

# Introduction to *proBAMr*

Xiaojing Wang

April 16, 2015

## Contents

|          |  |          |
|----------|--|----------|
| <b>1</b> | <b>Why proBAM file</b>                       | <b>1</b> |
| <b>2</b> | <b>What is proBAM file</b>                   | <b>2</b> |
| <b>3</b> | <b>How to build proBAM file</b>              | <b>3</b> |
| 3.1      | Preparing annotation files . . . . .         | 3        |
| 3.2      | Preparing PSMs table . . . . .               | 4        |
| 3.3      | Generate SAM file using PSMtab2SAM . . . . . | 5        |
| 3.4      | Convert SAM file to BAM and index . . . . .  | 6        |
| 3.5      | Visulize proteomics data in IGV . . . . .    | 7        |
| <b>4</b> | <b>Session Information</b>                   | <b>7</b> |

## 1 Why proBAM file

Recent advances of sequencing technologies have reformed our conception of genomic data analysis, storage and interpretation, instigating more research interest in exploring human proteome at a parallel scale. Shotgun proteomics holds this promise by surveying proteome both qualitatively and quantitatively. Over the last years large amount of proteomics data has been accumulated, an emerging demand is to combine these efforts to catalogue the wide dynamic range of protein expression and complexity of alternative isoforms. However, this task is daunting due to the fact that different studies use varying databases, search engines and assembly tools. Such a challenge calls for an efficient approach of integrating data from different proteomics studies and even with genomic data.

Here we provide an R package, *proBAMr*, that maps identified PSMs to the genome in BAM format, a binary format for efficient data storage and fast access in genomic research field. This method differs from other approaches because of its ability of building connections between peptide and genomic location and simultaneously maintaining spectra count information. PSMs are aligned under the same coordination framework regardless of the annotation systems (e.g. RefSeq, ENSEMBL) of the input proteomics data, which enables flexible protein assembly switch between different annotation or at different level (gene or protein). When genomic/transcriptomic information of the same individual is available, this approach allows the co-analysis with -omics data together.

## 2 What is proBAM file

Table 1: Mandatory field definition of proBAM file and compare to original BAM format for genomic studies.

| No. | NAME  | TYPE   | BAM Description                             | proBAM description                |
|-----|-------|--------|---|-----------------------------------|
| 1   | QNAME | String | Query template NAME                         | Spectrum name                     |
| 2   | FLAG  | Int    | Bitwise FLAG                                | Bitwise FLAG                      |
| 3   | RNAME | String | Reference sequence NAME                     | Reference sequence NAME           |
| 4   | POS   | Int    | 1-based leftmost mapping POSition           | 1-based leftmost mapping POSition |
| 5   | MAPQ  | Int    | MAPPING Quality (Phred-scaled)              | -                                 |
| 6   | CIGAR | String | Extended CIGAR string (operations: MIDNSHP) | CIGAR string                      |
| 7   | RNEXT | String | Mate Reference NAME ('=' if same as RNAME)  | -                                 |
| 8   | PNEXT | Int    | 1-Based leftmost Mate POSition              | -                                 |
| 9   | TLEN  | Int    | observed Template LENgth                    | -                                 |
| 10  | SEQ   | String | segment SEQUENCE                            | Coding sequence                   |
| 11  | QUAL  | String | Query QUALity (ASCII-33=Phred base quality) | -                                 |

To take full advantage of tools developed for processing BAM files in genomics studies, we designed proBAM by incorporating features from the BAM file format and other features specifically for proteomics. Like BAM, it contains a header section and an alignment section. A full description of the BAM format is available at <http://samtools.github.io/hts-specs/SAMv1.pdf>. A PSGM (peptide-spectrum-genomic location-match) is the basic unit in proBAM and is similar to a read in NGS data. In Table 1, we compared the BAM and proBAM description of each mandatory column in the alignment section.

Table 2: FLAG description in proBAM file.

| Description  | Bit        | FLAG         |
|--|------------|--------------|
| Peptide map to forward strand  | 0x00       | 0            |
| Peptide map to reverse strand  | 0x10       | 16           |
| Peptide map to forward strand and it is <b>NOT</b> the rank=1 peptide for the spectrum | 0x00+0x100 | 256 (0+256)  |
| Peptide map to reverse strand and it is <b>NOT</b> the rank=1 peptide for the spectrum | 0x10+0x100 | 272 (16+256) |
| Peptide map to the decoy sequence  | 0x4        | 4            |

proBAM allows for 5 FLAG values due to the less complicated requirements by shotgun proteomics data (Table 2). For the same reason, the CIGAR tag in proBAM file only supports 'M' for

match/mismatch and 'N' for skipped bases on the reference.

The optional field keep extra information from proteomics experiment platform or search engines. The definition and value format of each optional column is described in Table 3. It is important to note that this table is extendable depending on continuous development and input from the community.

