## Package 'CNAnorm'

April 12, 2022

**Version** 1.40.0 **Date** 2012-08-06

**Title** A normalization method for Copy Number Aberration in cancer samples

Author Stefano Berri <sberri@illumina.com>, Henry M. Wood

<H.M.Wood@leeds.ac.uk>, Arief Gusnanto <a.gusnanto@leeds.ac.uk>

Maintainer Stefano Berri <sberri@illumina.com>

**Depends** R (>= 2.10.1), methods

Description Performs ratio, GC content correction and normalization of data obtained using low coverage (one read every 100-10,000 bp) high troughput sequencing. It performs a ``discrete" normalization looking for the ploidy of the genome. It will also provide tumour content if at least two ploidy states can be found.

License GPL-2

Imports DNAcopy

Collate AllGenerics.R AllClasses.R dataFrame2object.R workflowWrapper.R initialize-methods.R summary-methods.R smoothseg.R bandsegment.R mixtureModel.R normalize.R length-methods.R CNAnorm-accessors.R CNAnorm-methods.R Data-methods.R makeDefaultGraphParamteres.R

URL http://www.r-project.org,

LazyLoad yes

**biocViews** CopyNumberVariation, Sequencing, Coverage, Normalization, WholeGenome, DNASeq, GenomicVariation

git\_url https://git.bioconductor.org/packages/CNAnorm

git\_branch RELEASE\_3\_14

git\_last\_commit fe146be

git\_last\_commit\_date 2021-10-26

**Date/Publication** 2022-04-12

2 addDNACopy

## **R** topics documented:

addD	NACopy Methods for Function addDNACopy in Package 'CNAnorm'	
Index		26
	vandation	ر کے
	validation	
	retrieve peaks and ploidy	
	ratio	
	plotPeaks	
	plotGenome	
	peakPloidy	18
	Params-class	17
	LS041	17
	InData-class	16
	hg19_hs_ideogr	15
	gPar	15
	gcNorm	14
	exportTable	13
	discreteNorm	12
	DerivData-class	
	dataFrame2object	10
	CNAnormWorkflow	8
	CNAnorm-class	6
	CN	6
	chromosomesPosition	5
	addSmooth	4
	addDNACopy	2

## Description

addSmooth segment ratio values in Package 'CNAnorm' using DNACopy

## Usage

```
## S4 method for signature 'CNAnorm'
addDNACopy(object, independent.arms = FALSE, ideograms = NULL,
    DNAcopy.smooth = list(), DNAcopy.weight = character(), DNAcopy.segment = list())
```

## Arguments

object An object of Class "CNAnorm" independent.arms

Boolean. If TRUE chromosomes arms will be treated as independent, ideograms

must be provided

ideograms A data frame containing information about banding. See ?hg19\_hs\_ideogr for

more information

addDNACopy 3

DNAcopy.smooth A list of parameters to be passed to function 'smooth.CNA' in package DNAcopy

DNAcopy.weight A character value of one of these values. 'poisson', 'gaussian', 'twoTailQuantile', 'oneTailQuantile'. It specifies a way to give weight to each window de-

pending on how much coverage in the normal deviates from the median for that chromosome. Options are listed in decreasing order of stringency. See Details

DNAcopy.segment

A list of parameters to be passed to function 'segment' in package DNAcopy. Parameters 'weights' and 'verbose' are not accepted

#### **Details**

'poisson': windows with coverage more or less than 2\*sqrt(mean) from the mean are weighted down. The most stringent. Recommended for unbiased genome wide sequencing.

'gaussian': windows with coverage more or less than 2\*sd from the median are weighted down. Recommended for genome wide sequencing where coverage in the normal is far from poisson distribution.

'twoTailQuantile': windows with coverage outside 5-95th quantile are weighted down. Recommended when coverage is far from a normal distribution - such as capture experiments -

'oneTailQuantile': windows with coverage lower than 5th quantile are weighted down. Recommended when coverage is far from a normal distribution - such as capture experiments. Does not weight down windows with very high coverage.

#### Value

An object of class "CNAnorm"

#### Methods

signature(object = "CNAnorm") Segment ratio values on an object of class "CNAnorm". Returns the same object with extra slots (segMean, segID)

#### Author(s)

Stefano Berri <s.berri@leeds.ac.uk> and Arief Gusnanto <a.gusnanto@leeds.ac.uk>

#### References

Venkatraman, E. S. and Olshen, A. B. (2007) A faster circular binary segmentation algorithm for the analysis of array CGH data. Bioinformatics

#### See Also

segMean, CNAnorm-class, DNAcopy, hg19\_hs\_ideogr

4 addSmooth

#### **Examples**

```
data(LS041)
CN <- dataFrame2object(LS041)
CN <- addDNACopy(CN)</pre>
```

addSmooth

Methods for Function addSmooth in Package 'CNAnorm'

## **Description**

addSmooth segment and smooth perform ratio values in Package 'CNAnorm'

## Usage

```
## S4 method for signature 'CNAnorm'
addSmooth(object, lambda = 7, ...)
```

## Arguments

object An object of Class "CNAnorm"

lambda Degree of smoothness. See reference for more details... Further arguments to pass to the function smoothseg

