

# Package ‘casper’

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**Title** Characterization of Alternative Splicing based on Paired-End Reads

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**Depends** R (>= 2.14.1), Biobase, IRanges, methods, GenomicRanges

**Imports** BiocGenerics, EBarrays, gaga, gtools, GenomeInfoDb, GenomicFeatures, limma, mgcV, Rsamtools, rtracklayer, S4Vectors, sqldf, survival, VGAM

**Enhances** parallel

**Description** Infer alternative splicing from paired-end RNA-seq data. The model is based on counting paths across exons, rather than pairwise exon connections, and estimates the fragment size and start distributions non-parametrically, which improves estimation precision.

**License** GPL (>=2)

**LazyLoad** yes

**Collate** GenericDefs.R ClassDefinitions.R asymmetryCheck.R calcDenovo.R calcExp.R casperVignettes.R createDenovoGenome.R genePlot.R getDistrs.R getRoc.R denovoExpr.R makeTranscriptDbFromGFF.R mergeBatches.R mergeExp.R modelPrior.R pathCounts.R probNonEquiv.R procBam.R procGenome.R qqnormGenomeWide.R relexprByGene.R rmShortInserts.R simMultSamples.R simPost.R simReads.R splitGenomeByLength.R truncnorm.R wrapKnown.R checks.R wrapDenovo.R

**biocViews** GeneExpression, DifferentialExpression, Transcription, RNASeq, Sequencing

**NeedsCompilation** yes

**R topics documented:**

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---

annotatedGenome-class *Class "annotatedGenome"*

---

### Description

Stores an annotated genome, either known or build de novo by combining a known genome with observed RNA-seq data.

### Objects from the Class

Objects are typically created with a call to `procGenome` (for known genomes) or to `createDenovoGenome` (for de novo genomes).

### Slots

**islands** GRangesList object with elements corresponding to gene islands. It indicates the start/end/name of each exon contained in the island

**transcripts** Each element in the list corresponds to a gene island. It indicates the exons contained in each known variant.

**exon2island** data.frame indicating the chromosome, start and end of each exon, and its corresponding gene island.

**exonsNI** GRanges indicating the chromosome, start/end and id of each exon

**aliases** data.frame indicating the aliases for each known transcript, i.e. transcripts having the exact same sequence of exons (as indicated in the UCSC database).

**genomeVersion** Character indicating the genome version from which the object was build, e.g. "hg19"

**dateCreated** Character indicating the date when the object was created. UCSC genomes change from time to time, so that an "hg19" genome from Jan 2012 may not be exactly the same as in Dec 2012.

**denovo** Logical variable. FALSE indicates that the object was created using available annotation only. TRUE indicates that new exons/islands were added based on the data observed in a particular RNA-seq experiment.

**txLength** Numeric vector storing transcript lengths.

**knownVars** List where each element corresponds to an island, and contains a character vector with names of isoforms that should be considered as known (i.e. always included in the model)

### Methods

**show** `signature(object = "annotatedGenome")`: Displays general information about the object.

### Author(s)

Camille Stephan-Otto Attolini

**See Also**

[procGenome](#) and [createDenovoGenome](#) to create annotatedGenome objects.

**Examples**

```
showClass("annotatedGenome")
```

---

asymmetryCheck	<i>Plot asymmetry coefficients for the observed data and compare to those expected under Normality.</i>
----------------	---

---

**Description**

Produces a boxplot for the asymmetry coefficients for each row in the input matrix. Normal observations are simulated using the observed sample means and variances, and their asymmetry coefficients are added to the plot.

**Usage**

```
asymmetryCheck(x, ...)
```

**Arguments**

x	ExpressionSet, matrix or data.frame with genes/isoforms in rows
...	Other arguments to be passed on to codeplot

**Value**

Boxplot with asymmetry coefficients for observed and simulated Normal data

**Author(s)**

David Rossell

**Examples**

```
mu <- rnorm(100)
x <- matrix(rnorm(100*5,mu),ncol=5)

asymmetryCheck(x)
```

---

calcDenovo	<i>Estimate expression of gene splicing variants de novo.</i>
------------	---

---

## Description

calcDenovo estimates expression of gene splicing variants, considering both known variants and variants that have not been previously described.

relativeExpr computes relative expression estimates from calcDenovo output via Bayesian model selection or averaging.

## Usage

```
calcDenovo(distrs, targetGenomeDB, knownGenomeDB=targetGenomeDB, pc,
readLength, islandid, priorq=3, mprior, minpp=0.001, selectBest=FALSE,
method="submodels", niter, exactMarginal=TRUE,
integrateMethod="plugin", verbose=TRUE, mc.cores=1)
```

```
relativeExpr(expr, summarize="modelAvg", minProbExpr=0.5, minExpr=0.05)
```

## Arguments

distrs	List of fragment distributions as generated by the getDistrs function
targetGenomeDB	annotatedGenome object with isoforms we wish to quantify. By default these are the same as in knownGenomeDB, but more typically targetGenomeDB is imported from a .gtf file produced by some isoform prediction software.
knownGenomeDB	annotatedGenome object with known isoforms, e.g. from UCSC or GENCODE annotations. Used to set the prior probability that any given isoform is expressed. knownGenome should be the same genome annotations used to create argument mprior (when provided)
pc	Named vector of exon path counts as returned by pathCounts
readLength	Read length in bp, e.g. in a paired-end experiment where 75bp are sequenced on each end one would set readLength=75.
islandid	Name of the gene island to be analyzed. If not specified, all gene islands are analyzed.
priorq	Parameter of the prior distribution on the proportion of reads coming from each variant. The prior is Dirichlet with prior sample size for each variant equal to priorq. We recommend priorq=3 as this defines a non-local prior that penalizes falsely predicted isoforms that show low expression.
mprior	Prior on the model space returned by modelPrior, used to favor isoforms consistent with knownGenomeDB. If left missing it is estimated from knownGenomeDB. See details.
minpp	Models (i.e. splicing configurations) with posterior probability less than minpp are not reported. This argument can help reduce substantially the amount of required memory to store the results.

selectBest	If set to TRUE only the model with highest posterior probability is reported. While this can save memory, we do not recommend this option as it may ignore a substantial amount of uncertainty.
method	Method used to perform the model search. "allmodels" enumerates all possible models (warning: this is not feasible for genes with >5 exons). "rwmcmc" uses a random-walk MCMC scheme to focus on models with high posterior probability. "submodels" considers that some isoforms in targetGenomeDB may not be expressed, but does not search for new variants. "auto" uses "allmodels" for genes with up to 5 exons and "rwmcmc" for longer genes. See details.
niter	Number of MCMC iterations.
exactMarginal	Set to FALSE to estimate posterior model probabilities as the proportion of MCMC visits. Set to TRUE to use the integrated likelihoods (default). See details.
integrateMethod	Method to compute integrated likelihoods. The default ('plugin') evaluates likelihood*prior at the posterior mode and is the faster option. Set 'Laplace' for Laplace approximations and 'IS' for Importance Sampling. The latter increases computation cost very substantially.
verbose	Set to TRUE to display progress information.
mc.cores	Number of processors to be used for parallel computation. Can only be used if the package multicore is available for your system. Warning: using multiple processors substantially increases the memory requirements, so set this value carefully.
expr	Object of class denovoGenomeExpr, as returned by calcDenovo.
summarize	Set to "modelAvg" to average estimates across models according to the posterior probability of each model. Set to "bestModel" to report the posterior mode, i.e. the estimate under the model with highest posterior probability.
minProbExpr	Variants with marginal posterior probability of being expressed below minProbExpr are not reported.
minExpr	Variants with estimated expression below minExpr are not reported.

## Details

calcDenovo explores which subset of the isoforms indicated in targetGenomeDB are truly expressed. It also adds new isoforms when some reads follow an exon path that is not possible under any of the isoforms in targetGenomeDB. calcDenovo the posterior probability of each model (i.e. configuration of expressed variants) via Bayes theorem.

$$P(\text{model}) \propto m(\text{ylmodel}) P(\text{model})$$

where  $m(\text{ylmodel})$  is the integrated likelihood and  $P(\text{model})$  is the prior probability of the model. For example, a gene with 20 predicted isoforms in targetGenome gives rise  $2^{20} - 1$  possible models (configurations of expressed isoforms).

Importantly,  $P(\text{model})$  can be set by analyzing available genome annotations in knownGenomeDB. For instance, genes with 20 exons have isoforms that tend to use most of the 20 exons. They also tend to express more isoforms than genes with 5 exons. The function modelPrior analyzes knownGenomeDB to set reasonable values for  $P(\text{model})$ .

An exhaustive enumeration of all possible models is not feasible unless the gene is very short (e.g. around 5 exons). For longer genes we use computational strategies to search a subset of "interesting" models. This is controlled by the argument `method` (see above).

In order to compute  $P(\text{model})$  one can either use the computed  $m(y|\text{model}) P(\text{model})$  (option `exactMarginal==TRUE`) or the proportion of MCMC visits (option `exactMarginal==FALSE`). Unless `niter` is large the former option typically provides more precise estimates.

### Value

Expression set with expression estimates. `featureNames` identify each transcript via RefSeq ids, and the `featureData` contains Entrez identifiers and the length of each transcript (in bp). If `citype` was set to a value other than "none", the `featureData` also contains the 95% credibility intervals (i.e. intervals that contain the true parameter value with 95% posterior probability).

### Author(s)

Camille Stephan-Otto Attolini, Manuel Kroiss, David Rossell

### References

Rossell D. QASPER: Quantifying Alternative Splicing from Paired End Reads. Technical report, IRB Barcelona (2010)

### See Also

[denovoExpr](#) to obtain expression estimates from the `calcDenovo` output. `plotExpr` to produce a plot with splicing variants and estimated expression.

### Examples

```
## See help(denovoExpr)
```

---

calcExp

*Estimate expression of a known set of gene splicing variants.*

---

### Description

Estimate expression of gene splicing variants, assuming that the set of variants is known. When `rpkms` is set to `TRUE`, fragments per kilobase per million are returned. Otherwise relative expression estimates are returned.

