

# Introduction to RBM package

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## Contents

<b>1 Overview</b>	<b>1</b>
<b>2 Getting started</b>	<b>2</b>
<b>3 RBM_T and RBM_F functions</b>	<b>2</b>
<b>4 Ovarian cancer methylation example using the RBM_T function</b>	<b>6</b>

## 1 Overview

This document provides an introduction to the RBM package. The RBM package executes the resampling-based empirical Bayes approach using either permutation or bootstrap tests based on moderated t-statistics through the following steps.

- Firstly, the RBM package computes the moderated t-statistics based on the observed data set for each feature using the lmFit and eBayes function.
- Secondly, the original data are permuted or bootstrapped in a way that matches the null hypothesis to generate permuted or bootstrapped resamples, and the reference distribution is constructed using the resampled moderated t-statistics calculated from permutation or bootstrap resamples.
- Finally, the p-values from permutation or bootstrap tests are calculated based on the proportion of the permuted or bootstrapped moderated t-statistics that are as extreme as, or more extreme than, the observed moderated t-statistics.

Additional detailed information regarding resampling-based empirical Bayes approach can be found elsewhere (Li et al., 2013).

## 2 Getting started

The `RBM` package can be installed and loaded through the following R code.  
Install the `RBM` package with:

```
> if (!requireNamespace("BiocManager", quietly=TRUE))
+   install.packages("BiocManager")
> BiocManager::install("RBM")
```

Load the `RBM` package with:

```
> library(RBM)
```

## 3 RBM\_T and RBM\_F functions

There are two functions in the `RBM` package: `RBM_T` and `RBM_F`. Both functions require input data in the matrix format with rows denoting features and columns denoting samples. `RBM_T` is used for two-group comparisons such as study designs with a treatment group and a control group. `RBM_F` can be used for more complex study designs such as more than two groups or time-course studies. Both functions need a vector for group notation, i.e., "1" denotes the treatment group and "0" denotes the control group. For the `RBM_F` function, a contrast vector need to be provided by users to perform pairwise comparisons between groups. For example, if the design has three groups (0, 1, 2), the `aContrast` parameter will be a vector such as ("X1-X0", "X2-X1", "X2-X0") to denote all pairwise comparisons. Users just need to add an extra "X" before the group labels to do the contrasts.

- Examples using the `RBM_T` function: `normdata` simulates a standardized gene expression data and `unifdata` simulates a methylation microarray data. The *p*-values from the `RBM_T` function could be further adjusted using the `p.adjust` function in the `stats` package through the Benjamini-Hochberg method.

```
> library(RBM)
> normdata <- matrix(rnorm(1000*6, 0, 1), 1000, 6)
> mydesign <- c(0,0,0,1,1,1)
> myresult <- RBM_T(normdata, mydesign, 100, 0.05)
> summary(myresult)

      Length Class  Mode
ordfit_t     1000 -none- numeric
ordfit_pvalue 1000 -none- numeric
ordfit_beta0  1000 -none- numeric
ordfit_beta1  1000 -none- numeric
permutation_p 1000 -none- numeric
bootstrap_p    1000 -none- numeric

> sum(myresult$permutation_p<=0.05)
```

```

[1] 63

> which(myresult$permutation_p<=0.05)

[1] 18 24 40 65 96 122 141 143 163 164 171 189 207 240 270 283 313 331 347
[20] 348 351 356 367 391 403 415 445 447 473 484 528 540 556 562 568 577 587 588
[39] 599 632 653 660 673 688 708 716 725 729 752 761 830 836 865 871 891 895 913
[58] 929 934 948 970 990 992

> sum(myresult$bootstrap_p<=0.05)

[1] 6

> which(myresult$bootstrap_p<=0.05)

[1] 40 148 473 716 948 990

> permutation_adjp <- p.adjust(myresult$permutation_p, "BH")
> sum(permutation_adjp<=0.05)

[1] 6

> bootstrap_adjp <- p.adjust(myresult$bootstrap_p, "BH")
> sum(bootstrap_adjp<=0.05)

[1] 0

> unifdata <- matrix(runif(1000*7, 0.10, 0.95), 1000, 7)
> mydesign2 <- c(0,0,0, 1,1,1,1)
> myresult2 <- RBM_T(unifdata,mydesign2,100,0.05)
> sum(myresult2$permutation_p<=0.05)

[1] 0

> sum(myresult2$bootstrap_p<=0.05)

[1] 30

> which(myresult2$bootstrap_p<=0.05)

[1] 124 129 137 143 162 177 218 225 303 333 458 464 499 516 532
[16] 598 661 663 666 674 709 719 726 758 791 828 923 990 993 1000

> bootstrap2_adjp <- p.adjust(myresult2$bootstrap_p, "BH")
> sum(bootstrap2_adjp<=0.05)

[1] 0