Table 3: Optional field definition of proBAM file.

| TAG | TYPE | Description   |
|-----|------|---|
| NH  | i    | Number genomic location the peptide mapping to  |
| XL  | i    | Number of peptides the spectrum mapping to  |
| XP  | Z    | Peptide sequence  |
| XS  | f    | score   |
| XC  | i    | Charge  |
| XA  | Z    | Whether the peptide is well annotated (0: yes ; 1: partially unknown; 2: totally unknown) |
| XM  | Z    | Modification site   |
| XN  | i    | Number of mis-cleavage  |
| XT  | I    | 0: non-tryptic; 1: semi-tryptic; 2: tryptic   |

The optional field follow the rule TAG:TYPE:VALUE defined by BAM file. There are three type of VALUE format: i, Signed 32-bit integer; Z, Printable string; f, Single-precision floating number.

### 3 How to build proBAM file

#### 3.1 Preparing annotation files

To map proteomics data to the genome, numerous pieces of genome annotation information are needed, such as genome elements region boundary, protein coding sequence and protein sequence et al. It is possible to manually download these data from different public resources (e.g. NCBI, UCSC and ENSEMBL) and then parse them to an appropriate format. To make this process more efficient and autonomous, we provide functions to prepare the gene/transcript annotation files from UCSC, ENSEMBL and GENCODE. The `PrepareAnnotationRefseq` and `PrepareAnnotationEnsembl` were included in another R package *customProDB* <http://bioconductor.org/packages/3.0/bioc/html/customProDB.html>. Here, we provide the function `PrepareAnnotationGENCODE` to prepare the annotation from GENCODE. This function requires users to download GTF file, coding sequence and protein sequence FASTA files from GENCODE ftp [ftp://ftp.sanger.ac.uk/pub/gencode/Gencode\\_human/](ftp://ftp.sanger.ac.uk/pub/gencode/Gencode_human/). Users should use the same version of annotations through the same project annalysis. All the annotations are saved to a specified directory for latter use.

```
> library(proBAMr)
```

```

> gtffFile <- system.file("extdata", "test.gtf", package="proBAMr")
> CDSfasta <- system.file("extdata", "coding_seq.fasta", package="proBAMr")
> pepfasta <- system.file("extdata", "pro_seq.fasta", package="proBAMr")
> annotation_path <- tempdir()
> PrepareAnnotationGENCODE(gtffFile, CDSfasta, pepfasta,
+                           annotation_path, dbsnp=NULL,
+                           splice_matrix=FALSE, COSMIC=FALSE)

```

### 3.2 Preparing PSMs table

After preparing all the annotation files, the R package *pepXMLTab* is used to extract confident PSMs and related information from pepXML files. Other tools are also applicable at this step, as long as it generates similar tabular files, as shown below.

```

> passedPSM <- read.table(system.file("extdata", "passedPSM.tab",
+   package="proBAMr"), sep='\t', header=TRUE)
> passedPSM[1:3, ]

```

|   | spectrum                                  |
|---|---|
| 1 | 00463_H12_P003361_B00L_A00_R1.9484.9484.2 |
| 2 | 00463_H12_P003361_B00L_A00_R1.9501.9501.2 |
| 3 | 00463_H12_P003361_B00L_A00_R1.9526.9526.2 |

  

|   | spectrumNativeID                              | start_scan | end_scan |
|---|---|------------|----------|
| 1 | controllerType=0 controllerNumber=1 scan=9484 | 9484       | 9484     |
| 2 | controllerType=0 controllerNumber=1 scan=9501 | 9501       | 9501     |
| 3 | controllerType=0 controllerNumber=1 scan=9526 | 9526       | 9526     |

  

|   | precursor_neutral_mass | assumed_charge | index | retention_time_sec |
|---|------------------------|----------------|-------|--------------------|
| 1 | 1945.011               | 2              | 1604  | 5941.112           |
| 2 | 1945.019               | 2              | 1614  | 5951.951           |
| 3 | 1945.016               | 2              | 1631  | 5963.760           |

  

|   | hit_rank | peptide            | peptide_prev_aa | peptide_next_aa |
|---|----------|--------------------|-----------------|-----------------|
| 1 | 1        | VNPTVFFDIAVDGEPLGR | M               | V               |
| 2 | 1        | VNPTVFFDIAVDGEPLGR | M               | V               |
| 3 | 1        | VNPTVFFDIAVDGEPLGR | M               | V               |

|   | proteins   |
|---|--|
| 1 | ENST00000415933.1 ENSG00000196262.9 OTTHUMG00000023687.5 OTTHUMT00000341788.1 PPIA-007 PPIA 12 |
| 2 | ENST00000415933.1 ENSG00000196262.9 OTTHUMG00000023687.5 OTTHUMT00000341788.1 PPIA-007 PPIA 12 |
| 3 | ENST00000415933.1 ENSG00000196262.9 OTTHUMG00000023687.5 OTTHUMT00000341788.1 PPIA-007 PPIA 12 |