### Usage

```
calcExp(distrs, genomeDB, pc, readLength, islandid, rpkms=TRUE, priorq=2,  
priorqGeneExpr=2, citype="none", niter=10^3, burnin=100, mc.cores=1, verbose=FALSE)
```

**Arguments**

distrs	List of fragment distributions as generated by the getDistrs function
genomeDB	knownGenome object containing annotated genome, as returned by the procGenome function.
pc	Named vector of exon path counts as returned by pathCounts
readLength	Read length in bp, e.g. in a paired-end experiment where 75bp are sequenced on each end one would set readLength=75.
islandid	Name of the gene island to be analyzed. If not specified, all gene islands are analyzed.
rpkM	Set to FALSE to return relative expression levels, i.e. the proportion of reads generated from each variant per gene. These proportions add up to 1 for each gene. Set to TRUE to return fragments per kilobase per million (RPKM).
priorq	Parameter of the prior distribution on the proportion of reads coming from each variant. The prior is Dirichlet with prior sample size for each variant equal to priorq. We recommend priorq=2 for estimation, as it pools the estimated expression away from 0 and 1 and returned lower estimation errors than priorq=1 in our simulated experiments.
priorqGeneExpr	Parameter for prior distribution on overall gene expression. Defaults to 2, which ensures non-zero estimates for all genes
ciType	Set to "none" to return no credibility intervals. Set to "asympt" to return approximate 95% CIs (obtained via the delta method). Set to "exact" to obtain exact CIs via Monte Carlo simulation. Options "asympt" and especially "exact" can increase the computation time substantially.
niter	Number of Monte Carlo iterations. Only used when ciType=="exact".
burnin	Number of burnin Monte Carlo iterations. Only used when ciType=="exact".
mc.cores	Number of processors to be used for parallel computation. Can only be used if the package multicore is available for your system.
verbose	Set to TRUE to display progress information.

**Value**

Expression set with expression estimates. featureNames identify each transcript via RefSeq ids, and the featureData contains further information. If ciType was set to a value other than "none", the featureData also contains the 95% credibility intervals (i.e. intervals that contain the true parameter value with 95% posterior probability).

**Author(s)**

Camille Stephan-Otto Attolini, Manuel Kroiss, David Rossell

**References**

Rossell D, Stephan-Otto Attolini C, Kroiss M, Stocker A. Quantifying Alternative Splicing from Paired-End RNA-sequencing data. *Annals of Applied Statistics*, 8(1):309-330.



**See Also**

relexprByGene

**Examples**

```
data(K562.r111)
data(hg19DB)

#Pre-process
bam0 <- rmShortInserts(K562.r111, isizeMin=100)
pbam0 <- procBam(bam0)
head(getReads(pbam0))

#Estimate distributions, get path counts
distrs <- getDistrs(hg19DB,bam=bam0,readLength=75)
pc <- pathCounts(pbam0, DB=hg19DB)

#Get estimates
eset <- calcExp(distrs=distrs, genomeDB=hg19DB, pc=pc, readLength=75, rpkm=FALSE)
head(exprs(eset))
head(fData(eset))

#Re-normalize relative expression to add up to 1 within gene_id rather
# than island_id
eset <- relexprByGene(eset)

#Add fake sample by permuting and combine
eset2 <- eset[sample(1:nrow(eset),replace=FALSE),]
esetall <- mergeExp(eset,eset2)

#After merge samples are correctly matched
head(exprs(esetall))
head(fData(esetall))
```

---

denovoExpr

*Estimate expression for de novo splicing variants.*

---

**Description**

Obtains expression estimates from denovoGenomeExpr objects, as returned by calcDenovo. When rpkm is set to TRUE, fragments per kilobase per million are returned. Otherwise relative expression estimates are returned.

The estimates can be obtained by Bayesian model averaging (default) or by selecting the model with highest posterior probability. See details.

**Usage**

```
denovoExpr(x, pc, rpkm = TRUE, summarize = "modelAvg", minProbExpr = 0.5, minExpr = 0.05)
```

**Arguments**

x	denovoGenomeExpr object returned by calcExp
pc	Named vector of exon path counts as returned by pathCounts
rpkm	Set to FALSE to return relative expression levels, i.e. the proportion of reads generated from each variant per gene. These proportions add up to 1 for each gene. Set to TRUE to return fragments per kilobase per million (RPKM).
summarize	Set to "modelAvg" to obtain model averaging estimates, or to "bestModel" to select the model with highest posterior probability. We recommend the former, as even the best model may have low posterior probability.
minProbExpr	Variants with (marginal posterior) probability of being expressed below minProbExpr are omitted from the results. This argument is useful to eliminate variants that are not at least moderately supported by the data.
minExpr	Variants with relative expression minExpr are omitted from the results. This is useful to eliminate variants to which few reads are assigned, e.g. due to read miss-alignments or biases.

**Value**

Expression set with expression estimates. The featureData indicates the gene island id, posterior probability that each variant is expressed (column "probExpressed"), the number of aligned reads per gene island (column "explCnts") and the exons in each variant.

**Author(s)**

David Rossell

**References**

Rossell D, Stephan-Otto Attolini C, Kroiss M, Stocker A. Quantifying Alternative Splicing from Paired-End RNA-sequencing data. COBRA (<http://biostats.bepress.com/cobra/art97>)

**Examples**

```
## NOTE: toy example with few reads & genes to illustrate code usage
##      Results with complete data are much more interesting!

data(K562.r111)
data(hg19DB)

#Pre-process
bam0 <- rmShortInserts(K562.r111, isizeMin=100)
pbam0 <- procBam(bam0)

#Estimate distributions, get path counts
distrs <- getDistrs(hg19DB,bam=bam0,readLength=75)
pc <- pathCounts(pbam0, DB=hg19DB)

#Set prior distrib on model space
```

```

mprior <- modelPrior(hg19DB, maxExons=40, smooth=FALSE)

#Fit model
denovo <- calcDenovo(distrs, targetGenomeDB=hg19DB, pc=pc, readLength=75, priorq=3, mprior=mprior, minpp=0)

head(names(denovo))
denovo[['6499']]
posprob(denovo[['6499']])

#Get estimates
eset <- denovoExpr(denovo, pc=pc, rpkm=TRUE, minProbExpr=0.5)

head(exprs(eset))
head(fData(eset))

```

---

denovoGeneExpr-class    *Class "denovoGeneExpr"*

---

## Description

denovoGeneExpr stores inferred expression for de novo splicing variants for a single gene. denovoGenomeExpr stores the information for several genes (typically, the whole genome).

## Objects from the Class

Objects are returned by calcDenovo. When running calcDenovo on multiple genes results are returned in a denovoGenomeExpr object. Results for a single gene can be retrieved using the [[] operator as usual, which returns a denovoGeneExpr object.

## Slots

**posprob** data.frame containing the posterior probability of each model

**expression** data.frame with the estimated expression of each variant under each model

**variants** matrix indicating the exons contained in each variant.

**integralSum** Sum of the log(integrated likelihood) + log(model prior probability) across all considered models.

**npathDeleted** Number of paths that had 0 probability under all considered variants and had to be excluded for model fitting purposes.

**priorq** Input parameter to calcDenovo

**txLength** Length of transcripts in bp (including new isoforms found by casper)

**Methods**

**show** signature(object = "denovoGeneExpr"): Displays general information about the object.  
**names** Show names (island ids)  
**"["** Selects a subset of genes  
**"["["** Selects a single gene  
**posprob** Accesses the posterior probabilities of each model (slot posprob)  
**variants** Accesses the variant names and their respective exons  
**variants<-** Replaces the value of the slot variants (can be useful for renaming variants, for instance)

**Author(s)**

David Rossell

**See Also**

[calcDenovo](#) to create objects from the class. [denovoExpr](#) to obtain expression estimates from denovoGenomeExpr objects.

**Examples**

```
showClass("denovoGeneExpr")
```

---

```
denovoGenomeExpr-class
      Class "denovoGenomeExpr"
```

---

**Description**

denovoGeneExpr stores inferred expression for de novo splicing variants for a single gene. denovoGenomeExpr stores the information for several genes (typically, the whole genome).

**Objects from the Class**

Objects are returned by calcDenovo.

**Slots**

**islands** A list of denovoGeneExpr objects, with each element containing results for an individual gene.

**Methods**

**show** signature(object = "denovoGenomeExpr"): Displays general information about the object.  
**as.list** Coerces the object to a list  
**"["** Selects a subset of genes  
**"["["** Selects a single gene

**Author(s)**

Camille Stephan-Otto Attolini

**See Also**

[procGenome](#) and [createDenovoGenome](#) to create denovoGenomeExpr objects.

**Examples**

```
showClass("denovoGeneExpr")
showClass("denovoGenomeExpr")
```

---

distrsGSE37704	<i>Estimated read start and insert size distributions from MiSeq data in GEO dataset GSE37704.</i>
----------------	--

---

**Description**

We downloaded the fastq files, aligned with TopHat and processed with wrapKnown to obtain the estimated distributions for each of the 6 samples. `distrsGSE37704` is a list with the 6 corresponding elements. The estimated distributions for HiSeq data were very similar, hence these distributions can be used as defaults for Illumina MiSeq and HiSeq experiments.

**Usage**

```
data(distrsGSE37704)
```

**Format**

An list with 6 elements of class `readDistrs`. See `help(getDistrs)` and `help(readDistrs-class)` for details.

**Examples**

```
data(distrsGSE37704)
distrsGSE37704
plot(distrsGSE37704[[1]], 'readSt')
lines(distrsGSE37704[[2]], 'readSt', col=2)
plot(distrsGSE37704[[1]], 'fragLength')
```

---

genePlot *Plot exon structure for each transcript of a given gene.*

---

### Description

Plot exon structure for each transcript of a given gene. Optionally, aligned reads can be added to the plot.

### Usage

```
genePlot(generanges, islandid, genomeDB, reads, exp, names.arg, xlab='',
         ylab='', xlim, cex=1, yaxt='n', col, ...)
```

### Arguments

generanges	Object containing the ranges with start/end of each exon.
islandid	If generanges is not specified, transcripts are obtained from island islandid from the annotated genome genomeDB.
genomeDB	Annotated genome produced with the "procGenome" function
reads	pbam object with aligned reads. This is an optional argument.
exp	ExpressionSet object with expression values, as returned by calcExp. This is an optional argument.
names.arg	Optionally, indicate the names of each transcript.
xlab	x-axis label
ylab	y-axis label
xlim	x-axis limits, defaults to start of 1st exon and end of last exon
cex	Character expansion
yaxt	The y-axis in the plot has no interpretation, hence by default it is not displayed.
col	Either single color or vector of colors to be used to draw each transcript. Defaults to rainbow colors.
...	Other arguments to be passed on to plot.

### Value

A plot is produced.