#### Value

An object of class "CNAnorm"

#### Methods

signature(object = "CNAnorm") Segment and smooth perform ratio values on an object of class "CNAnorm". Returns the same object with extra slot (ratio.s)

#### Author(s)

Stefano Berri <s.berri@leeds.ac.uk> and Arief Gusnanto <a.gusnanto@leeds.ac.uk>

#### References

Huang, J., Gusnanto, A., O'Sullivan, K., Staaf, J., Borg, A. and Pawitan, Y. (2007) *Robust smooth segmentation approach for array CGH data analysis*. Bioinformatics

#### See Also

```
ratio.s, CNAnorm-class
```

chromosomesPosition 5

#### **Examples**

```
data(LS041)
CN <- dataFrame2object(LS041)
CN.gcNorm <- gcNorm(CN, exclude = c("chrX", "chrY", "chrM"))
CN.smooth <- addSmooth(CN)</pre>
```

chromosomesPosition

Accessors methods for Function ratio in Package 'CNAnorm'

## **Description**

chrs returns/set the name of chromosomes/contigs

arms retruns the name of the chromosome and arm. A data frame containing ideogram information has to be provided. See ?hg19\_hs\_ideogr for an example

pos returns/set the position of starting window. **Be careful!** If you need to change data, it is better to change the input data and start over.

## Usage

```
chrs(object)
## S4 method for signature 'CNAnorm'
pos(object, show = "start")
## S4 method for signature 'CNAnorm'
arms(object, banding_df)
```

## **Arguments**

object An object of Class "CNAnorm" show The position to show: 'start', 'end'

banding\_df A data frame with infromation about ideogram

## Value

chrs and arms return a character vector, pos returns a numeric vector

#### Author(s)

Stefano Berri <s.berri@leeds.ac.uk>

#### See Also

```
gcNorm, CNAnorm-class, hg19_hs_ideogr
```

6 CNAnorm-class

#### **Examples**

```
data(LS041)
data(hg19_hs_ideogr)
CN <- dataFrame2object(LS041)
dataFrameNames <- as.character(LS041$Chr)
objectNames <- chrs(CN)
armNames <- arms(CN, hg19_hs_ideogr)
# check the names are, indeed, the same
all(dataFrameNames == objectNames)
# make shorter names, drop the first three letters ('chr')
shortNames <- substr(chrs(CN),4,nchar(chrs(CN)))
chrs(CN) <- shortNames

# retrieve all new names
unique(chrs(CN))
unique(armNames)</pre>
```

CN

A CNAnorm object with information about most abundant ploidy states, obtained from data LS041.

## **Description**

This data is to provide an object to use in several examples without having to wait for computing it. To see how it was generated, see documentation of function peakPloidy.

## Usage

data(CN)

#### **Format**

A CNAnorm object

CNAnorm-class

Class "CNAnorm"

#### **Description**

Class to Contain and Describe copy number aberration (CNA) data from low coverage (approx 0.01 - 0.5X) Next Generation Sequencing

## **Objects from the Class**

Objects can be created by calls of the form new("CNAnorm", InData).

CNAnorm-class 7

#### Slots

InData: Object of class "InData". Contains input data provided by the user. All slots have same length. Each element describe one window. See Class "InData"

DerivData: Object of class "DerivData". Contains data derived from "InData". It can be Retrieved by the user, but methods should be used to populate "DerivData". All slots have same length as input data. See Class DerivData

Res: Object of class "Res". Contains slots with obtained results. See Class "Res"

Params: Object of class "Params". Contains crucial parameters passed to some of the methods for reusing in later steps or for documentation.

#### Methods

Summary of methods for class "CNAnorm". Type "methods? methodName" for more details about methodName.

ratio signature(object = "CNAnorm"): Retrieve ratio (Test/Control). If gcNorm was called, ratio is GC normalized

ratio.n signature(object = "CNAnorm"): Retrieve normalized ratio (not smoothed)
ratio.s signature(object = "CNAnorm"): Retrieve smoothed ratio
ratio.n.s signature(object = "CNAnorm"): Retrieve normalized smoothed ratio

segMean signature(object = "CNAnorm"): Retrieve segmented ratio (as provided by DNAcopy)
segMean.n signature(object = "CNAnorm"): Retrieve normalized segmented ratio

#### Author(s)

Stefano Berri <s.berri@leeds.ac.uk> and Arief Gusnanto <a.gusnanto@leeds.ac.uk>

#### References

CNA-norm: Discrete Normalization of Copy Number Alteration data from clinical samples (in preparation)

#### See Also

InData, DerivData for documentation on the slots.

8 CNAnormWorkflow

#### **Examples**

```
data(LS041)
CNA <- new("CNAnorm", InData = new("InData", Chr = as.character(LS041$Chr), Pos = LS041$Pos,
    Test = LS041$Test, Norm = LS041$Norm, GC = LS041$GC))</pre>
```

CNAnormWorkflow

Wrapper to "CNAnorm" workflow

## **Description**

This function is a wrapper to use for a fully automated CNAnorm workflow where interactivity is not required. It contains MOST possible parameters. Defaults are set to to run a standard and conservative workflow.