```

- Examples using the RBM\_F function: normdata\_F simulates a standardized gene expression data and unifdata\_F simulates a methylation microarray data. In both examples, we were interested in pairwise comparisons.

```

> normdata_F <- matrix(rnorm(1000*9,0,2), 1000, 9)
> mydesign_F <- c(0, 0, 0, 1, 1, 1, 2, 2, 2)
> aContrast <- c("X1-X0", "X2-X1", "X2-X0")
> myresult_F <- RBM_F(normdata_F, mydesign_F, aContrast, 100, 0.05)
> summary(myresult_F)

      Length Class  Mode
ordfit_t     3000 -none- numeric
ordfit_pvalue 3000 -none- numeric
ordfit_beta1  3000 -none- numeric
permutation_p 3000 -none- numeric
bootstrap_p   3000 -none- numeric

> sum(myresult_F$permutation_p[, 1]<=0.05)
[1] 52

> sum(myresult_F$permutation_p[, 2]<=0.05)
[1] 58

> sum(myresult_F$permutation_p[, 3]<=0.05)
[1] 58

> which(myresult_F$permutation_p[, 1]<=0.05)
[1]  29  36  42  43  44  93 112 124 130 148 152 192 202 210 220 231 235 292 317
[20] 321 323 335 340 343 356 393 406 521 534 562 598 624 637 651 667 682 743 769
[39] 779 781 791 821 832 845 853 870 887 906 916 946 954 997

> which(myresult_F$permutation_p[, 2]<=0.05)
[1]  29  36  42  43  44  93 112 124 130 148 152 153 192 193 202 210 220 228 235
[20] 285 292 317 320 321 323 335 340 343 356 393 404 406 518 525 534 562 624 637
[39] 651 659 667 675 682 743 779 781 791 814 821 832 845 853 887 906 916 946 954
[58] 997

> which(myresult_F$permutation_p[, 3]<=0.05)
[1]  29  42  43  44  93 103 112 124 130 152 192 202 220 224 228 231 235 285 292
[20] 317 321 323 340 343 356 393 404 406 475 525 534 562 573 588 624 637 651 659
[39] 667 675 682 743 746 769 779 791 814 821 832 845 853 887 906 916 946 952 954
[58] 997

```

```

> con1_adjp <- p.adjust(myresult_F$permutation_p[, 1], "BH")
> sum(con1_adjp<=0.05/3)

[1] 2

> con2_adjp <- p.adjust(myresult_F$permutation_p[, 2], "BH")
> sum(con2_adjp<=0.05/3)

[1] 7

> con3_adjp <- p.adjust(myresult_F$permutation_p[, 3], "BH")
> sum(con3_adjp<=0.05/3)

[1] 12

> which(con2_adjp<=0.05/3)

[1] 43 93 343 393 624 779 853

> which(con3_adjp<=0.05/3)

[1] 42 124 220 292 321 343 624 667 779 853 916 946

> unifdata_F <- matrix(runif(1000*18, 0.15, 0.98), 1000, 18)
> mydesign2_F <- c(rep(0, 6), rep(1, 6), rep(2, 6))
> aContrast <- c("X1-X0", "X2-X1", "X2-X0")
> myresult2_F <- RBM_F(unifdata_F, mydesign2_F, aContrast, 100, 0.05)
> summary(myresult2_F)

      Length Class  Mode
ordfit_t     3000 -none- numeric
ordfit_pvalue 3000 -none- numeric
ordfit_beta1 3000 -none- numeric
permutation_p 3000 -none- numeric
bootstrap_p   3000 -none- numeric

> sum(myresult2_F$bootstrap_p[, 1]<=0.05)

[1] 47

> sum(myresult2_F$bootstrap_p[, 2]<=0.05)

[1] 44

> sum(myresult2_F$bootstrap_p[, 3]<=0.05)

[1] 45