### Methods

signature(generanges="CompressedIRangesList", islandid="ANY", genomeDB="ANY", reads="ANY", exp="ANY")  
 Plots a set of transcripts. Each element in the generanges corresponds to a transcript. Each transcript should contain exon start/end positions.

signature(generanges="IRanges", islandid="ANY", genomeDB="ANY", reads="ANY", exp="ANY")  
 Plots a single transcript. Each range indicates the start/end of a single exon.

`signature(generanges="IRangesList", islandid="ANY", genomeDB="ANY", reads="ANY", exp="ANY")`  
 Plots a set of transcripts. Each element in the `generanges` corresponds to a transcript. Each transcript should contain exon start/end positions.

`signature(generanges="GRangesList", islandid="ANY", genomeDB="ANY", reads="ANY", exp="ANY")`  
 Plots a set of transcripts. Each element in the `generanges` corresponds to a transcript. Each transcript should contain exon start/end positions.

`signature(generanges="GRanges", islandid="ANY", genomeDB="ANY", reads="ANY", exp="ANY")`  
 Plots a set of transcripts. Each space in `generanges` corresponds to a transcript. Each transcript should contain exon start/end positions.

`signature(generanges="missing", islandid="character", genomeDB="annotatedGenome", reads="GRanges", exp="ANY")`  
 Plots all transcripts stored in `genomeDB` for island with identifier `islandid`. Individual reads are added to the plot (`reads` contains start/end of individual read fragments).

`signature(generanges="missing", islandid="character", genomeDB="annotatedGenome", reads="missing", exp="ANY")`  
 Plots all transcripts stored in `genomeDB` for island with identifier `islandid`.

`signature(generanges="missing", islandid="character", genomeDB="annotatedGenome", reads="procBam", exp="ANY")`  
 Plots all transcripts stored in `genomeDB` for island with identifier `islandid`. Individual reads are added to the plot (`reads` contains start/end of individual read fragments).

`signature(generanges="missing", islandid="character", genomeDB="annotatedGenome", reads="procBam", exp="ANY", expEst="ANY")`  
 Plots all transcripts stored in `genomeDB` for island with identifier `islandid`. Individual reads and estimated expression are added to the plot (`reads` contains start/end of individual read fragments).

### Author(s)

Camille Stephan-Otto Attolini, David Rossell

### Examples

```
data(hg19DB)

#Plot an IRangesList
txs <- transcripts(entrezid="27", genomeDB=hg19DB)
genePlot(txs)

#Equivalently, indicate islandid
islandid <- getIsland(entrezid="27", genomeDB=hg19DB)
genePlot(islandid=islandid, genomeDB=hg19DB)
```

---

getDistrs

*Compute fragment start and fragment length distributions*

---

### Description

Compute fragment start distributions by using reads aligned to genes with only one annotated variant. Estimate fragment length distribution using fragments aligned to long exons (>1000nt). Fragment length is defined as the distance between the start of the left-end read and the end of the right-end read.

**Usage**

```
getDistrs(DB, bam, pbam, islandid, verbose, nreads=4*10^6, readLength)
```

**Arguments**

DB	Annotated genome. Object of class knownGenome as returned by procGenome.
bam	Aligned reads, as returned by scanBam. It must be a list with elements 'qname', 'rname', 'pos' and 'mpos'. Ignored when argument pbam is specified.
pbam	Processed BAM object of class procBam, as returned by function procBam. Arguments bam and readLength are ignored when pbam is specified.
islandid	Island IDs of islands to be used in the read start distribution calculations (defaults to genes with only one annotated variant)
verbose	Set to TRUE to print progress information.
nreads	To speed up computations, only the first nreads are used to obtain the estimates. The default value of 4 millions usually gives highly precise estimates.
readLength	Read length in bp, e.g. in a paired-end experiment where 75bp are sequenced on each end one would set readLength=75.

**Value**

An object of class readDistrs with slots:

lenDis	Table with number of fragments with a given length
stDis	Cumulative distribution function (object of type closure) for relative start position

**Author(s)**

Camille Stephan-Otto Attolini, David Rossell

**Examples**

```
data(K562.r111)
data(hg19DB)
bam0 <- rmShortInserts(K562.r111, isizeMin=100)

distrs <- getDistrs(hg19DB,bam=bam0,readLength=75)

#Fragment length distribution
plot(distrs,'fragLength')

#Fragment start distribution (relative to transcript length)
plot(distrs,'readSt')
```



---

getIsland	<i>getIsland returns the island id associated to a given entrez or transcript id in an annotatedGenome object. getChr indicates the chromosome for a given Entrez, transcript or island id.</i>
-----------	---

---

### Description

annotatedGenome objects store information regarding genes and transcripts. When there's an overlap in exons between several genes, these genes are grouped into gene islands. getIsland retrieves the island to which each gene or transcript was assigned, while getChr indicates the chromosome.

### Usage

```
getIsland(entrezid, txid, genomeDB)
getChr(entrezid, txid, islandid, genomeDB)
```

### Arguments

entrezid	Character indicating single Entrez identifier. Can be left missing and specify another identifier instead.
txid	Character indicating a single RefSeq transcript identifier. Can be left missing and specify another identifier instead.
islandid	Character indicating the gene island identifier. Can be left missing and specify another identifier instead.
genomeDB	Object of class annotatedGenome

### Value

Character with island identifier

### Methods

```
signature(entrezid='character', txid='missing', genomeDB='annotatedGenome') Return
  island id for given Entrez identifier
signature(entrezid='missing', txid='character', genomeDB='annotatedGenome') Return
  island id for given transcript identifier (RefSeq)
signature(entrezid='character', txid='missing', islandid='missing', genomeDB='annotatedGenome')
  Return chromosome for given Entrez identifier (RefSeq)
signature(entrezid='missing', txid='character', islandid='missing', genomeDB='annotatedGenome')
  Return chromosome for given transcript identifier (RefSeq)
signature(entrezid='missing', txid='missing', islandid='character', genomeDB='annotatedGenome')
  Return chromosome for given island identifier

signature(entrezid='character', txid='missing', islandid='missing') Return chromo-
  some for given Entrez identifier
```

signature(entrezid='missing', txid='character', islandid='missing') Return chromosome for given transcript identifier (RefSeq)

signature(entrezid='missing', txid='character', islandid='missing') Return chromosome for given island identifier

### Examples

```
data(hg19DB)
getIsland(entrezid="27", genomeDB=hg19DB)
getIsland(txid="NM_005158", genomeDB=hg19DB)

getChr(entrezid="27", genomeDB=hg19DB)
getChr(txid="NM_005158", genomeDB=hg19DB)
```

---

getNreads	<i>Get total number of paths in each island from a pathCounts object.</i>
-----------	---

---

### Description

getNreads returns a numeric vector with the total number of path counts in each island from a pathCounts object.

### Usage

```
getNreads(pc)
```

### Arguments

pc                    pathCounts object generated by pathCounts()

### Value

Numeric vector with total number of path counts in each island of pc.

### Methods

signature(pathCounts='pathCounts') Returns numeric vector with total number of path counts for each island in the pathCounts object.

### Author(s)

Camille Stephan-Otto Attolini

### Examples

```
##---- Should be DIRECTLY executable !! ----
##-- ==> Define data, use random,
##--or do help(data=index) for the standard data sets.
```

---

getReads	<i>getReads returns the reads stored in a procBam object.</i>
----------	---

---

**Description**

procBam objects store reads that have been split according to their CIGAR codes. getReads accesses these reads.

**Usage**

```
getReads(x)
```

**Arguments**

x	Object of class procBam
---	-------------------------

**Value**

RangedData object with reads stored in x.

**Methods**

signature(x='procBam') Return reads stored in x.

**Examples**

```
#See example in calcExp
```

---

getRoc	<i>Operating characteristics of differential expression analysis</i>
--------	--

---

**Description**

getRoc compares simulation truth and data analysis results to determine False Positives (FP), False Negatives (FN), True Positives (TP), True Negatives (TN), Positives (FP+TP), False Discovery Proportion (FP/P) and Power (TP/(TP+FN)).

**Usage**

```
getRoc(simTruth, decision)
```

**Arguments**

simTruth	Binary vector or matrix indicating simulation truth (FALSE or 0 for non differential expression, TRUE or 1 for differential expression)
decision	Binary vector or matrix with differential expression calls based on some data analysis.

**Value**

data.frame with TP, FP, TN, FN, P, FDR and Power.

**Methods**

`signature(simTruth='logical',decision='logical')` Operating characteristics are computed for a single simulation

`signature(simTruth='numeric',decision='numeric')` Operating characteristics are computed for a single simulation

`signature(simTruth='matrix',decision='matrix')` `simTruth` and `decision` contain truth and calls for several simulations (in columns). `getRoc` returns a data.frame with operating characteristics in each simulation.

**Author(s)**

David Rossell

**Examples**

```
## See help(probNonEquiv) for an example
```

---

hg19DB

*Subset of human genome (UCSC hg19 version)*

---

**Description**

We downloaded the human genome hg19 via `procGenome` and selected a few genes from chromosome 1 to use as a toy data for the vignette and examples.

**Usage**

```
data(hg19DB)
```

**Format**

An `annotatedGenome` object. See `help(procGenome)` and `help(annotatedGenome-class)` for details.

**Examples**

```
data(hg19DB)
hg19DB
slotNames(hg19DB)
```

---

`K562.r111`*Toy RNA-seq data from RGASP project.*

---

**Description**

The paired-end RNA-seq data is from the RGASP project sample K562\_2x75 (replicate 1, lane 1) and was obtained at [ftp://ftp.sanger.ac.uk/pub/gencode/rgasp/RGASP1/inputdata/human\\_fastq](ftp://ftp.sanger.ac.uk/pub/gencode/rgasp/RGASP1/inputdata/human_fastq). Reads were aligned against hg19 with tophat 2.0.2 and bowtie 0.12.5, setting the insert size at -r 200, and imported into R using scanBam from package Rsamtools. For illustration purposes, we selected reads mapping to a few genes only (namely, the genes that were also selected for the toy genome annotation in `data(hg19DB)`).

**Usage**

```
data(K562.r111)
```

**Format**

A list indicating read id, chromosome, start and end locations and the position of the pair, as returned by scanBam.

**Source**

```
ftp://ftp.sanger.ac.uk/pub/gencode/rgasp/RGASP1/inputdata/human_fastq
```

**References**

C Trapnell, L Pachter, SL Salzberg. TopHat: discovering splice junctions with RNA-Seq. *Bioinformatics*, 2009, 25, 1105-1111. doi=10.1093/bioinformatics/btp120.