#### Usage

```
CNAnormWorkflow(dataFrame, gc.do=FALSE, gc.exclude=character(0),
    gc.maxNumPoints=10000, smooth.do=TRUE, smooth.lambda=7,
    smooth.other=list(), peak.method='closest', peak.exclude=character(0),
    peak.ploidyToTest=12, peak.sd=5, peak.dThresh=0.01, peak.n=2048,
    peak.adjust=.9, peak.force.smooth=TRUE, peak.reg=FALSE, peak.ds=1.5,
    peak.zero.count=FALSE, peak.other=list(), DNAcopy.do=TRUE,
    DNAcopy.independent.arms=FALSE, DNAcopy.ideograms=NULL,
    DNAcopy.smooth=list(), DNAcopy.segment=list(), DNAcopy.weight=character(),
    dNorm.normBy=NULL)
```

#### **Arguments**

All arguments are explained in the relative functions

A data frame with columns Chr, Pos, Test, Norm and optional GC. See dataFrame2object

gatdbrame Specify if GC correction need to be done. See gcNorm

gc.exclude See gcNorm

gc.maxNumPoints

See gcNorm

smooth.do Specify if smoothing need to be done. See addSmooth

smooth.lambda See addSmooth

smooth.other A list of other parameters to pass to the smoothing function. See addSmooth

peak.method See peakPloidy
peak.exclude See peakPloidy

peak.ploidyToTest

See peakPloidy

peak.sd See peakPloidy

CNAnormWorkflow 9

peak.dThresh See peakPloidy
peak.n See peakPloidy
peak.adjust See peakPloidy
peak.force.smooth

See peakPloidy

peak.reg See peakPloidy peak.ds See peakPloidy

peak.zero.count

See peakPloidy

peak.other A list of other parameters to be passed to funtions for peak detection See peak-

Ploidy

DNAcopy .do Specify if segmentation with DNAcopy need to be done. See addDNACopy

DNAcopy.independent.arms

See addDNACopy

DNAcopy.ideograms

See addDNACopy

DNAcopy.smooth See addDNACopy

DNAcopy.segment

See addDNACopy

DNAcopy.weight See addDNACopy
dNorm.normBy See discreteNorm

#### Value

An object of Class "CNAnorm"

## Author(s)

Stefano Berri <s.berri@leeds.ac.uk>

#### See Also

dataFrame2object, gcNorm, addSmooth, peakPloidy, addDNACopy, discreteNorm

```
data(LS041)
CN <- CNAnormWorkflow(LS041)</pre>
```

10 DerivData-class

dataFrame2object

Convert a data frame into an object of Class "CNAnorm"

## Description

Convert a data frame with column: Chr, Pos, Test, Norm and optional GC into object of class "CNAnorm"

## Usage

```
dataFrame2object(dataFrame)
```

## **Arguments**

dataFrame

A data frame with columns Chr, Pos, Test, Norm and optional GC

#### Value

An object of Class "CNAnorm"

#### Author(s)

Stefano Berri <s.berri@leeds.ac.uk>

#### See Also

```
CNAnorm-class, InData-class, data.frame
```

## **Examples**

```
data(LS041)
CN <- dataFrame2object(LS041)</pre>
```

DerivData-class

Class "DerivData"

## **Description**

A Class containing data derived from InData used for further computation and plotting.

## **Objects from the Class**

Objects can be created by calls of the form new("DerivData"), however DerivData is typically populated using methods. If a slot has not been populated yet, it has zero length, otherwise slots have the same length as InData.

DerivData-class 11

#### **Slots**

```
ratio: Numeric vector with ratio Test/Normal. Optionally GC corrected.
```

ratio.s: Numeric vector with smoothed ratio.

ratio.n: Numeric vector with normalized ratio.

ratio.s.n: Numeric vector with normalized and smoothed ratio.

segID: Numeric vecotr with ID of segmented data (as provided by DNACopy). Each segment has a different ID.

segMean: Numeric vector with mean value of the segment (as provided by DNACopy.)

segMean.n: Numeric vector with normalized segMean.

ok4density: Logical vector. Specify wich values have been used to calculate density.

#### Methods

length signature(x = "DerivData"): Returns number of windows.

