3Unifilter", getwd(), unifltr, group=c("GrpA", "GrpA", "GrpB", "GrpB"), ver
str(rma.ufr)

## End(Not run)
```

---

UniFilter-class

*Class UniFilter*

---

**Description**

Class UniFilter allows to apply different unitest filters to class [ExprTreeSet](#), i.e. to the expression level data.frame data.

**Objects from the Class**

Objects can be created by calls of the form `new("UniFilter", ...)`. Alternatively, the constructor [UniFilter](#) can be used.

**Slots**

**foldchange:** Object of class "list" describing parameters for `fcFilter`.  
**prescall:** Object of class "list" describing parameters for `callFilter`.  
**unifilter:** Object of class "list" describing parameters for `unitestFilter`.  
**unitest:** Object of class "list" describing parameters for `uniTest`.  
**numfilters:** Object of class "numeric" giving the number of filters applied.

**Extends**

Class "[Filter](#)", directly.

**Methods**

**callFilter** signature(object = "UniFilter"): extracts slot prescall.  
**callFilter<-** signature(object = "UniFilter", value = "character"): replaces slot prescall with character vector c(cutoff, samples, condition).  
**fcFilter** signature(object = "UniFilter"): extracts slot foldchange.  
**fcFilter<-** signature(object = "UniFilter", value = "numeric"): replaces slot foldchange with numeric vector c(cutoff, direction).  
**uniTest** signature(object = "UniFilter"): extracts slot unitest.  
**uniTest<-** signature(object = "UniFilter", value = "character"): replaces slot unitest with character vector c(type, alternative, correction, numperm, mu, paired, conflevel, varequ).  
**unitestFilter** signature(object = "UniFilter"): extracts slot unifilter.  
**unitestFilter<-** signature(object = "UniFilter", value = "character"): replaces slot unifilter with character vector c(cutoff, variable).

**Author(s)**

Christian Stratowa

**See Also**

related classes [Filter](#), [PreFilter](#).

**Examples**

```
unifltr <- new("UniFilter", unitest=list("t.test"))
fcFilter(unifltr) <- c(1.5,"both")
unitestFilter(unifltr) <- c(0.01,"pval")
str(unifltr)
```

---

UniFilter-constructor *Constructor for Class UniFilter*

---

**Description**

Constructor for class UniFilter allows to apply different unitest filters to class [ExprTreeSet](#), i.e. to the expression level data.frame data.

**Usage**

```
UniFilter(unitest = "t.test",
          foldchange = character(),
          prescall = character(),
          unifilter = character())
```

**Arguments**

unitest	"character" vector describing parameters for <a href="#">uniTest</a> .
foldchange	"character" vector describing parameters for <a href="#">fcFilter</a> .
prescall	"character" vector describing parameters for <a href="#">callFilter</a> .
unifilter	"character" vector describing parameters for <a href="#">unitestFilter</a> .

**Details**

The UniFilter constructor allows to apply the following unitest filters to class [ExprTreeSet](#):

unitest:	character vector <code>c(type,alternative,correction.numperm,mu,paired,conflevel,varequ)</code> .
foldchange:	character vector <code>c(cutoff,direction)</code> .
prescall:	character vector <code>c(cutoff,samples,condition)</code> .
unifilter:	character vector <code>c(cutoff,variable)</code> .

**Value**

An object of type "[UniFilter](#)"

**Note**

Function `UniFilter` is used as constructor for class `UniFilter` so that the user need not know details for creating S4 classes.

**Author(s)**

Christian Stratowa

**See Also**

[UniFilter](#), [PreFilter](#)

**Examples**

```
## fill character vectors within constructor
unifltr <- UniFilter(unitest=c("t.test","two.sided","none",0,0.0,FALSE,0.95,TRUE),
                    foldchange=c(1.3,"both"),unifilter=c(0.1,"pval"))
str(unifltr)

## alternatively add character vectors as methods after creation of constructor
unifltr <- UniFilter()
fcFilter(unifltr) <- c(1.5,"both")
unitestFilter(unifltr) <- c(0.01,"pval")
str(unifltr)
```



uniTest-methods

*A Two-Group Unittest***Description**

Unittest performs a a two group uni-test such as the `t.test` on each row of the expression dataframe. The Unittest returns a dataframe containing the results of the test.

*Usage*