B Langmead, C Trapnell, M Pop, SL Salzberg. Ultrafast and memory-efficient alignment of short DNA sequences to the human genome. *Genome Biology*, 2009, 10:R25.

**Examples**

```
data(K562.r111)
names(K562.r111)
```

---

mergeBatches	<i>Merge two ExpressionSet objects by doing quantile normalization and computing partial residuals (i.e. subtracting group mean expression in each batch). As currently implemented the method is only valid for balanced designs, e.g. each batch has the same number of samples per group.</i>
--------------	--

---

## Description

mergeBatches combines x and y into an ExpressionSet, performs quantile normalization and adjusts for batch effects by subtracting the mean expression in each batch (and then adding the grand mean so that the mean expression per gene is unaltered).

## Usage

```
mergeBatches(x, y, mc.cores=1)
```

## Arguments

x	ExpressionSet object with data from batch 1.
y	Either ExpressionSet object with data from batch 2, or simulatedSamples object with data from multiple simulations.
mc.cores	Number of processors to be used (ignored when y is an ExpressionSet)

## Value

When y is an ExpressionSet, mergeBatches returns an ExpressionSet with combined expressions. Its featureData contains a variable "batch" indicating the batch that each sample corresponded to.

When y is a simulatedSamples object, mergeBatches is applied to combine x with each dataset in y and a list of ExpressionSet objects is returned.

## Author(s)

David Rossell

## Examples

```
#Fake data from 2 batches
x <- matrix(rnorm(6),nrow=2)
colnames(x) <- paste('x',1:3,sep='')
y <- matrix(1+rnorm(6),nrow=2)
colnames(y) <- paste('y',1:3,sep='')
x <- new("ExpressionSet",exprs=x)
y <- new("ExpressionSet",exprs=y)
exprs(x)
exprs(y)
```

```
#Merge & adjust
z <- mergeBatches(x,y)
exprs(z)
```

---

mergeExp	<i>Merge splicing variant expression from multiple samples</i>
----------	--

---

### Description

mergeExp combines the output of calcExp from multiple samples, i.e. multiple ExpressionSet objects, into a single ExpressionSet

### Usage

```
mergeExp(..., sampleNames, keep=c('transcript','gene_id','island_id'))
```

### Arguments

...	ExpressionSet objects to be combined.
sampleNames	Character vector indicating the name of each sample. Defaults to 'Sample1', 'Sample2', etc.
keep	Variables in the featureData of each individual ExpressionSet to keep in the merged output.

### Details

mergeExp runs some checks to ensure that object can be combined (e.g. making sure that measurements are obtained on same set of genes), then sorts and formats each input ExpressionSet.

A label with the sample name is appended to variables in the featureData that appear in multiple samples, e.g. variable 'se' reporting standard errors (obtained by setting citype='asympt' in calcExp).

### Value

Object of class ExpressionSet combining the input ExpressionSets. Its featureData contains the columns indicated in the keep argument, plus a column readCount with the total number of reads mapped to each gene (or gene island, when multiple genes have overlapping exons).

### Author(s)

David Rossell

### See Also

calcExp to obtain an ExpressionSet for an individual sample.

**Examples**

```
#See example in calcExp
```

---

modelPrior	<i>Set prior distribution on expressed splicing variants.</i>
------------	---

---

**Description**

Set prior on expressed splicing variants using the genome annotation contained in a knownGenome object.

The prior probability of variants  $V_1, \dots, V_n$  being expressed depends on  $n$ , on the number of exons in each variant  $V_1, \dots, V_n$  and the number of exons in the gene. See the details section.

**Usage**

```
modelPrior(genomeDB, maxExons=40, smooth=TRUE, verbose=TRUE)
```

**Arguments**

genomeDB	Object of class knownGenome
maxExons	The prior distribution is estimated for genes with 1 up to maxExons exons. As there are fewer genes with many exons, the prior parameters are estimated poorly. To avoid this common estimate is used for all genes with more than maxExons exons
smooth	If set to TRUE the estimated prior distribution parameters for the number of exons in a gene are smoothed using Generalized Additive Models. This step typically improves the precision of the estimates, and is only applied to genes with 10 or more exons.
verbose	Set to TRUE to print progress information.

**Details**

The goal is to set a prior that takes into account the number of annotated variants for genes with  $E$  exons, as well as the number of exons in each variant.

Suppose we have a gene with  $E$  exons. Let  $V_1, \dots, V_n$  be  $n$  variants of interest and let  $|V_1|, \dots, |V_n|$  be the corresponding number of exons in each variant. The prior probability of variants  $V_1, \dots, V_n$  being expressed is modeled as

$$P(V_1, \dots, V_n | E) = P(n | E) P(|V_1| | E) \dots P(|V_n| | E)$$

where  $P(n | E) = \text{NegBinom}(n; k_E, r_E) I(0 < n < 2^E)$  and  $P(|V_{il}| | E) = \text{BetaBinomial}(|V_{il}| - 1; E - 1, \alpha_E, \beta_E)$ .

The parameters  $k_E, r_E, \alpha_E, \beta_E$  depend on  $E$  (the number of exons in the gene) and are estimated from the available annotation via maximum likelihood. Parameters are estimated jointly for all genes with  $E \geq \text{maxExons}$  in order to improve the precision.

For `smooth==TRUE`,  $\alpha_E$  and  $\beta_E$  are modeled as a smooth function of  $E$  by calling `gam` and setting the smoothing parameter via cross-validation. Estimates for genes with  $E \geq 10$  are substituted by their smooth versions, which typically helps improve stability in the estimates.



**Value**

List with 2 components.

nvarPrior	List with prior distribution on the number of expressed variants for genes with 1,2,3... exons. Each element contains the truncated Negative Binomial parameters, observed and predicted frequencies (counting the number of genes with a given number of variants).
nexonPrior	List with prior distribution on the number of exons in a variant for genes with 1,2,3... exons. Each element contains the Beta-Binomial parameters, observed and predicted frequencies (counting the number of variants with a given number of exons)

**Author(s)**

David Rossell, Camille Stephan-Otto Attolini

**Examples**

```
data(hg19DB)
mprior <- modelPrior(hg19DB, maxExons=10)

##Prior on number of expressed variants
##Genes with 2 exons
##mprior$nvarPrior[['2']]
##Genes with 3 exons
##mprior$nvarPrior[['3']]

##Prior on the number of exons in an expressed variant
##Genes with 2 exons
##mprior$nexonPrior[['2']]
##Genes with 3 exons
##mprior$nexonPrior[['3']]
```

---

modelPriorAS-class      *Class "modelPriorAS"*

---

**Description**

modelPriorAS stores parameters for the prior distribution on all possible alternative splicing configuration (i.e. prior on model space). This information is used for de novo reconstruction of splicing variants.

**Objects from the Class**

Objects are created by function modelPrior.

**Slots**

**nvarPrior** Prior on the number of variants per gene. A list with components "nbpar" containing the parameters of the Negative Binomial distribution, "obs" containing the observed counts and "pred" the Negative Binomial predicted counts.

**nexonPrior** Prior on the number of exons in an expressed variant. A list with components "bbpar" containing Beta-Binomial parameters, "obs" containing the observed counts and "pred" the Beta-Binomial predicted counts.

**Methods**

**show** signature(object = "modelPriorAS"): Displays general information about the object.

**"["** Selects prior parameters for genes with the specified number of exons

**coef** Selects a single gene

**Author(s)**

David Rossell

**See Also**

[procGenome](#) and [createDenovoGenome](#) to create modelPriorAS objects.

**Examples**

```
showClass("modelPriorAS")
```

---

pathCounts

*Compute exon path counts*

---

**Description**

Compute counts for exon paths visited by aligned reads

**Usage**

```
pathCounts(reads, DB, mc.cores = 1, verbose=FALSE)
```

**Arguments**

reads	Object of class procBam containing aligned reads, as returned by procBam.
DB	Object of class annotatedGenome containing either a known or de novo annotated genome.
mc.cores	Number of processors to be used for parallel computing. Requires having package multicore installed and loaded.
verbose	Set to TRUE to print progress information.

**Value**

Named integer vector with counts of exon paths. Names are character strings built as ".exon1.exon2-exon3.exon4.", with dashes making the split between exons visited by left and right-end reads correspondingly.

**Methods**

`signature(reads='list')` Computes counts for exon paths from a list of `procBam` objects (usually reads processed and split by chromosome).

`signature(reads='procBam')` Compute counts for exon paths from a `procBam` object of processed reads.

**Author(s)**

Camille Stephan-Otto Attolini

**See Also**

[procGenome](#) to create an annotated genome object, [createDenovoGenome](#) to create a de novo annotated genome. See `help(getNreads)` to get number of fragments mapping to each island.

**Examples**

```
##---- Should be DIRECTLY executable !! ----
##-- ==> Define data, use random,
##--or do help(data=index) for the standard data sets.
```

---

pathCounts-class      *Class "pathCounts"*

---

**Description**

Stores exon path counts.

**Objects from the Class**

Objects are created with a call to `pathCounts`.

**Slots**

**counts** List with one element per gene island. For each island, it contains a named vector with exon path counts. The names indicate the visited exons.

For instance, consider that for gene '1' with 2 exons we observe 10 reads in which the left end falls completely in exon 1 and the right end in exon 2. Suppose that for 5 reads the left end bridges exons 1-2 and the right end falls in exon 2. Then `pc[['1']]` would contain `c(10,5)` and `names(pc[['1']])` would contain `c(".1-2.", ".1.2-2.")`

**denovo** Logical variable. FALSE indicates that the counts correspond to a known genome (i.e. created with procGenome), and TRUE to a de novo annotated genome (i.e. created with createDenovoGenome).

**stranded** Logical variable. TRUE indicates that the path counts were obtained from an RNA-seq experiment where strand information was preserved.

## Methods

**show** signature(object = "pathCounts"): Displays general information about the object.

## Author(s)

Camille Stephan-Otto Attolini

## Examples

```
showClass("pathCounts")
```

---

plot-methods

*Plot estimated read start and fragment length distributions.*

---

## Description

Plots the estimated fragment length (insert size) distribution and the relative read start distribution (0 indicating transcription start, 1 transcription end). The former checks that the insert size distribution matches that described in the experimental protocol. The latter checks the extent to which reads are non-uniformly distributed (note: casper does NOT assume reads to be uniformly distributed, so a lack of uniformity is not a problem per se).