#### Author(s)

Stefano Berri and Arief Gusnanto

#### References

Gusnanto, A., Wood, H.M., Pawitan, Y., Rabbitts, P. and Berri, S. (2011) *Correcting for cancer genome size and tumor cell content enables better estimation of copy number alterations from next generation sequence data*. Bioinformatics

## See Also

```
CNAnorm, InData-class
```

```
data(LS041)
inObject <- new("InData", Chr = as.character(LS041$Chr),
    Pos = LS041$Pos, Test = LS041$Test, Norm = LS041$Norm,
    GC = LS041$GC)
CNA <- new("CNAnorm", InData = inObject)</pre>
```

12 discreteNorm

discreteNorm

Methods for Function addSmooth in Package 'CNAnorm'

#### **Description**

discreteNorm performs normalization of data using information on ploidy. Implicitly calls "validation" if no validation was performed

#### Usage

```
## S4 method for signature 'CNAnorm'
discreteNorm(object, normBy = object)
```

#### **Arguments**

object An object of Class "CNAnorm" to normalize

normBy An object of Class "CNAnorm" used to set normalization. It is possible, for

instance, to normalize a sample at high resolution, using information obtained

from the same sample at low resolution

#### Value

An object of class "CNAnorm"

#### Author(s)

Stefano Berri <s.berri@leeds.ac.uk> and Arief Gusnanto <a.gusnanto@leeds.ac.uk>

#### References

Gusnanto, A., Wood, H.M., Pawitan, Y., Rabbitts, P. and Berri, S. (2011) Correcting for cancer genome size and tumour cell content enables better estimation of copy number alterations from next generation sequence data. Bioinformatics

## See Also

```
validation, peakPloidy
```

```
\mbox{data(CN)} # see peakPloidy documentation to know how object CN is created CN <- discreteNorm(CN)
```

exportTable 13

exportTable Methods for Functi	on exportTable in Package 'CNAnorm'
--------------------------------	-------------------------------------

#### **Description**

exportTable write a table with normalised values of each window. A wrapper to "write.table"

#### Usage

```
## S4 method for signature 'CNAnorm'
exportTable(object, file = "CNAnorm_table.tab", show = 'ratio',
    sep = "\t", row.names = FALSE, ...)
```

## **Arguments**

object an object of Class "CNAnorm"

file name of the file to save to

show what should be reported in the table: "ratio": the normalized ratio (a value of 1 means diploid). "ploidy": the same as ratio \* 2. "center": report ratio centered on the most abbundant copy. Ratio of 1 means that the most abbundant "state" is centered to 1

sep the field separator string.

row.names either a logical value indicating whether the row number should be written or a

character vector of row names to be written.

... Extra arguments to be passed to "write.table"

#### **Details**

It produces a tab delimited text file with the following columns:

Chr: Chromosome/contig name.

Pos: Starting position of the window.

Ratio: Ratio Test/Normal for each window after GC correction.

Ratio.n: Ratio Test/Normal or ploidy for each window after normalisation.

Ratio.s.n: Smoothed and normalised ratio Test/Normal or ploidy for each window.

SegMean: Mean of the segment this window belongs to.

SegMean.n: Normalised mean ratio Test/Normal or ploidy of the segment this window belongs to.

#### Value

An object of class "CNAnorm"

#### Author(s)

Stefano Berri <s.berri@leeds.ac.uk>

14 gcNorm

#### See Also

```
write.table
```

#### **Examples**

```
data(CN)
CN <- validation(CN)
CN <- discreteNorm(CN)
exportTable(CN, file = "CNAnorm_table.tab", show = 'ploidy')</pre>
```

gcNorm

Methods for Function gcNorm in Package 'CNAnorm'

## Description

gcNorm perform GC content normalization on ratio Test/Normal in Package 'CNAnorm'

#### Usage

```
## S4 method for signature 'CNAnorm'
gcNorm(object, exclude = character(0), maxNumPoints = 10000)
```

## **Arguments**

object An object of Class "CNAnorm"

exclude A character vector with name of chromosomes/contigues not to use to calculate

GC content correction. All genome, however, will be corrected

maxNumPoints Maximum number of data points to fit the loess correction. For computational

pourposes, if the number of points in ratio(object) is greater than maxNumPoints,

only maxNumPoints randomly selected will be used

#### Value

An object of class "CNAnorm"

#### Methods

signature(object = "CNAnorm") Perform GC content correction on an object of class "CNAnorm". Returns the same object with corrected ratio

#### Author(s)

Stefano Berri <s.berri@leeds.ac.uk>

#### See Also

loess, CNAnorm-class, ratio

gPar 15

#### **Examples**

```
data(LS041)
CN <- dataFrame2object(LS041)
# correct for GC content, but ignoring data from sex chromosomes and
# mitocondria
CN.gcNorm <- gcNorm(CN, exclude = c("chrX", "chrY", "chrM"))</pre>
```

gPar

An object with the default graphical parameters

## **Description**

This data object is used by some plotting methods and contains the default values. User can change graphical parameters by changing this object

The object consists of several layers refering to different plots and different properties. Here an indicative description

gPar\$genome: prameters here refer to the plot produced by plotGenome

graphical parameters: see ?par

\$colors: specify colors \$cex: specify relative size - points, text... \$lwd: specify line width \$lty: specify line type - solid, dashed \$mar: specify margins

#### Usage

data(gPar)

#### Format

A S3 object

hg19\_hs\_ideogr

An object with the ideogram information for homo sapiens - hg19

## **Description**

This is bundles data that can be provided to plotGenome in order to plot location of the centromere. In future release it might be used to produce an ideogram too

## Usage

```
data(hg19_hs_ideogr)
```

#### **Format**

A data.frame

16 InData-class

InData-class

Class "InData" ~~~

#### **Description**

A Class containing input data for CNA

#### **Objects from the Class**

Objects can be created by calls of the form new("InData", Chr, Pos, Test, Norm, ratio, GC).