```
uniTest(object)
uniTest(object, value)<-
```

**Arguments**

`object` object of class `UniFilter`.  
`value` character vector `c(type, alternative, correction, numperm, mu, paired, conflevel, varequ)`

**Details**

The method `uniTest` initializes the following parameters:

`type`: a character string specifying the type of test: currently `"t.test"` (default) or `"normal.test"`.  
`alternative`: a character string specifying the alternative hypothesis, must be one of `"two.sided"` (default), `"greater"`, or `"less"`.  
`correction`: a correction to adjust p-values for multiple comparisons:  
`correction="none"`: no correction (default).  
`correction="bonferroni"`: Bonferroni correction.  
`correction="BH" or "fdr"`: correction for false discovery rate (Benjamini & Hochberg).  
`correction="BY"`: correction for false discovery rate (Benjamini & Yekutieli).  
`correction="hochberg"`: Hochberg correction.  
`correction="holm"`: Holm correction.  
`correction="wy"`: Westfall-Young step-down adjusted p-chance (E.Manduchi).  
`numperm`: optional number of permutations used to determine p-chance (default is 0).  
`mu`: a number indicating the true value of the difference in means for a two sample test (default is 0).  
`paired`: a logical indicating whether you want a paired uni-test (default is FALSE).  
`conflevel`: confidence level of the interval (default is 0.95).  
`varequ`: a logical variable indicating whether to treat the two variances as being equal. If TRUE then the pooled

**Value**

An initialized `UniFilter` object.

**Author(s)**

Christian Stratowa

**References**

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Dudoit S., Yang Y.H., Callow M.J., Speed T.P. (2000) Statistical methods for identifying differentially expressed genes in replicated cDNA microarray experiments. *Technical report 578*; UC Berkeley.

Manduchi E. (2000) Software: tpWY, see: <http://www.cbil.upenn.edu/tpWY/>

## Examples

```
unifltr <- UniFilter()
uniTest(unifltr) <- c("t.test", "two.sided", "none", 0, 0.0, FALSE, 0.98, TRUE)
str(unifltr)
```

---

unitestFilter-methods *Unitest Filter*

---

## Description

This method initializes the Unitest Filter.

Applying an unitest such as the `t.test` to two groups returns the p-value for the test and the value of the t-statistic. The Unitest Filter allows to select only rows satisfying e.g. a certain p-value as cutoff.

The Unitest Filter flags all rows with: `flag = (variable <= cutoff)`

### Usage

```
unitestFilter(object)
unitestFilter(object, value)<-
```

## Arguments

object	object of class UniFilter.
value	character vector <code>c(cutoff, variable)</code> .

## Details

The method `unitestFilter` initializes the following parameters:

cutoff:	the cutoff level for the filter.
variable:	<code>variable="pval"</code> (default): p-value. <code>variable="stat"</code> : univariate statistic. <code>variable="padj"</code> : optional adjusted p-value. <code>variable="pcha"</code> : optional p-value obtained by permutations.

## Value

An initialized `UniFilter` object.

**Author(s)**

Christian Stratowa

**Examples**

```
unifltr <- UniFilter()
unitestFilter(unifltr) <- c(0.01,"pval")
str(unifltr)
```

---

validCall-methods      *Get Valid Detection Call Values*

---

**Description**

Extracts valid present call values with unit names as row names.

*Usage*

```
validCall(object, which = "UnitName")
validPVal(object, which = "UnitName")
```

**Arguments**

object	object of class CallTreeSet.
which	name of column containing unit name.

**Details**

Method `validCall` returns the present calls from slot `detcall` as `data.frame` and uses column `which` as row names, usually the probeset IDs stored in column "UnitName".

Method `validPVal` returns the detection call p-values from slot `data` as `data.frame` and uses column `which` as row names, usually the probeset IDs stored in column "UnitName".

**Value**

A `data.frame`.

**Author(s)**

Christian Stratowa

**See Also**

[validData](#), [validExpr](#)

---

 validData-methods      *Extract Subset of Data*


---

**Description**

Extracts a subset of valid data from data.frame data.

*Usage*