## Arguments

x	Object of type readDistrs, as returned by getDistrs.
y	Set to "fragLength" to plot the estimated insert size distribution. Set to "readSt" to plot a histogram of the estimated read start distribution.
...	Further arguments to be passed on to plot.

## Methods

signature(x = "readDistrs", y = "ANY") x is an object of type readDistrs, as returned by getDistrs. The plot function allows to visualize the fragment length and read start distributions.

signature(x = "readDistrs") x is an object of type readDistrs, as returned by getDistrs. The plot function allows to visualize the fragment length and read start distributions.

## Examples

```
#See getDistrs examples
```

---

plotExpr *Plot inferred gene structure and expression.*

---

### Description

Plots variants with sufficiently large posterior probability of being expressed along with their (marginal) estimated expression.

### Usage

```
plotExpr(gene, minProbExpr = 0.5, minExpr = 0.1,
         xlab = "(kb)", ylab = "", xlim, cex = 1, yaxt = "n", col, ...)
```

### Arguments

gene	denovoGeneExpr object containing results for a single gene, as returned by calcDenovo.
minProbExpr	Variants with marginal posterior probability of expression below minProbExpr are not reported
minExpr	Variants with (marginal) estimated expression below minExpr are not reported. Can be useful to remove sequence preference artifacts.
xlab	x-axis label, passed on to plot
ylab	y-axis label, passed on to plot
xlim	x-axis limits, passed on to plot
cex	Character expansion, passed on to plot
yaxt	Type of y-axis, passed on to plot
col	Colors for each variant, defaults to rainbow colors. It is possible to specify a single color.
...	Other arguments to be passed on to plot

### Details

The marginal posterior probability that a variant is expressed is the sum of the posterior probabilities of all models containing that variant.

The marginal estimated expression is the average expression across all models (including those where the variant has 0 expression) weighted by the posterior probability of each model.

### Methods

signature(gene = "denovoGeneExpr") gene contains the results from a de novo isoform expression analysis for a single gene, as returned by calcDenovo. When calcDenovo is run on multiple genes simultaneously, the desired gene can be selected using the "[[" operator as usual.

**Examples**

```
#See calcDenovo examples
```

---

plotPriorAS	<i>Plot prior distribution on set of expressed variants (i.e. the model space).</i>
-------------	---

---

**Description**

Plots the prior distribution on the number of expressed variants and the number of exons per variant in genes with exons (as returned by function modelPrior). The prior distribution is compared to the observed frequencies to check that the assumed distributional forms are reasonable.

**Usage**

```
plotPriorAS(object, type="nbVariants", exons=1:9, xlab,
            ylab="Probability", col=c("red","blue"))
```

**Arguments**

object	modelPriorAS object with prior distribution on model space.
type	Set to "nbVariants" to plot the prior on the number of variants per gene. Set to "nbExons" to plot the prior on the number of exons.
exons	Vector with integers. The plot is only produced with number of exons indicated in exons.
xlab	x-axis label, passed on to plot
ylab	y-axis label, passed on to plot
col	Colors for bars showing prior probabilities and frequencies in the known genome

**Methods**

signature(object = "modelPriorAS") object contains the prior distribution on the model space, as returned by function modelPrior

**Examples**

```
#See modelPrior examples
```

---

probNonEquiv	probNonEquiv performs a Bayesian hypothesis test for equivalence between group means. It returns the posterior probability that $ \mu_1 - \mu_2  > \log_{fc}$ . pvalTreat is a wrapper to treat in package limma, which returns P-values for the same hypothesis test.
--------------	--

---

### Description

probNonEquiv computes  $v_i = P(|\theta_i| > \log_{fc} \mid \text{data})$ , where  $\theta_i$  is the difference between group means for gene  $i$ . This posterior probability is based on the NNGCV model from package EBarrays, which has a formulation similar to limma in an empirical Bayes framework. Notice that the null hypothesis here is that  $|\theta_i| < \log_{fc}$ , e.g. isoforms with small fold changes are regarded as uninteresting.

Subsequent differential expression calls are based on selecting large  $v_i$ . For instance, selecting  $v_i \geq 0.95$  guarantees that the posterior expected false discovery proportion (a Bayesian FDR analog) is below 0.05.

### Usage

```
probNonEquiv(x, groups, logfc = log(2), minCount, method = "plugin", mc.cores=1)
pvalTreat(x, groups, logfc = log(2), minCount, p.adjust.method='none', mc.cores = 1)
```

### Arguments

x	ExpressionSet containing expression levels, or list of ExpressionSets
groups	Variable in fData(x) indicating the two groups to compare (the case with more than 2 groups is not implemented).
logfc	Biologically relevant threshold for the log fold change, i.e. difference between groups means in log-scale
minCount	If specified, probabilities are only computed for rows with <code>fData(x)\$readCount &gt;= minCount</code>
method	Set to 'exact' for exact posterior probabilities (slower), 'plugin' for plug-in approximation (much faster). Typically both give very similar results.
mc.cores	Number of parallel processors to use. Ignored unless x is a list.
p.adjust.method	P-value adjustment method, passed on to p.adjust

### Value

If x is a single ExpressionSet, probNonEquiv returns a vector with posterior probabilities (NA for rows with less than minCount reads). pvalTreat returns TREAT P-values instead.

If x is a list of ExpressionSet, the function is applied to each element separately and results are returned as columns in the output matrix.

**Author(s)**

Victor Pena, David Rossell

**References**

Rossell D, Stephan-Otto Attolini C, Kroiss M, Stocker A. Quantifying Alternative Splicing from Paired-End RNA-sequencing data. *Annals of Applied Statistics*, 8(1):309-330

McCarthy DJ, Smyth GK. Testing significance relative to a fold-change threshold is a TREAT. *Bioinformatics*, 25(6):765-771

**See Also**

treat in package limma, p.adjust

**Examples**

```
#Simulate toy data
p <- 50; n <- 10
x <- matrix(rnorm(p*2*n),nrow=p)
x[(p-10):p,1:n] <- x[(p-10):p,1:n] + 1.5
x <- new("ExpressionSet",exprs=x)
x$group <- rep(c('group1','group2'),each=n)

#Posterior probabilities
pp <- probNonEquiv(x, groups='group', logfc=0.5)
d <- rowMeans(exprs(x[,1:n])) - rowMeans(exprs(x[,-1:-n]))
plot(d,pp,xlab='Observed log-FC')
abline(v=c(-.5,.5))

#Check false positives
truth <- rep(c(FALSE,TRUE),c(p-11,11))
getRoc(truth, pp>.9)
getRoc(truth, pp>.5)
```

---

procBam

*Process BAM object*

---

**Description**

Process paired-end data stored in BAM object generated by scanBam. Outputs GRanges objects for reads and junctions.

**Usage**

```
procBam(bam, stranded, seed, verbose=FALSE, rname, keep.junx=FALSE,
keep.flag=FALSE, ...)
```



**Arguments**

bam	BAM object generated by scanBam
stranded	Set to TRUE to indicate that the RNA-seq experiment preserved the strand information.
seed	Seed for random number generator
verbose	Set to TRUE to print progress information.
rname	Chromosome to process be combined with the <code>\`which\`</code> argument in the scanBam function)
keep.junx	Option to store junction information. Only useful for finding denovo exons and transcripts.
keep.flag	Option to store alignment flag information.
...	Other arguments to be passed on to procBam.

**Details**

In case of multihits with same start position for both reads but different insertions/deletions patterns only one alignment is chosen at random.

**Value**

An object of class procBam containing reads with both ends correctly aligned and split according to the corresponding CIGAR. Unique identifiers by fragment are stored. Junctions spanned by reads are also stored in GRanges object if the argument `\`keep.junx\`` is set to TRUE.

**Methods**

`signature(bam='list',stranded='logical',seed='integer',verbose='logical', rname='character',keep.junx='logical')`  
Process paired-end data stored in BAM object generated by scanBam. Outputs GRanges objects for reads and (optionally) junctions.

**Author(s)**

Camille Stephan-Otto Attolini

**See Also**

scanBam from package Rsamtools, `help("procBam-class")`, `getReads`.

**Examples**

```
##See example in calcExp
```

---

procBam-class	Class "procBam"
---------------	-----------------

---

### Description

Stores processed bam files in a RangedData format. Each read is split into disjoint ranges according to its cigar code.

### Objects from the Class

Objects are created with a call to procBam.

### Slots

**pbam** GRanges indicating chromosome, start and end of each disjoint range. The pair id and read id within the pair are also stored.

**junx** GRanges indicating chromosome, start and end of junctions spanned by reads.

**stranded** Logical variable. TRUE indicates that the reads were obtained from an RNA-seq experiment where strand information was preserved.

In the case of stranded experiments:

**plus** GRanges indicating chromosome, start and end of each disjoint range for fragments originated from the positive strand. The pair id and read id within the pair are also stored.

**minus** GRanges indicating chromosome, start and end of each disjoint range for fragments originated from the negative strand. The pair id and read id within the pair are also stored.

**pjunx** GRanges indicating chromosome, start and end of junctions spanned by reads originated from the positive strand.

**mjunx** GRanges indicating chromosome, start and end of junctions spanned by reads originated from the negative strand.

### Methods

**show** signature(object = "procBam"): Displays general information about the object.

**getReads** signature(x = "procBam"): Extracts the aligned reads stored in x.

### Author(s)

Camille Stephan-Otto Attolini, David Rossell

### See Also

getReads

### Examples

```
showClass("procBam")
```

---

procGenome	<i>Create an annotatedGenome object that stores information about genes and transcripts</i>
------------	---

---

## Description

procGenome processes annotations for a given transcriptome, either from a TxDb object created by GenomicFeatures package (e.g. from UCSC) or from a user-provided GRanges object (e.g. by importing a gtf file).

createDenovoGenome creates a de novo annotated genome by combining UCSC annotations and observed RNA-seq data.