#### **Slots**

Chr: Object of class "character". Name of the Chromosomes/Contigs of each window.

Pos: Object of class "numeric". Starting position of the each window.

Test: Object of class "numeric". Number of reads from Test in each window.

Norm: Object of class "numeric". Number of reads from Normal (Control) in each window.

ratio: Object of class "numeric". Ratio Test/Control in each window. Automatically computed if Test and Norm are provided, or user generated if Test and Norm are not know.

GC: Object of class "numeric". GC content of each window.

## Methods

**length** signature(x = "InData"): Returns number of windows from input data.

## Author(s)

Stefano Berri

#### References

Gusnanto, A., Wood, H.M., Pawitan, Y., Rabbitts, P. and Berri, S. (2011) *Correcting for cancer genome size and tumor cell content enables better estimation of copy number alterations from next generation sequence data*. Bioinformatics

#### See Also

**CNAnorm** 

```
data(LS041)
inObject <- new("InData", Chr = as.character(LS041$Chr), Pos = LS041$Pos,
    Test = LS041$Test, Norm = LS041$Norm, GC = LS041$GC)
CNA <- new("CNAnorm", InData = inObject)</pre>
```

LS041 17

LS041

Mapped reads in tumor and matched blood for patient LS041

#### Description

This data set provide reads in tumor and matched blood for patient LS041. Each row has information about non-overlapping window across the genome. In particular it reports: chromosome name, starting position of the window (1 based), number of mapped reads in the test (lung tumor), number of reads in the control (matched blood) and GC content of the window.

#### Usage

data(LS041)

#### **Format**

A dataframe

#### References

Gusnanto, A., Wood, H.M., Pawitan, Y., Rabbitts, P. and Berri, S. (2011) Correcting for cancer genome size and tumour cell content enables better estimation of copy number alterations from next generation sequence data. Bioinformatics

Params-class

Class "Params"

## Description

A Class containing some Parameters used in the analysis. Not heavily used at the moment.

## **Objects from the Class**

Objects can be created by calls of the form new("Params"), it is usually iniziated and populated with methods and functions of class CNAnorm.

#### Slots

method: variable of class "character". Record if the peakPloidy function was called using density or mixture.

density.n: The variable "n" used when calling peakPloidy. This variable is saved so that can be used later for drawing plots.

density.adjust: The variable "adjust" used when calling peakPloidy. This variable is saved so that can be used later for drawing plots

18 peakPloidy

- gc.excludeFromGCNorm: Vector of class "character". Name of the Chromosomes/Contigs not used for GC content correction.
- gc.maxNumPoints: One element vector of class "numeric". Specify how many points to use for GC correction
- gp.excludeFromDensity: Vector of class "character". Name of the Chromosomes/Contigs not used for peak guessing

#### Methods

```
length signature(x = "Params")
```

#### Author(s)

Stefano Berri

#### References

Gusnanto, A., Wood, H.M., Pawitan, Y., Rabbitts, P. and Berri, S. (2011) Correcting for cancer genome size and tumor cell content enables better estimation of copy number alterations from next generation sequence data. Bioinformatics

#### See Also

**CNAnorm** 

#### **Examples**

```
data(LS041)
inObject <- new("InData", Chr = as.character(LS041$Chr), Pos = LS041$Pos,
   Test = LS041$Test, Norm = LS041$Norm, GC = LS041$GC)
CNA <- new("CNAnorm", InData = inObject)</pre>
```

peakPloidy

Methods for Function peakPloidy in Package 'CNAnorm'

#### **Description**

peakPloidy Estimate most likely ploidy of genome looking at distribution of smoothed ratio.

## Usage

```
## S4 method for signature 'CNAnorm'
peakPloidy(object, method = 'mixture', exclude = character(0),
    ploidyToTest = 12, sd = 5, dThresh = 0.01, n = 2048, adjust = .9, force.smooth = TRUE,
    reg = FALSE, ds = 1.5, zero.cont = FALSE, ...)
```

peakPloidy 19

#### **Arguments**

object An object of Class "CNAnorm"

exclude A character vector with names of Chromosomes/Contigs **not** to use to estimate

ploidy.

method A character element matching either "mixture", "density", "median", "mode"

or "closest". "mixture" will fit a mixture model to find peaks; "density" will use the density function to find peaks; "median" "mode" and "closest" will only find one peak at the median, mode or peak closest to the median respectively. No tumour content or reliable estimated ploidy will be provided. These methods are ment to perform a more "standard" normalisation, without stratching the data. Suggested for germline CNV or a fully automated process that does not requires a normalisation on integer copy number or for highly heterogeneous sample where such normalisation would not be possible. Non

ambigous partial matches can be used.

ploidyToTest Maximum ploidy allowed. Warnings! Computation time increases exponen-

tially with this parameter if using "density".

adjust The "adjust" parameter passed to the density function.

n The "n" parameter passed to the density function.

force.smooth If the input object does not have smoothed ratio, it will smooth using "addSmooth".

