## Plotting using Genominator and GenomeGraphs (Beta)

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This vignette is preliminary, and should be viewed as subject to change. A number of the functions are not directly exported by the package – there is a reason for that.

In this vignette we demonstrate how to visualize data using the *GenomeGraphs* package. The main idea is that we want to build a plotting function which we can use to plot regions. The simplest case is the following:

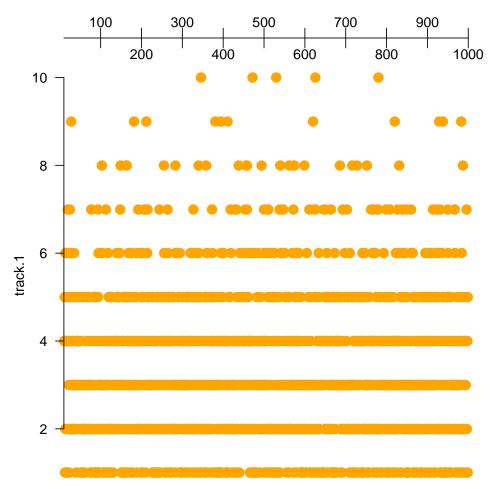
First, we make a database:

```
> require(Genominator)
> options(verbose = FALSE)
> N <- 1e+05
> K <- 100
> df <- data.frame(chr = sample(1:16, size = N, replace = TRUE),
+
      location = sample(1:1000, size = N, replace = TRUE), strand = sample(c(1L,
+
          -1L), size = N, replace = TRUE))
 eData <- aggregateExpData(importToExpData(df, dbFilename = "pmy.db",</pre>
>
      overwrite = TRUE, tablename = "ex_tbl"))
+
 annoData <- data.frame(chr = sample(1:16, size = K, replace = TRUE),
>
+
      strand = sample(c(1, -1), size = K, replace = TRUE), start = (st <- sample(1:1000,</pre>
+
          size = K, replace = TRUE)), end = st + rpois(K, 75),
      feature = c("gene", "intergenic")[sample(1:2, size = K, replace = TRUE)])
+
> rownames(annoData) <- paste("elt", 1:K, sep = ".")</pre>
> rp <- Genominator:::makeRegionPlotter(list(track.1 = list(expData = eData,
      what = "counts")))
> args(rp)
function (chr, start, end, overlays = NULL, title = NULL, ...)
```

This constructs a function which can be called to view particular pieces of data.

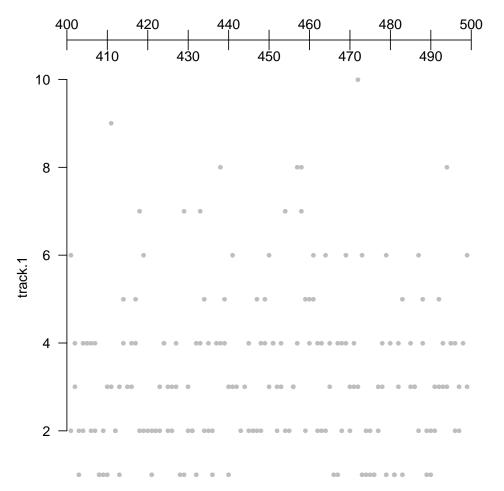
> rp(1, 10, 1000)

NULL

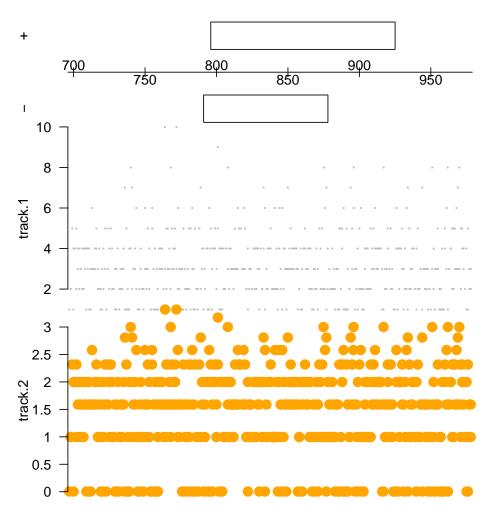


GenomeGraphs provides a wealth of customization options and means of plotting which for the most part are transferable using the list.

```
> rp <- Genominator:::makeRegionPlotter(list(track.1 = list(expData = eData,
+ what = "counts", dp = DisplayPars(lwd = 0.45, color = "grey"))))
> rp(1, 400, 500)
```