```

```

> which(myresult2_F$bootstrap_p[, 1]<=0.05)

[1] 1 4 13 37 45 52 65 87 120 131 184 213 229 241 303 308 311 313 332
[20] 335 347 349 400 417 447 487 489 520 541 562 572 586 625 634 670 690 712 755
[39] 828 832 860 898 902 936 941 954 957

> which(myresult2_F$bootstrap_p[, 2]<=0.05)

[1] 4 13 23 37 45 52 87 120 131 165 213 241 303 308 313 332 335 349 400
[20] 417 447 487 489 520 527 541 562 572 586 666 670 690 737 755 828 832 875 898
[39] 902 916 936 941 954 957

> which(myresult2_F$bootstrap_p[, 3]<=0.05)

[1] 1 4 13 23 37 52 65 83 87 130 184 213 241 292 303 308 313 335 349
[20] 400 417 447 487 489 528 541 572 573 586 666 670 690 712 737 755 824 828 832
[39] 860 875 898 936 941 954 957

> con21_adjp <- p.adjust(myresult2_F$bootstrap_p[, 1], "BH")
> sum(con21_adjp<=0.05/3)

[1] 4

> con22_adjp <- p.adjust(myresult2_F$bootstrap_p[, 2], "BH")
> sum(con22_adjp<=0.05/3)

[1] 10

> con23_adjp <- p.adjust(myresult2_F$bootstrap_p[, 3], "BH")
> sum(con23_adjp<=0.05/3)

[1] 8

```

## 4 Ovarian cancer methylation example using the RBM\_T function

Two-group comparisons are the most common contrast in biological and biomedical field. The ovarian cancer methylation example is used to illustrate the application of `RBM_T` in identifying differentially methylated loci. The ovarian cancer methylation example is taken from the genome-wide DNA methylation profiling of United Kingdom Ovarian Cancer Population Study (UKOPS). This study used Illumina Infinium 27k Human DNA methylation Beadchip v1.2 to obtain DNA methylation profiles on over 27,000 CpGs in whole blood cells from 266 ovarian cancer women and 274 age-matched healthy controls. The data are downloaded from the NCBI GEO website with access number GSE19711. For illustration purpose, we chose the first 1000 loci in 8 randomly selected women with 4 ovarian cancer cases (pre-treatment) and 4 healthy controls. The following codes show the process of generating significant differential DNA methylation loci using the `RBM_T` function and presenting the results for further validation and investigations.

```

> system.file("data", package = "RBM")
[1] "/private/tmp/RtmpQBoTFp/Rinst64337611defb/RBM/data"

> data(ovarian_cancer_methylation)
> summary(ovarian_cancer_methylation)

    IlmnID      Beta      exmdata2[, 2]      exmdata3[, 2]
cg00000292: 1   Min.   :0.01058   Min.   :0.01187   Min.   :0.009103
cg00002426: 1   1st Qu.:0.04111   1st Qu.:0.04407   1st Qu.:0.041543
cg00003994: 1   Median :0.08284   Median :0.09531   Median :0.087042
cg00005847: 1   Mean    :0.27397   Mean    :0.28872   Mean    :0.283729
cg00006414: 1   3rd Qu.:0.52135   3rd Qu.:0.59032   3rd Qu.:0.558575
cg00007981: 1   Max.    :0.97069   Max.    :0.96937   Max.    :0.970155
(Other)     :994          NA's    :4
exmdata4[, 2]  exmdata5[, 2]  exmdata6[, 2]  exmdata7[, 2]
Min.   :0.01019   Min.   :0.01108   Min.   :0.01937   Min.   :0.01278
1st Qu.:0.04092   1st Qu.:0.04059   1st Qu.:0.05060   1st Qu.:0.04260
Median :0.09042   Median :0.08527   Median :0.09502   Median :0.09362
Mean   :0.28508   Mean   :0.28482   Mean   :0.27348   Mean   :0.27563
3rd Qu.:0.57502   3rd Qu.:0.57300   3rd Qu.:0.52099   3rd Qu.:0.52240
Max.   :0.96658   Max.   :0.97516   Max.   :0.96681   Max.   :0.95974
NA's   :1

exmdata8[, 2]
Min.   :0.01357
1st Qu.:0.04387
Median :0.09282
Mean   :0.28679
3rd Qu.:0.57217
Max.   :0.96268

> ovarian_cancer_data <- ovarian_cancer_methylation[, -1]
> label <- c(1, 1, 0, 0, 1, 1, 0, 0)
> diff_results <- RBM_T(aData=ovarian_cancer_data, vec_trt=label, repetition=100, alpha=0.05)
> summary(diff_results)

      Length Class  Mode
ordfit_t     1000  -none- numeric
ordfit_pvalue 1000  -none- numeric
ordfit_beta0  1000  -none- numeric
ordfit_beta1  1000  -none- numeric
permutation_p 1000  -none- numeric
bootstrap_p   1000  -none- numeric

> sum(diff_results$ordfit_pvalue<=0.05)
[1] 45