```
validData(object, which = "", unitID = NULL, unittype = "transcript")
```

**Arguments**

object	object of class DataTreeSet, ExprTreeSet or CallTreeSet.
which	type of probes to be returned for DataTreeSet, otherwise name of column containing unit name.
unitID	optional vector of UNIT_IDs.
unittype	character vector, "transcript" or "probeset".

**Details**

For class DataTreeSet and expression arrays, validData returns all the perfect match or mismatch probes on the arrays the object represents as data.frame, i.e. which can have the following values:

pm:	perfect match probes.
mm:	mismatch probes.
both:	both perfect match and mismatch probes.

For class DataTreeSet and exon arrays, validData returns the probes of the different exon levels as data.frame, i.e. which can have one of the following values:

core:	probesets supported by RefSeq and full-length GenBank transcripts.
metacore:	core meta-probesets.
extended:	probesets with other cDNA support.
metaextended:	extended meta-probesets.
full:	probesets supported by gene predictions only.
metafull:	full meta-probesets.
affx:	standard AFFX controls.
all:	combination of above.
genomic:	genomic background probes.
antigenomic:	antigenomic background probes.

For class ExprTreeSet validData returns the valid expression levels from slot data with unit names as row names, usually the probeset IDs stored in column which="UnitName".

For class CallTreeSet validData returns the valid detection call p-values from slot data with unit names as row names, usually the probeset IDs stored in column which="UnitName".

**Value**

A [data.frame](#).

**Author(s)**

Christian Stratowa

**See Also**

[pm](#), [mm](#), [validExpr](#), [validCall](#)

---

validExpr-methods      *Get Valid Expression Levels*

---

**Description**

Extracts valid expression levels with unit names as row names from data.frame data.

*Usage*

```
validExpr(object, which = "UnitName")
```

**Arguments**

object	object of class ExprTreeSet.
which	name of column containing unit name.

**Details**

Method validExpr returns the expression levels from slot data and uses column which as row names, usually the probeset IDs stored in column "UnitName".

**Value**

A [data.frame](#).

**Author(s)**

Christian Stratowa

**See Also**

[validData](#), [validCall](#)

validSE-methods

*Get Valid Standard Errors*

---

**Description**

Extracts valid standard errors with unit names as row names.

*Usage*

```
validSE(object, which = "UnitName")
```

**Arguments**

object	object of class ExprTreeSet.
which	name of column containing unit name.

**Details**

Method validSE returns the standard errors (or standard deviations) from the expression trees and uses column which as row names, usually the probeset IDs stored in column "UnitName".

**Value**

A [data.frame](#).

**Author(s)**

Christian Stratowa

**See Also**

[validExpr](#)

---

validTreetype*Validate Tree Type*

---

**Description**

Validate tree type for corresponding data type.

**Usage**