## Usage

```
procGenome(genDB, genome, mc.cores = 1)
```

```
createDenovoGenome(reads, DB, minLinks=2,
maxLinkDist=1e+05, maxDist=1000, minConn=2, minJunx=3, minLen=12, mc.cores=1)
```

## Arguments

genDB	Either a TxDb object with annotations (e.g. from UCSC or a gtf file or a GRanges object as returned by import from rtracklayer package). See details.
genome	Character indicating genome version (e.g. "hg19", "dm3")
mc.cores	Number of cores to use in parallel processing (multicore package required)
DB	annotatedGenome object, as returned by procGenome
minLinks	Minimum number of reads joining two exons to merge their corresponding genes
maxLinkDist	Maximum distance between two exons to merge their corresponding genes. A value of 0 disables this option.
maxDist	Maximum distance between two exons with reads joining them to merge their corresponding genes.
minConn	Minimum number of fragments connecting a new exon to an annotated one to add to denovo genome.
minJunx	Minimum number of junctions needed to redefine an annotated exon's end or start.
minLen	Minimum length of a junction to consider as a putative intron.
reads	Processed reads stored in a RangedData, as returned by procBam

## Details

These functions create the annotation objects that are needed for subsequent functions. Typically these objects are created only once for a set of samples.

If interested in quantifying expression for known transcripts only, one would typically use `procGenome` with a `TxDb` from the usual Bioconductor annotations, e.g. `genDB<-makeTranscriptDbFromUCSC(genome="hg19",tablename="refGene")` or imported from a `gtf` file e.g. `genDB<-makeTranscriptDbFromGFF('transcripts.gtf',format='gtf')`. `GRanges` object (e.g. `genDB <- import('transcripts.gtf')`). Package `GenomicFeatures` contains more info about how to create `TxDb` objects. Alternatively, one can provide annotations as a `GRanges` object which is returned when importing a `gtf` file with function `import` (package `rtracklayer`).

The output from `procGenome` can be used in combination with `wrapKnown`, which quantifies expression for a set of known transcripts, or `wrapDenovo`, which uses Bayesian model selection methods to assess which transcripts are truly expressed. When using `wrapDenovo`, you should create a single `annotatedGenome` object that combines information from all samples (e.g. from a `gtf` file produced by running your favorite isoform prediction software jointly on all samples), as this increases the power to detect new exons and isoforms.

## Value

Object of class `annotatedGenome`.

## Methods

`signature(genDB = "transcriptDb")` `genDB` is usually obtained with a call to `makeTranscriptDbFromUCSC` (package `GenomicFeatures`), e.g. `genDB<-makeTranscriptDbFromUCSC(genome="hg19",tablename="refGene")`

`signature(genDB = "GRanges")` `genDB` stores information about all transcripts and their respective exons. Chromosome, start, end and strand are stored as usual in `GRanges` objects. `genDB` must have a column named "type" taking the value "transcript" for rows corresponding to transcript and "exon" for rows corresponding to exons. It must also store transcript and gene ids. For instance, `Cufflinks` `RABT` module creates a `gtf` file with information formatted in this manner for known and de novo predicted isoforms.

## Author(s)

Camille Stephan-Otto Attolini

## See Also

See `annotatedGenome-class` for a description of the class. To obtain the island id for an entrez gene id or a transcript id see `help(getIsland)`. To extract the transcripts for a entrez or island id see `help(transcripts)`. See `splitGenomeByLength` for splitting an `annotatedGenome` according to gene length.

## Examples

```
##Known transcripts from UCSC
## genDB<-makeTranscriptDbFromUCSC(genome="hg19", tablename="refGene")
## hg19DB <- procGenome(genDB, "hg19")
```

```
##Transcripts inferred by Cufflinks-RABT module
##genDB.Cuff <- import('transcripts.gtf')
##hg19DB.Cuff <- procGenome(genDB.Cuff, genome='hg19')
```

---

qqnormGenomeWide      *Genome-wide qq-normal plots*

---

### Description

Overlays quantile-quantile normal plots (qqnorm) for a series of genes (rows in the input matrix), to provide an overall assessment of Normality.

### Usage

```
qqnormGenomeWide(x, ngenes=min(1000, nrow(x)), ...)
```

### Arguments

x	ExpressionSet, matrix or data.frame with genes/isoforms in rows
ngenes	A qqnorm plot is produced for the first ngenes rows in x
...	Other arguments to be passed on to codeplot

### Value

Produces a figure overlaying qq-normal plots for ngenes comparing observed vs. theoretical quantiles

### Author(s)

David Rossell

### Examples

```
mu <- rnorm(100)
x <- matrix(rnorm(100*5, mu), ncol=5)

qqnormGenomeWide(x)
```

---

quantileNorm	<i>Apply quantile normalization</i>
--------------	-------------------------------------

---

**Description**

Perform quantile normalization on the columns of a matrix or ExpressionSet

**Usage**

```
quantileNorm(x)
```

**Arguments**

x                    ExpressionSet or matrix

**Value**

Returns x with quantile normalized columns

**Author(s)**

David Rossell

**Examples**

```
x <- cbind(rnorm(1000), rnorm(1000, 2, 4))
boxplot(x)

xnorm <- quantileNorm(x)
boxplot(xnorm)
```

---

relexprByGene	<i>Compute relative expressions within each gene</i>
---------------	--

---

**Description**

Transforms relative expressions that add up to 1 within each gene island (the default output of casper) to relative expressions that add up to 1 per gene.

**Usage**

```
relexprByGene(x, normbylength=FALSE, genomeDB)
```

**Arguments**

x	ExpressionSet containing relative expressions. (typically, adding up to 1 for each island_id) Column gene_id in fData(x) should contain a unique gene identifier.
normbylength	If set to TRUE, isoform expressions are divided by isoform length before re-normalizing. This is useful for taking into account that longer isoforms produce more reads than shorter isoforms.
genomeDB	If normbylength==TRUE, genomeDB should be an annotatedGenome object containing the annotated genome (see procGenome)

**Value**

ExpressionSet with relative expressions adding up to one for each gene\_id.

**Author(s)**

David Rossell

**Examples**

```
#See help(calcExp)
```

---

rmShortInserts	<i>Remove reads with short insert sizes from imported BAM files.</i>
----------------	--

---

**Description**

In paired-end experiments short inserts (i.e. the 2 ends being very close to each other), may indicate RNA degradation or that a short RNA (e.g. miRNA) is being sequenced. Typically the goal is not to study alternative splicing for such short/degraded RNA; in this case it is recommendable to remove such short inserts to avoid biasing the insert size distribution. Requiring a minimum insert size can also result in significantly faster computations when quantifying alternative splicing via calc or calcDenovo.

**Usage**

```
rmShortInserts(bam, isizeMin=100)
```

**Arguments**

bam	Object with aligned reads, as returned by scanBam
isizeMin	Reads with insert size smaller than isizeMin will be removed.

**Value**

Named list, in the same format as that returned by scanBam.

**Note**

The insert size is stored in objects imported with scanBam in the element named isize.

**Author(s)**

David Rossell

**Examples**

```
##---- Should be DIRECTLY executable !! ----
##-- ==> Define data, use random,
##--or do help(data=index) for the standard data sets.
```

---

simMAE	<i>Simulate Mean Absolute Error (MAE) in estimating isoform expression under various experimental settings.</i>
--------	---

---

**Description**

Simulate several future RNA-seq data under various experimental settings (sequencing depth, read length, insert sizes), estimate isoform expression and assess the MAE incurred in the estimation process. The function is a wrapper combining functions simReads and calcExp.

**Usage**

```
simMAE(nsim, islandid, nreads, readLength, fragLength,
burnin=1000, pc, distr, readLength.pilot=readLength,
eset.pilot, usePilot=FALSE, retTxError=FALSE, genomeDB,
mc.cores=1, mc.cores.int=1, verbose=FALSE)
```

**Arguments**

nsim	Number of RNA-seq datasets to generate (often as little as nsim=10 suffice)
islandid	When specified this argument indicates to run the simulations only for gene islands with identifiers in islandid. When not specified genome-wide simulations are performed.
nreads	Vector indicating the target number of read pairs for each experimental setting. The actual number of reads differs from nreads to account for non-mappability and random read yield (see details)
readLength	Vector indicating the read length in each experimental setting
fragLength	Vector indicating the mean insert size in each experimental setting
burnin	Number of MCMC burn-in samples (passed on to calcExp)
pc	Observed path counts in pilot data. When not specified, these are simulated from eset.pilot



distr	Estimated read start and insert size distributions in pilot data
readLength.pilot	Read length in pilot data
eset.pilot	ExpressionSet with pilot data expression in log2-RPKM, used to simulate pc when not specified by the user. See details
usePilot	By default casper assumes that the pilot data is from a related experiment rather than the current tissue of interest (usePilot=FALSE). Hence, the pilot data is used to simulate new RNA-seq data but not to estimate its expression. However, in some cases we may be interested in re-sequencing the pilot sample at deeper length, in which case one would want to combine the pilot data with the new data to obtain more precise estimates. This can be achieved by setting usePilot=TRUE
retTxsError	If retTxsError=TRUE, simMAE returns posterior expected MAE for each individual isoform. This option is not available when eset.pilot is specified instead of pc. Else the output is a data.frame with overall MAE across all isoforms
genomeDB	annotatedGenome object, as returned by procGenome
mc.cores	Number of cores to use in the expression estimation step, passed on to calcExp
mc.cores.int	Number of cores to simulate RNA-seq datasets in parallel
verbose	Set verbose=TRUE to print progress information

## Details

simMAE simulates `nsim` datasets under each experimental setting defined by `nreads`, `readLength`, `fragLength`. For each dataset the following steps are performed:

1. The number of reads is  $nreads * readYield * pmapped$ , where  $readYield = runif(1,0.8,1.2)$  accounts for deviations in read yield and  $pmapped = runif(1,0.6,0.9) * pmappable$  is the proportion of mapped reads (60%-90% of the mappable reads according to the piecewise-linear power law of Li et al (2014))
2. True expression levels  $\pi_i$  are generated from their posterior distribution given the pilot data.
3. Conditional on  $\pi_i$ , RNA-seq data are generated and expression estimates  $\hat{\pi}_i$  are obtained using `calcExp`
4. The mean absolute estimation error  $\sum(\text{abs}(\hat{\pi}_i - \pi_i))$  across all isoforms is computed

Ideally simMAE should use pilot data from a relevant related experiment to simulate what future data may look like for the current experiment of interest. The recommended way to do this is to download a .bam file from such a related experiment and processing it in casper with function `wrapKnown`, as then both gene and isoform expression can be estimated accurately. The object output by `wrapKnown` is a list with elements named 'pc', 'distr' which can be given as input to simMAE.

As an alternative to specifying pc, simMAE allows setting `eset.pilot` as pilot data. Gene and isoform expression are then simulated as follows:

1. The number of reads per gene is generated from a Multinomial distribution with success probabilities proportional to  $2^{\text{exprs}\{\text{eset.pilot}\}}$ .
2. Relative isoform expression within each gene are generated from a symmetric Dirichlet distribution with parameter  $1/I_g$ , where  $I_g$  is the number of isoforms in gene  $g$ .