It is highly recomended to use "force.smooth = TRUE"

sd Parameter to filter outliers. Values greater than i median + sd \* standard deiva-

tions will be ignored while detecting peaks and ploidy.

dThresh Parameter to filter outliers. Values with a density lower than max(density)\*dThresh

will be ignored while detecting peaks and ploidy.

reg Parameter for mixture model: If set TRUE, the starting point for EM algorithm

will be optimized through several methods including regular grid on the ratio distribution. The default is FALSE, by which the starting values are taken from

the quantiles of the distribution.

ds Parameter for mixture model: A constraint in EM algorithm of minimum dis-

tance between mean estimates, in terms of median standard deviation of the

mixture components.

zero.cont Parameter for mixture model: An argument for the mixture model. If set TRUE,

the EM algorithm considers exactly-zero ratios as a mixture component.

.. Extra parameters to be passed to funtions for peak detection, specific to each of

the methods (deinsity or mixture), se below for details.

#### Value

An object of class "CNAnorm"

#### Note

Other optional parameters to be passed (...)

mixture method

20 plotGenome

#### density method

**peakRatio**Threshold used to call a peak. Peaks smaller than maxPeakHight/peakRatio are not considered peaks.

**spacingTolerance**A parameter smaller than 1 which describes how strict the program should be on alternative solutions. Only solution with the best  $R^2 >= \max(R^2)^*$  spacingTolerance will be considered as valid.

**interceptRatio**Minimum value for the intercept of the linear model. Ideally, should be zero, but the default allows a little flexibility.

#### Author(s)

Stefano Berri <s.berri@leeds.ac.uk> and Arief Gusnanto <a.gusnanto@leeds.ac.uk>

#### References

Gusnanto, A., Wood, H.M., Pawitan, Y., Rabbitts, P. and Berri, S. (2011) Correcting for cancer genome size and tumour cell content enables better estimation of copy number alterations from next generation sequence data. Bioinformatics

#### See Also

```
CNAnorm-class, density
```

#### **Examples**

```
data(LS041)
CN <- dataFrame2object(LS041)
chr2skip <- c("chrY", "chrM")
CN <- gcNorm(CN, exclude = chr2skip)
CN <- addSmooth(CN, lambda = 3)
CN <- peakPloidy(CN, exclude = chr2skip)
# this object CN is what you obtain when you load
# data(CN)</pre>
```

plotGenome

Methods for Function plotGenome in Package 'CNAnorm'

#### **Description**

plotGenome plot normalized ratio and optionally segmented and/or smoothed normalized ratio values in Package 'CNAnorm'. It also shows annotation.

## Usage

```
## S4 method for signature 'CNAnorm'
plotGenome(object, maxRatio = 8, minRatio = -1,
    superimpose = character(0), gPar = NULL, numHorLables = 10,
    colorful = FALSE, show.centromeres = TRUE, idiogram = NULL, fixVAxes = FALSE,
    supLineColor = character(0), supLineCex = character(0), dot.cex = .2, ...)
```

plotGenome 21

## Arguments

object	An object of Class "CNAnorm"
maxRatio	The maximum ratio to be shown on the plot. Values or ratio greater than maxRatio will be displayed as green triangulars
minRatio	The minimum ratio to be shown on the plot. Values or ratio smaller than min- Ratio will be displayed as green triangulars
superimpose	A character verctor with one or both of the following: "smooth", "DNACopy"
numHorLables	. Number of maximum horizontal lables. The function will try to annotate numHorLables so that they are approximately equally spaced.
colorful	A switch to decide if the background dots representing the ratio of each window should be gray or colored according their value in relation to the peak closest to the median
show.centromere	es
	A switch to decide if location of centromere are displayed on the graph. The location of the centromere is stored in idiogram
idiogram	A data frame containing banding information. if NULL -default- human information will be loaded by data(hg19_hs_ideogr)
fixVAxes	A switch to decide if the vertical axes should be fixed to minRatio and maxRatio or fit the data within minRatio and maxRatio.
gPar	a S3 object with all graphical parameters. If NULL (default) data(gPar) is called
supLineColor	A three element character vector with colors to be used for first superimposed line, second superimposed line, normalized ratio dots. If none is provided, defaults are: c("black", "cyan", "grey60")
supLineCex	A two element vector with cex valeus to be used for width of first superimposed line and second superimposed line. If none is provided, defaults are: $c(0.5,0.5)$
dot.cex	size of the dots in the plot
• • •	Further arguments to pass to the function plot

## Value

An object of class "CNAnorm"

## Author(s)

Stefano Berri <s.berri@leeds.ac.uk> and Arief Gusnanto <a.gusnanto@leeds.ac.uk>

## See Also

plot, par, peakPloidy, gPar, hg19\_hs\_ideogr

22 plotPeaks

#### **Examples**

```
data(CN)
# see peakPloidy documentation to know how object CN is created
CN <- addDNACopy(CN)
CN <- validation(CN)
CN <- discreteNorm(CN)
plotGenome(CN, superimpose = 'DNACopy')</pre>
```

plotPeaks

Methods for Function plotPeaks in Package 'CNAnorm'