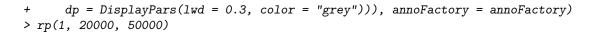


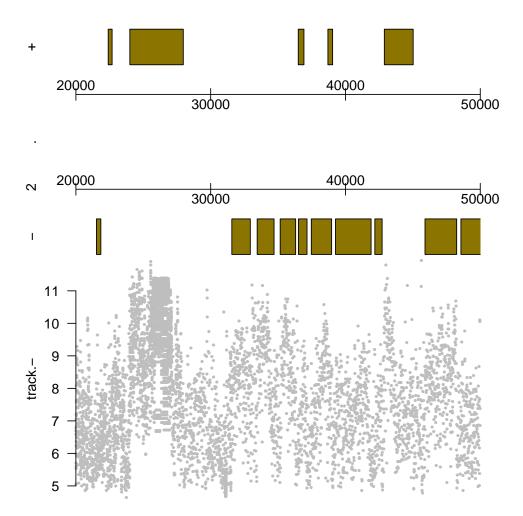
Here we can plot our annotation using the annotation factory construct. This is probably a little advanced. An easier thing is to use Ensembl to do the plotting of the annotation. Often, however, you will want to augment the annotation produced by Ensembl.



*GenomeGraphs* also offers a nice way to plot annotation for a given region using data from Ensembl or other sources of annotation - in some cases you have to do a little work because of the way that Biomart indexes the annotation and the way the *Genominator* package works (in this case yeast annotation is stored with Roman numerals denoting the chromosomes).

```
> require("biomaRt")
> mart <- useMart("ensembl", dataset = "scerevisiae_gene_ensembl")</pre>
> annoFactory <- Genominator:::makeAnnoFactory(mart, chrFunction = function(chr) as.roman(chr))
> load(system.file("data", "chr1_yeast.rda", package = "Genominator"))
> head(chr1_yeast)
  chr location strand
                        mRNA_1
                                  mRNA_2
    1
             1
                   -1 9.038919 8.614710
1
2
    1
             1
                   -1 9.172428 8.558421
3
    1
             2
                   -1 9.422065 9.131857
4
    1
             2
                   -1 8.679480 8.442943
5
    1
             2
                   -1 8.546894 8.794416
6
             2
                   -1 8.784635 8.918863
    1
> yData <- importToExpData(chr1_yeast, dbFilename = "my.db", tablename = "yeast",
+
      overwrite = TRUE)
> rp <- Genominator:::makeRegionPlotter(list(`track.-` = list(expData = yData,
      what = c("mRNA_1", "mRNA_2"), fx = rowMeans, strand = -1,
```





## SessionInfo

- R version 2.13.0 (2011-04-13), x86\_64-unknown-linux-gnu
- Locale: LC\_CTYPE=en\_US.UTF-8, LC\_NUMERIC=C, LC\_TIME=en\_US.UTF-8, LC\_COLLATE=C, LC\_MONETARY=C, LC\_MESSAGES=en\_US.UTF-8, LC\_PAPER=en\_US.UTF-8, LC\_NAME=C, LC\_ADDRESS=C, LC\_TELEPHONE=C, LC\_MEASUREMENT=en\_US.UTF-8, LC\_IDENTIFICATION=C
- Base packages: base, datasets, grDevices, graphics, grid, methods, stats, utils
- Other packages: DBI 0.2-5, GenomeGraphs 1.12.0, Genominator 1.6.0, IRanges 1.10.0, RSQLite 0.9-4, biomaRt 2.8.0
- Loaded via a namespace (and not attached): RCurl 1.5-0, XML 3.2-0, tools 2.13.0