  

|   | num_tot_proteins | calc_neutral_pep_mass | massdiff    | num_tol_term |
|---|------------------|-----------------------|-------------|--------------|
| 1 | 4                | 1944.995              | -0.01667663 | 2            |
| 2 | 4                | 1944.995              | -0.02461120 | 2            |
| 3 | 4                | 1944.995              | -0.02192565 | 2            |

  

|   | num_missed_cleavages | num_matched_ions | tot_num_ions | mvh      |
|---|----------------------|------------------|--------------|----------|
| 1 | 0                    | 21               | 31           | 46.20442 |
| 2 | 0                    | 26               | 31           | 60.50605 |
| 3 | 0                    | 22               | 31           | 52.51659 |

|   | mzFidelity | xcorr    | modification | NTT |
|---|------------|----------|--------------|-----|
| 1 | 81.97849   | 4.276119 | <NA>         | 1   |
| 2 | 104.25397  | 5.334539 | <NA>         | 1   |
| 3 | 86.18693   | 5.057394 | <NA>         | 1   |

### 3.3 Generate SAM file using PSMtab2SAM

The function PSMtab2SAM first finds the peptide location in protein sequences, then maps the coding sequence of the peptide back to the genome according to the annotation.

```
> load(system.file("extdata/GENCODE", "exon_anno.RData", package="proBAMr"))
> load(system.file("extdata/GENCODE", "proseq.RData", package="proBAMr"))
> load(system.file("extdata/GENCODE", "procodingseq.RData", package="proBAMr"))
> options(stringsAsFactors=FALSE)
> passedPSM <- read.table(system.file("extdata", "passedPSM.tab",
+   package="proBAMr"), sep='\t', header=TRUE)
> SAM <- PSMtab2SAM(passedPSM, XScolumn='mvh', exon, proteinseq,
+   procodingseq)
> write.table(SAM, file=paste(tempdir(), '/test.sam', sep=''),
+   sep='\t', quote=FALSE, row.names=FALSE, col.names=FALSE)
> dim(SAM)
```

```
[1] 40 21
```

```
> SAM[20:27, ]
```

|    | QNAME                                  | X1 | X2    | X3        | X4  |
|----|--|----|-------|-----------|-----|
| 20 | 00463_H12_P003361_B00L_A00_R1.0.1.7307 | 16 | chr11 | 65622810  | 255 |
| 21 | 00463_H12_P003361_B00L_A00_R1.0.1.7350 | 0  | chr7  | 44839340  | 255 |
| 22 | 00463_H12_P003361_B00L_A00_R1.0.1.7441 | 0  | chr7  | 44836381  | 255 |
| 23 | 00463_H12_P003361_B00L_A00_R1.0.1.7457 | 0  | chr7  | 44836381  | 255 |
| 24 | 00463_H12_P003361_B00L_A00_R1.0.1.7898 | 16 | chr5  | 133509648 | 255 |
| 25 | 00463_H12_P003361_B00L_A00_R1.0.1.7915 | 16 | chr5  | 133509648 | 255 |
| 26 | 00463_H12_P003361_B00L_A00_R1.0.1.7933 | 16 | chr5  | 133509648 | 255 |
| 27 | 00463_H12_P003361_B00L_A00_R1.0.1.7952 | 16 | chr1  | 26230237  | 255 |

  

|    | X5          | X6 | X7 | X8 |
|----|-------------|----|----|----|
| 20 | 42M         | *  | 0  | 0  |
| 21 | 45M         | *  | 0  | 0  |
| 22 | 12M2453N24M | *  | 0  | 0  |
| 23 | 12M2453N24M | *  | 0  | 0  |
| 24 | 51M         | *  | 0  | 0  |
| 25 | 51M         | *  | 0  | 0  |
| 26 | 51M         | *  | 0  | 0  |
| 27 | 39M         | *  | 0  | 0  |

  