## Description

plotPeaks plot annotated distribution of ratio Test/Normal

## Usage

## Arguments

object	An object of Class "CNAnorm"
special1	The chromosome/contig whose distribution will be shown with a different color
special2	The chromosome/contig whose distribution will be shown with a different color
show	A character verctor with one or both of the following: "suggested", "validated". Specify what need to be plotted
	Further arguments to pass to the function plot

## Author(s)

```
Stefano Berri <s.berri@leeds.ac.uk>
```

## See Also

```
plot, validation, peakPloidy
```

```
data(CN)
# see peakPloidy documentation to know how object CN is created
plotPeaks(CN, special1 = 'chrX', special2 = 'chrY')
```

ratio 23

ratio

Methods for Function ratio in Package 'CNAnorm'

## **Description**

ratio returns the Test/Normal ratio from an object of class CNAnorm. ratio is corrected for GC content if gcNorm was called.

ratio.n returns the Test/Normal **normalized** ratio from an object of class CNAnorm after normalization. Its input is ratio(object)

ratio.s returns the Test/Normal **smoothed** ratio from an object of class CNAnorm Its input is ratio(object)

ratio.s.n returns the Test/Normal **smoothed and normalized** ratio from an object of class CNAnorm. Its input is ratio.s(object)

segMean returns the mean of the segments as produced by DNACopy

segMean.n returns the **normalized** mean of the segments

## Usage

```
ratio(object)
ratio.n(object)
ratio.s(object)
ratio.s.n(object)
segMean(object)
segMean.n(object)
```

## Arguments

object

An object of Class "CNAnorm"

## Value

A numeric vector

#### Author(s)

Stefano Berri <s.berri@leeds.ac.uk>

#### See Also

```
gcNorm, CNAnorm-class, DNAcopy
```

## **Examples**

```
data(LS041)
CN <- dataFrame2object(LS041)
ratio.original <- ratio(CN)
CN.gcNorm <- gcNorm(CN, exclude = c("chrX", "chrY", "chrM"))
ratio.gc.corrected <- ratio(CN.gcNorm)</pre>
```

retrieve peaks and ploidy

Methods for Function to retrieve suggested/validated ploidy and peaks in Package 'CNAnorm'

## **Description**

sugg.peaks returns the location of peaks before normalization as suggested by peakPloidy.

sugg.ploidy returns the ploidy of the peaks as suggested by peakPloidy.

valid.peaks returns the location of peaks before normalization as validated after calling method "validation"

valid.ploidy returns the validated ploidy of the peaks as validated after calling method "validation"

## Usage

```
sugg.peaks(object)
sugg.ploidy(object)
valid.peaks(object)
valid.ploidy(object)
```

## **Arguments**

object

An object of Class "CNAnorm" after method "peakPloidy" was called

#### Value

A numeric vector

#### Author(s)

Stefano Berri <s.berri@leeds.ac.uk>

#### See Also

```
gcNorm, CNAnorm-class, DNAcopy
```

validation 25

#### **Examples**

```
data(CN)
# see peakPloidy documentation to know how object CN is created
plot(sugg.ploidy(CN), sugg.peaks(CN))
```

validation

Methods for Function addSmooth in Package 'CNAnorm'

## Description

validation segment and smooth perform ratio values in Package 'CNAnorm'

## Usage

```
## S4 method for signature 'CNAnorm'
validation(object, peaks = sugg.peaks(object),
    ploidy = sugg.ploidy(object))
```

## Arguments

object An object of Class "CNAnorm"

peaks The user validated location (ratio Test/Normal) of the peaks before normaliza-

tion.

ploidy The user validated ploidy of the peaks before normalization.

#### Value

An object of class "CNAnorm"

## Note

It is implicitly called by discreteNorm if no validation was manually performed

#### Author(s)

```
Stefano Berri <s.berri@leeds.ac.uk>
```

#### See Also

```
ratio.s, CNAnorm-class
```

```
data(CN) # see peakPloidy documentation to know how object CN is created CN <- validation(CN)
```