```
validTreetype(treetype, datatype)
```

**Arguments**

treetype	tree type.
datatype	data type.

## Details

Every **ROOT** tree has an extension, which describes the type of data stored in this tree. For example, 'TestA1.cel' is the tree name that stores the CEL-file data for 'TestA1.CEL'.

Trees with datatype="scheme" have the following extensions:

scm: scheme tree containing (x,y)-coordinates and mask for UNIT\_ID.

idx: unit tree containing UnitName (i.e. probeset id), NumCells, NumAtoms, UnitType, for UNIT\_ID.

prb: probe tree containing probe sequences.

ann: transcript annotation tree.

anx: exon annotation tree; exon arrays only.

anp: probeset annotation tree; exon arrays only.

cxy: coordinate tree containing CLF-file information; exon arrays only.

exn: exon tree; exon arrays only.

pbs: probeset tree; exon arrays only.

Trees with datatype="rawdata" have the following extensions:

cel: data tree containing CEL-file data.

Trees with datatype="preprocess" have the following extensions:

int: intensity tree containing background-corrected intensities.

sbg: background tree containing MAS4 sector background levels.

wbg: background tree containing MAS5 weighted sector background levels.

rbg: background tree containing RMA background levels.

gbg: background tree containing GC-content background levels.

cmn: cell tree containing preprocessed intensities using algorithm 'mean'.

cmd: cell tree containing preprocessed intensities using algorithm 'median'.

c1w: cell tree containing preprocessed intensities using algorithm 'lowess'.

css: cell tree containing preprocessed intensities using algorithm 'supsmu'.

cqu: cell tree containing preprocessed intensities using algorithm 'quantile'.

dc5: detection tree containing MAS5 detection call and p-value.

dab: detection tree containing DABG detection call and p-value.

amn: expression tree containing expression levels computed with 'arithmetic mean'.

gmn: expression tree containing expression levels computed with 'geometric mean'.

wmn: expression tree containing expression levels computed with 'weighted mean'.

wdf: expression tree containing expression levels computed with 'weighted difference'.

adf: expression tree containing expression levels computed with 'average difference'.

tbw: expression tree containing expression levels computed with 'tukey biweight'.

mdp: expression tree containing expression levels computed with 'median polish'.

r1m: quality tree containing expression levels, NUSE, RLE computed with 'median polish'.

res: residual tree containing the residual SE and the model fit weights.

brd: border tree containing border intensities, mean border intensities and COI.

Trees with datatype="normation" have the following extensions:

tmn: expression tree after normalization using algorithm 'trimmed mean'.

med: expression tree after normalization using algorithm 'median'.

ksm: expression tree after normalization using algorithm 'kernel smoother'.

low: expression tree after normalization using algorithm 'lowess'.

sup: expression tree after normalization using algorithm 'supsmu'.

qua: expression tree after normalization using algorithm 'quantile'.

mdp: expression tree after normalization using algorithm 'median polish'.

## Value

Returns the valid treetype, otherwise an error message is returned.

**Note**

Not all tree types are used in the current package.

**Author(s)**

Christian Stratowa

**See Also**

[getDatatype](#), [type2Exten](#)

**Examples**

```
validTreetype("prb", "scheme")
validTreetype("cel", "rawdata")
validTreetype("tbw", "preprocess")
```

---

varFilter-methods      *Variance Filter*

---

**Description**

This method initializes the Variance Filter.

The Variance Filter flags all rows with:  $\text{flag} = (\text{var}/\text{mean} \geq \text{cutoff})$

*Usage*

```
varFilter(object)
varFilter(object, value)<-
```

**Arguments**

**object**            object of class `PreFilter`.  
**value**             numeric vector `c(cutoff, trim, epsilon)`.

**Details**

The method `varFilter` initializes the following parameters:

**cutoff:**    the cutoff level for the filter.  
**trim:**       the trim value for trimmed mean (default is `trim=0`).  
**epsilon:**    value to replace mean (default is `epsilon=0.01`):  
               `epsilon > 0`: replace `mean=0` with `epsilon`.  
               `epsilon = 0`: always set `mean=1`.

Note, that for `epsilon = 0` the filter flags all rows with:  $\text{variance} \geq \text{cutoff}$

**Value**

An initialized `PreFilter` object.



**Author(s)**

Christian Stratowa

**Examples**

```
prefltr <- PreFilter()
varFilter(prefltr) <- c(0.6,0.02,0.01)
str(prefltr)
```

---

volcanoplot-methods    *Volcano Plot*


---

**Description**

Produce a scatter plot of fold-change values vs p-values, called volcano plot.

*Usage*