|    | X9  | X10 | X11    |
|----|---|-----|--------|
| 20 | CAAAGGCTTGCCCTCCAGGGAGATGACGGCACTGCCCCCAG | *   | XA:Z:0 |

```

21      TCCATCTATGGGGAGAAAATTTGAAGATGAGAACTTCATCCTAAAG      * XA:Z:0
22      GTCTCCTTTGAGCTGTTTGCAGACAAGGTCCCAAAG      * XA:Z:0
23      GTCTCCTTTGAGCTGTTTGCAGACAAGGTCCCAAAG      * XA:Z:0
24 TTTGGCAATTTCCACATCAACTTCAAATATCTCTCCATCAGAACTCTGCAA      * XA:Z:0
25 TTTGGCAATTTCCACATCAACTTCAAATATCTCTCCATCAGAACTCTGCAA      * XA:Z:0
26 TTTGGCAATTTCCACATCAACTTCAAATATCTCTCCATCAGAACTCTGCAA      * XA:Z:0
27      CCGAGGGCTGAGAATCAGCTCAAAAGCCTGGCCTGAGGC      * XA:Z:0
      NH      XL      XP      XC      XS      XM
20 NH:i:1 XL:i:1      XP:Z:LGGSAVISLEGKPL XC:i:2 XS:f:30.6722 XM:Z:-
21 NH:i:1 XL:i:1      XP:Z:SIYGEKFEDENFILK XC:i:2 XS:f:22.4909 XM:Z:-
22 NH:i:1 XL:i:1      XP:Z:VSFELFADKVPK XC:i:2 XS:f:21.321 XM:Z:-
23 NH:i:1 XL:i:1      XP:Z:VSFELFADKVPK XC:i:2 XS:f:25.2581 XM:Z:-
24 NH:i:1 XL:i:1 XP:Z:LQSSDGEIFEVDVEIAK XC:i:2 XS:f:30.9829 XM:Z:-
25 NH:i:1 XL:i:1 XP:Z:LQSSDGEIFEVDVEIAK XC:i:2 XS:f:42.8235 XM:Z:-
26 NH:i:1 XL:i:1 XP:Z:LQSSDGEIFEVDVEIAK XC:i:2 XS:f:33.9951 XM:Z:-
27 NH:i:1 XL:i:1      XP:Z:ASGQAFELILSPR XC:i:2 XS:f:23.4655 XM:Z:-
      XN      XT      XG
20 XN:i:0 XT:i:2 XG:Z:N
21 XN:i:1 XT:i:2 XG:Z:N
22 XN:i:1 XT:i:2 XG:Z:N
23 XN:i:1 XT:i:2 XG:Z:N
24 XN:i:0 XT:i:2 XG:Z:N
25 XN:i:0 XT:i:2 XG:Z:N
26 XN:i:0 XT:i:2 XG:Z:N
27 XN:i:0 XT:i:2 XG:Z:N

```

### 3.4 Convert SAM file to BAM and index

Add the header to the SAM file. Converted them to the binary BAM files using samtools <http://samtools.sourceforge.net/>. Sort and index them for fast access.

The bullet list below summarizes the steps after the SAM file been generated.

```

> paste('cat header test.sam > test_header.sam')

[1] "cat header test.sam > test_header.sam"

> paste('samtools view -S -b test_header.sam > test_header.bam')

[1] "samtools view -S -b test_header.sam > test_header.bam"

> paste('samtools sort test_header.bam > test_header_sort')

[1] "samtools sort test_header.bam > test_header_sort"

> paste('samtools index test_header_sort')

[1] "samtools index test_header_sort"

```

### 3.5 Visualize proteomics data in IGV

The proBAM files can be visualized in IGV directly. Furthermore, users can co-visualize their proteomics data with the paired genomics/transcriptomics data, as shown in Fig 1.

Figure 1: IGV snapshot of a homozygous mutation in gene ALDH1B1 in both proteomics and RNA-Seq data (inside read box)



## 4 Session Information

R version 3.2.0 RC (2015-04-08 r68161)

Platform: x86\_64-w64-mingw32/x64 (64-bit)

Running under: Windows Server 2008 R2 x64 (build 7601) Service Pack 1

locale:

```
[1] LC_COLLATE=C
[2] LC_CTYPE=English_United States.1252
[3] LC_MONETARY=English_United States.1252
[4] LC_NUMERIC=C
[5] LC_TIME=English_United States.1252
```

attached base packages:

```
[1] stats4    parallel  stats     graphics grDevices utils
```

```
[7] datasets methods base
```

other attached packages:

```
[1] proBAMr_1.2.0      AnnotationDbi_1.30.0 GenomeInfoDb_1.4.0  
[4] Biobase_2.28.0     IRanges_2.2.0      S4Vectors_0.6.0  
[7] BiocGenerics_0.14.0
```

loaded via a namespace (and not attached):

```
[1] XVector_0.8.0      GenomicRanges_1.20.0  
[3] zlibbioc_1.14.0    GenomicAlignments_1.4.0  
[5] BiocParallel_1.2.0 tools_3.2.0  
[7] DBI_0.3.1          lambda.r_1.1.7  
[9] futile.logger_1.4  rtracklayer_1.28.0  
[11] futile.options_1.0.0 bitops_1.0-6  
[13] RCurl_1.95-4.5     biomaRt_2.24.0  
[15] RSQLite_1.0.0      GenomicFeatures_1.20.0  
[17] Biostrings_2.36.0  Rsamtools_1.20.0  
[19] XML_3.98-1.1
```