# **Index**

* classes	arms-methods (chromosomesPosition), 5
CNAnorm-class, 6	
DerivData-class, 10	${\it chromosomesPosition}, 5$
InData-class, 16	chrs (chromosomesPosition), 5
Params-class, 17	chrs, CNAnorm-method (CNAnorm-class), 6
* convert, data.frame	chrs-methods (chromosomesPosition), 5
dataFrame2object, 10	<pre>chrs&lt;- (chromosomesPosition), 5</pre>
* datasets, dataframe	chrs<-,CNAnorm-method(CNAnorm-class),6
LS041, 17	<pre>chrs<methods (chromosomesposition),="" 5<="" pre=""></methods></pre>
* datasets, object	CN, 6
CN, 6	CNAnorm, <i>11</i> , <i>16</i> , <i>18</i>
gPar, 15	CNAnorm (CNAnorm-class), 6
hg19_hs_ideogr, 15	CNAnorm-class, 6
* methods, accessor, get, set	CNAnormWorkflow, 8
ratio, 23	
retrieve peaks and ploidy, 24	data.frame, <i>10</i>
* methods, accessor, get	dataFrame2object, $9$ , $10$
chromosomesPosition, 5	density, 20
* methods, export, dataframe, table	DerivData, 7
exportTable, 13	DerivData-class, 10
* methods, normalization	discreteNorm, $9$ , $12$
gcNorm, 14	discreteNorm,CNAnorm-method
peakPloidy, 18	(discreteNorm), 12
* methods, segmentation, smoothing	discreteNorm-methods (discreteNorm), 12
addDNACopy, 2	
addSmooth, 4	exportTable, 13
* workflow	exportTable,CNAnorm-method
CNAnormWorkflow, 8	(exportTable), 13
[,CNAnorm-method (CNAnorm-class), 6	<pre>exportTable-methods (exportTable), 13</pre>
E, GIAMOT III III CETIOU (CIAMOT III CETU33), V	
addDNACopy, 2, 9	gcNorm, 5, 9, 14, 23, 24
addDNACopy, CNAnorm-method (addDNACopy),	gcNorm, CNAnorm-method (gcNorm), 14
2	gcNorm-methods (gcNorm), 14
addDNACopy-methods (addDNACopy), 2	gPar, 15, <i>21</i>
addSmooth, 4, 9	h-10 h- :d 2 5 15 21
addSmooth, CNAnorm-method (addSmooth), 4	hg19_hs_ideogr, 3, 5, 15, 21
addSmooth-methods (addSmooth), 4	InData 7
arms (chromosomesPosition), 5	InData-alass 16
arms, CNAnorm-method	InData-class, 16
(chromosomesPosition), 5	length, CNAnorm-method (CNAnorm-class), 6
(CITI OHIOSOHICSI OSTETOTI), S	rengen, environ in method (environ in class),

INDEX 27

<pre>length,DerivData-method     (DerivData-class), 10</pre>	<pre>segMean, CNAnorm-method (CNAnorm-class),</pre>
length, InData-method (InData-class), 16	segMean-methods(ratio), 23
loess, 14	segMean.n (ratio), 23
LS041, 17	segMean.n,CNAnorm-method
L3041, 17	(CNAnorm-class), 6
21	segMean.n-methods (ratio), 23
par, 21	sugg.peaks (retrieve peaks and ploidy),
Params-class, 17	24
peakPloidy, 9, 12, 18, 21, 22	<del>-</del> ·
<pre>peakPloidy, CNAnorm-method (peakPloidy),</pre>	sugg.peaks, CNAnorm-method
18	(CNAnorm-class), 6
<pre>peakPloidy-methods (peakPloidy), 18</pre>	sugg.peaks-methods(retrieve peaks and
plot, 21, 22	ploidy), 24
plotGenome, 20	sugg.ploidy(retrieve peaks and
<pre>plotGenome, CNAnorm-method (plotGenome),</pre>	ploidy), 24
20	sugg.ploidy,CNAnorm-method
plotGenome-methods (plotGenome), 20	(CNAnorm-class), 6
plotPeaks, 22	<pre>sugg.ploidy-methods(retrieve peaks</pre>
plotPeaks, CNAnorm-method (plotPeaks), 22	and ploidy), 24
plotPeaks-methods (plotPeaks), 22	
pos (chromosomesPosition), 5	valid.peaks (retrieve peaks and
pos, CNAnorm-method	ploidy), 24
(chromosomesPosition), 5	valid.peaks,CNAnorm-method
pos-methods (chromosomesPosition), 5	(CNAnorm-class), 6
pos<- (chromosomesPosition), 5	valid.peaks-methods(retrieve peaks
	and ploidy), 24
pos<-, CNAnorm-method (CNAnorm-class), 6	valid.ploidy(retrieve peaks and
pos <methods (chromosomesposition),="" 5<="" td=""><td>ploidy), 24</td></methods>	ploidy), 24
	valid.ploidy,CNAnorm-method
ratio, 14, 23	(CNAnorm-class), 6
ratio, CNAnorm-method (CNAnorm-class), 6	valid.ploidy-methods(retrieve peaks
ratio-methods (ratio), 23	and ploidy), 24
ratio.n (ratio), 23	validation, <i>12</i> , <i>22</i> , <i>25</i>
ratio.n,CNAnorm-method(CNAnorm-class),	validation, CNAnorm-method (validation),
6	25
ratio.n-methods(ratio), 23	validation-methods (validation), 25
ratio.s, 4, 25	, , , , , , , , , , , , , , , , , , , ,
ratio.s (ratio), 23	write.table, <i>14</i>
<pre>ratio.s,CNAnorm-method(CNAnorm-class),</pre>	
6	
ratio.s-methods(ratio), 23	
ratio.s.n(ratio), 23	
ratio.s.n,CNAnorm-method	
(CNAnorm-class), 6	
ratio.s.n-methods (ratio), 23	
retrieve peaks and ploidy, 24	
Tet. Tere peaks and proray, 27	
segMean, 3	
segMean (ratio). 23	
SEKITEALL (LALLU), 43	