We emphasize that relative isoform expressions are not trained from the pilot data, and that while the distribution of gene expression levels resembles that in `eset.pilot`, no attempt is made to match gene identifiers and hence the results for individual genes should not be trusted (hence this option is only available when `retTxError==FALSE`).

### Value

If `retTxError==TRUE`, `simMAE` returns posterior expected MAE for each individual isoform. Else the output is a `data.frame` with overall MAE across all isoforms

### References

Stephan-Otto Attolini C., Pena V., Rossell D. Bayesian designs for personalized alternative splicing RNA-seq studies (2014)

Li, W. and Freudenberg, J. and Miramontes, P. Diminishing return for increased Mappability with longer sequencing reads: implications of the k-mer distributions in the human genome. *BMC Bioinformatics*, 15, 2 (2014)

### See Also

`wrapKnown`, `simReads`, `calcExp`

### Examples

```
## maybe str(simMAE) ; plot(simMAE) ...
```

---

simMAEcheck

*Model checking for One Sample Problems.*

---

### Description

Simulates RNA-seq data under the same experimental setting as in the observed data, and compares the observed vector of number of reads per gene with the simulations.

### Usage

```
simMAEcheck(nsim, islandid, burnin=1000, pc, distr, readLength.pilot, eset.pilot, usePilot=FALSE, ret
```

### Arguments

<code>nsim</code>	Number of RNA-seq datasets to generate (often as little as <code>nsim=10</code> suffice)
<code>islandid</code>	When specified this argument indicates to run the simulations only for gene islands with identifiers in <code>islandid</code> . When not specified genome-wide simulations are performed.
<code>burnin</code>	Number of MCMC burn-in samples (passed on to <code>calcExp</code> )
<code>pc</code>	Observed path counts in pilot data. When not specified, these are simulated from <code>eset.pilot</code>

distr	Estimated read start and insert size distributions in pilot data
readLength.pilot	Read length in pilot data
eset.pilot	ExpressionSet with pilot data expression in log2-RPKM, used to simulate pc when not specified by the user. See details
usePilot	By default casper assumes that the pilot data is from a related experiment rather than the current tissue of interest (usePilot=FALSE). Hence, the pilot data is used to simulate new RNA-seq data but not to estimate its expression. However, in some cases we may be interested in re-sequencing the pilot sample at deeper length, in which case one would want to combine the pilot data with the new data to obtain more precise estimates. This can be achieved by setting usePilot=TRUE
retTxsError	If retTxsError=TRUE, simMAE returns posterior expected MAE for each individual isoform. This option is not available when eset.pilot is specified instead of pc. Else the output is a data.frame with overall MAE across all isoforms
genomeDB	annotatedGenome object, as returned by procGenome
mc.cores	Number of cores to use in the expression estimation step, passed on to calcExp
mc.cores.int	Number of cores to simulate RNA-seq datasets in parallel
verbose	Set verbose=TRUE to print progress information

### Details

simMAEcheck simulates `nsim` datasets under the same experimental setting as in the observed data. For more details, please check the documentation for `simMAE`, which is the basis of this function.

### Value

The output is a list with 2 entries. The first entry is a `data.frame` with overall MAE across all isoforms in the simulations (see `simMAE` for details). The second entry contains the expected number of genes for which the number of reads in the data lies in the range of the posterior predictive simulations (under the hypothesis that they have the same distribution) and the actual number of genes for which the condition is satisfied.

### References

Stephan-Otto Attolini C., Pena V., Rossell D. Bayesian designs for personalized alternative splicing RNA-seq studies (2014)

Li, W. and Freudenberg, J. and Miramontes, P. Diminishing return for increased Mappability with longer sequencing reads: implications of the k-mer distributions in the human genome. BMC Bioinformatics, 15, 2 (2014)

### See Also

`wrapKnown`, `simReads`, `calcExp`

**Examples**

```
#Run casperDesign() to see full manual with examples
```

---

simMultSamples	<i>Simulate paired end reads for multiple future samples based on pilot data, and obtain their expression estimates via casper</i>
----------------	--

---

**Description**

Simulate true expression levels and observed data (casper expression estimates) for future samples within each group.

These simulations serve as the basis for sample size calculation: if one were to sequence `nsamples` new RNA-seq samples, what data would we expect to see? The simulation is posterior predictive, i.e. based on the current available data `x`.

**Usage**

```
simMultSamples(nsim, nsamples, nreads, readLength, fragLength=300, x,
  groups='group', distrs, genomeDB, verbose=TRUE, mc.cores=1)
```

**Arguments**

<code>nsim</code>	Number of simulations to obtain
<code>nsamples</code>	Vector indicating number of future samples per group, e.g. <code>nsamples=c(5,5)</code> to simulate 5 new samples for 2 groups.
<code>nreads</code>	Desired number of paired-end reads per sample. The actual number of aligned reads for any given sample differs from this amount, see details.
<code>readLength</code>	Read length, i.e. in an experiment with paired reads at 100bp each, <code>readLength=100</code> .
<code>fragLength</code>	Desired average insert size (size of RNA molecules after fragmentation)
<code>x</code>	ExpressionSet containing pilot data. <code>x[[group]]</code> indicates groups to be compared
<code>groups</code>	Name of column in <code>pData(x)</code> indicating the groups
<code>distrs</code>	Fragment start and length distributions. It can be either an object or a list of objects of class <code>readDistrs</code> . In the latter case, an element is chosen at random for each individual sample to consider uncertainty in these distributions. If not specified, it defaults to <code>data(distrsGSE37704)</code> .
<code>genomeDB</code>	<code>annotatedGenome</code> object
<code>verbose</code>	Set to <code>TRUE</code> to print progress
<code>mc.cores</code>	Number of cores to use in function. <code>mc.cores&gt;1</code> requires package <code>parallel</code>

## Details

The posterior predictive simulations is based on four steps: (1) simulate true expression for each group (mean and SD), (2) simulate true expression for future samples, (3) simulate paired reads for each future sample, (4) estimate expression from the reads via Casper. Below are some more details.

1. Simulate true mean expression in each group and residual variance for each gene. This is based on the NNGCV model in package EBarrays, adapted to take into account that the expression estimates in the pilot data  $x$  are noisy (which is why `simMultSamples` requires the SE / posterior SD associated to `exprs(x)`). The simulated values are returned in component `"simTruth"` of the `simMultSamples` output.

2. Simulate true isoform expression for each of the future samples. These are independent Normal draws with mean and variance generated in step 1. True gene expression is derived from the isoform expressions.

3. Determine the number of reads to be simulated for each gene based on its true expression (generated in step 2) and a Multinomial sampling model. For each sample:

- The number of reads yielded by the experiment is `Unif(.8*nreads,1.2*nreads)` - A proportion of non-mappable reads is discarded using the power law in Li et al (2014) - Amongst remaining reads, we assume that a proportion `Unif(0.6,0.9)` were aligned (consistently with reports from ENCODE project)

The final number of simulated reads is reported in component `"simExpr"` of the `simMultSamples` output.

4. Obtain expression estimates from the path counts produced in step 3 via `calcExp`. These are reported in component `"simExpr"` of the `simMultSamples` output.

## Value

Object of class `simulatedSamples`, which extends a list of length `nsim`. See the class documentation for some helpful methods (e.g. `coef`, `exprs`, `mergeBatches`). Each element is itself a list containing an individual simulation.

<code>simTruth</code>	<code>data.frame</code> indicating the mean and standard deviation of the Normal distribution used to generate data from each group
<code>simExpr</code>	<code>ExpressionSet</code> with Casper expression estimates, as returned by <code>calcExp</code> . <code>pData(simExpr)</code> indicates group information, and <code>fData(simExpr)</code> the number of simulated reads for each sample (in columns <code>'explCnts'</code> ) and across all samples (in column <code>'readCount'</code> )

## Author(s)

Victor Pena, David Rossell

## References

Stephan-Otto Attolini C., Pena V., Rossell D. Bayesian designs for personalized alternative splicing RNA-seq studies (2014)

**Examples**

```
#Run casperDesign() to see full manual with examples
```

---

simReads	<i>Function to simulate paired end reads following given read start and fragment length distributions and gene and variant expressions.</i>
----------	---

---

**Description**

This function generates path counts and bam files with simulated paired end reads according to given read start distribution, fragment length distribution and gene and variant expressions.

**Usage**

```
simReads(islandid, nSimReads, pis, rl, seed, writeBam, distrs, genomeDB,
repSims=FALSE, bamFile=NULL, stranded=FALSE, verbose=TRUE, chr=NULL, mc.cores=1)
```

**Arguments**

islandid	Island ID's from the genomeDB object to simulate reads
nSimReads	Named numeric vector with number of fragments to simulate in each island.
pis	Named numeric vector with relative expression of transcripts. Expressions add up to one for each island to simulate.
rl	Read length
seed	Seed of the random numbers generator
writeBam	Set to 1 to generate bam files with the simulated reads
distrs	Object of class 'readDistrs' with read start and fragment length distributions
genomeDB	Object of class 'annotatedGenome' with the genome to generate reads from
repSims	Set to TRUE to return relative read starts and fragment lengths from the simulation
bamFile	Name of the bam file to write reads to. Must end with '.bam'
stranded	Set to TRUE to preserve gene strand when generating reads. The 'XS' tag will be added to reads in the bam file and the returned 'pc' object will be stranded
verbose	Set to TRUE to print progress
chr	Characters vector with chromosomes to simulate. Defaults to whole genome simulations.
mc.cores	Number of cores to use in function

**Value**

Nsim	Numerical vector with the number of reads simulated for each island.
pc	Object of class 'pathCounts' with simulated path counts
sims	Only if 'repSims' is set to TRUE. List with vectors of length 'n' with the following elements: -'varl': Length of variant for corresponding read -'st' Start of fragment relative to variant start (not in genomic coordinates) -len:Fragment length -'strand':Strand of gene for simulated read

**Author(s)**

Camille Stephan-Otto Attolini

**Examples**

```

data(hg19DB)
data(K562.r111)
distrs <- getDistrs(hg19DB,bam=K562.r111,readLength=75)

islandid <- c('10319','463')
txs <- unlist(lapply(hg19DB@transcripts[islandid], names))
pis <- vector(mode='numeric', length=length(txs))
npis <- sapply(hg19DB@transcripts[islandid],length)
pis[1:npis[1]] <- rep(1/npis[1],npis[1])
pis[-1:-npis[1]] <- rep(1/npis[2],npis[2])
names(pis) <- txs
nSimReads <- c(100, 100)
names(nSimReads) <- islandid

simpc <- simReads(islandid=islandid, nSimReads=nSimReads, pis=pis,
r1=75, repSims=TRUE, seed=1, writeBam=FALSE, distrs=distrs,genomeDB=hg19DB)

```

---

simulatedSamples-class

*Class "simulatedSamples"*

---

**Description**

simulatedSamples stores multiple simulated isoform expression datasets. Each dataset contains the (simulation) true mean expression in each group and residual variance, as well as the estimated expression in each individual sample.