```

```

> sum(diff_results$permutation_p<=0.05)
[1] 69

> sum(diff_results$bootstrap_p<=0.05)
[1] 53

> ordfit_adjp <- p.adjust(diff_results$ordfit_pvalue, "BH")
> sum(ordfit_adjp<=0.05)

[1] 0

> perm_adjp <- p.adjust(diff_results$permutation_p, "BH")
> sum(perm_adjp<=0.05)

[1] 4

> boot_adjp <- p.adjust(diff_results$bootstrap_p, "BH")
> sum(boot_adjp<=0.05)

[1] 7

> diff_list_perm <- which(perm_adjp<=0.05)
> diff_list_boot <- which(boot_adjp<=0.05)
> sig_results_perm <- cbind(ovarian_cancer_methylation[, diff_results$ordfit_t[diff_list_perm]], diff_results$permutation_p[diff_list_perm])
> print(sig_results_perm)

      IlmnID      Beta exmdata2[, 2] exmdata3[, 2] exmdata4[, 2]
19  cg00016968  0.80628480          NA   0.81440820   0.83623180
83  cg00072216  0.04505377   0.04598964   0.04000674   0.03231534
237 cg00215066  0.94926640   0.95311870   0.94634910   0.94561120
245 cg00224508  0.04479948   0.04972043   0.04152814   0.04189373
               exmdata5[, 2] exmdata6[, 2] exmdata7[, 2] exmdata8[, 2]
19        0.80831380    0.73306440    0.82968340   0.84917800
83        0.04965089    0.04833366    0.03466159   0.04390894
237       0.94837410    0.94665570    0.94089070   0.94600090
245       0.04208405    0.05284988    0.03775905   0.03955271
      diff_results$ordfit_t[diff_list_perm]
19                           -2.446404
83                           2.514109
237                          1.419654
245                          1.962457
      diff_results$permutation_p[diff_list_perm]
19                           0
83                           0
237                          0
245                          0

```

```

> sig_results_boot <- cbind(ovarian_cancer_methylation[, diff_list_boot], diff_results$ordfit_t[diff_list_boot])
> print(sig_results_boot)

    IlmnID      Beta exmdata2[, 2] exmdata3[, 2] exmdata4[, 2]
131 cg00121904 0.15449580     0.17949750     0.23608110     0.24354150
146 cg00134539 0.61101320     0.53321780     0.45999340     0.46787420
259 cg00234961 0.04192170     0.04321576     0.05707140     0.05327565
280 cg00260778 0.64319890     0.60488960     0.56735060     0.53150910
833 cg00814580 0.09348613     0.09619816     0.12010440     0.11534240
882 cg00858899 0.11427700     0.11919540     0.07690343     0.08321229
979 cg00945507 0.13432250     0.23854600     0.34749760     0.28903340
               exmdata5[, 2] exmdata6[, 2] exmdata7[, 2] exmdata8[, 2]
131     0.17352980     0.12564280     0.18193170     0.20847670
146     0.67191510     0.63137380     0.47929610     0.45428300
259     0.04030003     0.03996053     0.05086962     0.05445672
280     0.61920530     0.61925200     0.46753250     0.55632410
833     0.09577040     0.11598850     0.12860890     0.14111200
882     0.08961409     0.10730660     0.09203980     0.08726349
979     0.11848510     0.16653850     0.30718420     0.26624740
               diff_results$ordfit_t[diff_list_boot]
131                  -3.451679
146                  5.394750
259                 -4.052697
280                  4.170347
833                 -3.428319
882                  3.179415
979                 -4.750997
               diff_results$bootstrap_p[diff_list_boot]
131                      0
146                      0
259                      0
280                      0
833                      0
882                      0
979                      0

```