```
volcanoplot(x, labels = "", p.value = "pval", mask = FALSE
```

**Arguments**

x	object of class <a href="#">AnalysisTreeSet</a> .
labels	optional transcript labels to be drawn at plotting points.
p.value	type of p-value, 'pval' for p-value, 'padj' for adjusted p-value, or 'pcha' for p-chance.
mask	logical, if TRUE draw only points for transcripts satisfying the univariate test.
show.cutoff	logical, if TRUE draw lines indicating cutoff.
cex.text	magnification to be used for optional labels.
col.text	color to be used for optional labels.
col.cutoff	color to be used for lines indicating cutoff, if show.cutoff=TRUE.
xlim	optional range for the plotted fold-change values.
xlab	label of x-axis.
ylab	label of y-axis.
pch	either an integer specifying a symbol or a single character to be used as the default in plotting points.
...	optional arguments to be passed to plot.

**Details**

Produces a volcano plot for slot data for an object of class [AnalysisTreeSet](#).

It is possible to label the points of the volcano plot, whereby the following labels parameters are valid:

fUnitName:	unit name (probeset ID).
fName:	gene name.
fSymbol:	gene symbol.
fChromosome:	chromosome.
fCytoBand:	cytoband.

**Author(s)**

Christian Stratowa

---

xpsOptions	<i>xps Options</i>
------------	--------------------

---

**Description**

Options for xps

**Usage**

xpsOptions(debug=FALSE)

**Arguments**

debug                    logical, if TRUE, print debug information.

**Details**

Currently only used to set debug to FALSE or TRUE.

**Value**

A global variable debug.xps can be set to TRUE.

**Author(s)**

Christian Stratowa

---

xpsQAReport	<i>Create Quality Assessment Report.</i>
-------------	--

---

**Description**

Create a quality assessment report.

**Usage**

```
xpsQAReport(xps.data,
            xps.expr   = NULL,
            xps.call   = NULL,
            xps.qual   = NULL,
            dataset    = character(0),
            title      = "Quality Report",
            date       = "October, 2011",
            author     = "Christian Stratowa",
            outdir     = file.path(getwd(), "QAReport"),
            add.pseudo = FALSE,
            overwrite  = FALSE,
            verbose    = TRUE,
            ...)
```

**Arguments**

xps.data	object of class <a href="#">DataTreeSet</a> .
xps.expr	object of class <a href="#">ExprTreeSet</a> .
xps.call	object of class <a href="#">CallTreeSet</a> .
xps.qual	object of class <a href="#">QualTreeSet</a> .
dataset	name of the dataset.
title	title of quality report.
date	date of quality report.
author	author(s) of quality report.
outdir	name of directory where to create the quality report.
add.pseudo	logical, if TRUE add pseudo-images to the quality report.
overwrite	logical, if TRUE overwrite outdir and its contents.
verbose	logical, if TRUE print status information.
...	optional arguments to be passed to xpsQARepor.

**Details**

Function `xpsQARepor` creates a quality assessment report "QARepor.pdf" for all `TreeSets`, which are passed as parameters to the function. It calls `library(tools)` and uses its function `buildVignettes` to create the report.

If parameter `xps.qual` is supplied, it is possible to create pseudo-images for every CEL-file by setting parameter `add.pseudo=TRUE`.

**Value**

None, the output is a pdf-file.

**Note**

Function `xpsQARepor` requires a working LaTeX implementation and so will only work on Windows platforms, and on OS X, if the user has installed the necessary LaTeX tools.

**Author(s)**

Christian Stratowa, based on ideas of package `affyQCReport`.

**Examples**

```
## Not run:
## first, load ROOT scheme file and ROOT data file
scheme.test3 <- root.scheme(paste(path.package("xps"), "schemes/SchemeTest3.root", sep="/"))
data.test3 <- root.data(scheme.test3, paste(path.package("xps"), "rootdata/DataTest3_cel.root", sep="/"))

## optional normalized expression levels
data.rma <- rma(data.test3, "Test3RMA", tmpdir="", background="pmonly", normalize=TRUE, verbose=FALSE)

## optional MAS5 detection call
call.mas5 <- mas5.call(data.test3, "Test3Call", tmpdir="", verbose=FALSE)

## optional quality measures
```

```
rlm.all <- rmaPLM(data.test3, "tmp_Test3RLMall", filedir=getwd(), tmpdir="", qualopt="all", option="transcr  
## quality assessment report  
xpsQAReport(data.test3, data.rma, call.mas5, rlm.all, dataset="My Dataset", add.pseudo=TRUE, overwrite=TRUE  
## End(Not run)
```

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