**Objects from the Class**

Objects are returned by simMultSamples.

**Slots**

The class extends a list directly.

**.Data** A list, each element containing a different simulated dataset

**Methods**

**show** signature(object = "simulatedSamples"): Displays general information about the object.

**coef** signature(object = "simulatedSamples"): Returns a matrix with difference between group means (simulation truth) in all simulated datasets

**exprs** signature(object = "simulatedSamples"): Returns a list of ExpressionSets containing the estimated expressions in each simulation.

**mergeBatches** signature(x="ExpressionSet",y="simulatedSamples"): Combines x with each element in exprs in y, and returns a list. See help(mergeBatches) for more details.

"[" x[i] selects a subset of simulations, x[,j] a subset of the samples in each simulation

**Author(s)**

David Rossell

**See Also**

[mergeBatches](#)

**Examples**

```
showClass("simulatedSamples")
```

---

splitGenomeByLength	<i>Split an annotatedGenome object into subsets according to gene length</i>
---------------------	--

---

**Description**

splitGenomeByLength splits an annotatedGenome according to gene length (bp), which allows estimating the fragment start and length distribution for each subset separately.

**Usage**

```
splitGenomeByLength(DB, breaks=c(0,3000,5000,Inf))
```

**Arguments**

DB Object containing annotated genome. Must be of class annotatedGenome, as returned by procGenome or createDenovoGenome.

breaks Breakpoints to define gene subgroups.



**Details**

By default groups are <3000bp, 3000-5000bp, >5000bp, which work well for the human genome. Further sub-divisions may result in unstable estimates of fragment start and length distributions.

**Value**

List where each component is of class annotatedGenome.

**Author(s)**

David Rossell

**See Also**

procGenome and createDenovoGenome for creating annotatedGenome objects. getDistrs for estimating fragment start and length distribution.

**Examples**

```
##Not run
## genDB<-makeTranscriptDbFromUCSC(genome="hg19", tablename="refGene")
## hg19DB <- procGenome(genDB, "hg19")
## hg19split <- splitGenomeByLength(hg19DB)
```

---

subsetGenome	<i>subsetGenome subsets an object of class annotatedGenome for a set of island IDs or chromosome names.</i>
--------------	---

---

**Description**

~~ Methods for function subsetGenome in package **casper** ~~ Subset an annotatedGenome object by islands or chromosomes.

**Usage**

```
subsetGenome(islands, chr, genomeDB)
```

**Arguments**

islands	Vector of characters with the island IDs to retrieve from genome.
chr	Vector of characters with the names of chromosomes to retrieve from genome.
genomeDB	annotatedGenome object with genome to subset.

**Methods**

```
signature(islands = "character", chr = "missing", genomeDB = "annotatedGenome")
  Subset annotatedGenome object by a set of island IDs.
signature(islands = "missing", chr = "character", genomeDB = "annotatedGenome")
  Subset annotatedGenome object by chromosomes.
```

---

transcripts	<i>Returns all transcripts associated to a given island or entrez id stored in an annotatedGenome object.</i>
-------------	---

---

### Description

annotatedGenome objects store information regarding genes and transcripts. When there's an overlap in exons between several genes, these genes are grouped into gene islands.

The function transcripts retrieves all stored transcripts for a given gene or island.

### Usage

```
transcripts(entrezid, islandid, genomeDB)
```

### Arguments

entrezid	Character indicating single Entrez identifier. Can be left missing and specify islandid instead.
islandid	Character indicating island identifier. Can be left missing and specify entrezid instead
genomeDB	Object of class annotatedGenome

### Value

IRangesList where each element in the list corresponds to a different transcript.

### Methods

signature(x = "readDistrs") x is an object of type readDistrs, as returned by getDistrs.  
The plot function allows to visualize the fragment length and read start distributions.

### See Also

genePlot to plot the resulting transcripts

### Examples

```
data(hg19DB)
tx <- transcripts(entrezid="27", genomeDB=hg19DB)
tx
```

---

txLength                      *~~ Methods for Function txLength in Package **casper** ~~*

---

## Description

~~ Methods for function txLength in package **casper** ~~ Function to retrieve transcript lengths from annotated genome (class genomeDB).

## Usage

```
txLength(islandid, txid, genomeDB)
```

## Arguments

islandid	Retrieve length for transcripts in island islandid.
txid	Retrieve length for txid transcripts.
genomeDB	Annotated genome of class genomeDB.

## Details

When called for the first time lengths are calculated and stored in the object genomeDB. Subsequent calls refer to these computed values.

## Value

Named numeric vector with transcript lengths.

## Methods

```
signature(islandid = "character", txid = "missing", genomeDB = "annotatedGenome")  
Retrieve lengths from genomeDB for transcripts in islandid islands.
```

```
signature(islandid = "missing", txid = "character", genomeDB = "annotatedGenome")  
Retrieve lengths from genomeDB for txid transcripts.
```

```
signature(islandid = "missing", txid = "missing", genomeDB = "annotatedGenome")  
Retrieve or calculate lengths for all transcripts in the annotated genome genomeDB.
```

---

wrapKnown	<i>Run all necessary steps to get expression estimates from multiple bam files with the casper pipeline.</i>
-----------	--

---

### Description

Function to analyze bam files to generate an ExpressionSet with expression estimates for all samples, read start and fragment length distributions, path counts and optionally processed reads.

### Usage

```
wrapKnown(bamFile, verbose=FALSE, seed=1, mc.cores.int=1,
mc.cores=1, genomeDB, readLength, rpkm=TRUE, priorq=2, priorqGeneExpr=2,
citype='none', niter=10^3, burnin=100, keep.pbam=FALSE,
keep.multihits=TRUE, chroms=NULL)
```

### Arguments

bamFile	Names of bam files with the sample to analyze. These must sorted and indexed, and the index must be in the same directory.
verbose	Set to TRUE to display progress information.
seed	Set seed of random number generator.
mc.cores.int	Number of cores to use when loading bam files. This is a memory intensive step, therefore number of cores must be chosen according to available RAM memory.
mc.cores	Number of cores to use in expression estimation.
genomeDB	annotatedGenome object containing annotated genome, as returned by the procGenome function.
readLength	Read length in bp, e.g. in a paired-end experiment where 75bp are sequenced on each end one would set readLength=75.
rpkm	Set to TRUE to return reads per kilobase per million (RPKM). Set to FALSE to return relative expression levels. Important, relative expression adds up to 1 within gene island, NOT within gene. To get relative expressions within gene run relexprByGene afterwards. See details.
priorq	Parameter of the prior distribution on the proportion of reads coming from each variant. The prior is Dirichlet with prior sample size for each variant equal to priorq. We recommend priorq=2 for estimation, as it pools the estimated expression away from 0 and 1 and returned lower estimation errors than priorq=1 in our simulated experiments.
priorqGeneExpr	Parameter for prior distribution on overall gene expression. Defaults to 2, which ensures non-zero estimates for all genes
citype	Set to "none" to return no credibility intervals. Set to "asypm" to return approximate 95% CIs (obtained via the delta method). Set to "exact" to obtain exact CIs via Monte Carlo simulation. Options "asypm" and especially "exact" can increase the computation time substantially.

niter	Number of Monte Carlo iterations. Only used when <code>citype=="exact"</code> .
burnin	Number of burnin Monte Carlo iterations. Only used when <code>citype=="exact"</code> .
keep.pbam	Set to TRUE to save processed bam object, as returned by <code>procBam</code> . This object can require substantial memory during execution and disk storage upon saving and is not needed for a default analysis.
keep.multihits	Set to FALSE to discard reads aligned to multiple positions.
chroms	Manually set chromosomes to be processed. By default only main chromosomes are considered (except 'chrM')

### Details

The function executes the functions `procBam`, `getDistrs` and `pathCounts` in parallel for each chromosome, but is much more efficient in cpu speed and memory usage than running these functions separately. Data from multiple samples are then combined using `mergeExp`. Note that further normalization (e.g. `quantileNorm`) may be needed preliminary to actual data analysis.

When `rpkm` is false the function returns the estimated proportion of reads arising from each isoform within a gene island. `casper` groups two or more genes into a gene island whenever these genes share an exon (or part of an exon). Because exons are shared, isoform quantification must be done simultaneously for all those genes.

That is, the output from `wrapKnown` when `rpkm` is FALSE are proportions that add up to 1 within each island. If you would like to re-normalize these expressions so that they add up to 1 within each gene, see the help for function `reexprByGene`.

One last remark: `casper` returns the estimated proportion of reads generated by each isoform, which is not the same as relative isoform expressions. Longer isoforms tend to produce more reads than shorter isoforms. This is easily accounted for by dividing relative expressions by isoform length, see `reexprByGene`.

### Value

<code>distr</code>	Object of class <code>readDistrs</code>
<code>pbam</code>	List of objects of class <code>procBam</code> with one element per chromosome
<code>pc</code>	Object of class <code>pathCounts</code>
<code>exp</code>	Object of class <code>ExpressionSet</code>

### Author(s)

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### References

Rossell D, Stephan-Otto Attolini C, Kroiss M, Stocker A. Quantifying Alternative Splicing from Paired-End RNA-sequencing data. *Annals of Applied Statistics*, 8(1):309-330.

### See Also

`procGenome`, `reexprByGene`, `quantileNorm`

**Examples**

```
## genDB<-makeTranscriptDbFromUCSC(genome="hg19", tablename="refGene")
## hg19DB <- procGenome(genDB, "hg19")
## bamFile="/path_to_bam/sorted.bam"
## ans <- wrapKnown(bamFile=bamFile, mc.cores.int=4, mc.cores=3, genomeDB=hg19DB, readLength=101)
## names(ans)
## head(exprs(ans$exp))
```

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