

# phenoTest

October 25, 2011

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`ClusterPhenoTest` *Test association of clusters with phenotype.*

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## Description

Test the associations between clusters that each sample belongs to (based on gene expression) and each phenotype.

## Usage

```
ClusterPhenoTest(x, cluster, vars2test, B=10^4, p.adjust.method='none')
```

## Arguments

<code>x</code>	ExpressionSet with phenotype information stored in <code>pData(x)</code> .
<code>cluster</code>	variable of class <code>character</code> or <code>factor</code> telling at which cluster each sample belongs to.
<code>vars2test</code>	list with components <code>'continuous'</code> , <code>'categorical'</code> , <code>'ordinal'</code> and <code>'survival'</code> indicating which phenotype variables should be tested. <code>'continuous'</code> , <code>'categorical'</code> and <code>'ordinal'</code> must be character vectors, <code>'survival'</code> a matrix with columns named <code>'time'</code> and <code>'event'</code> . The names must match names in <code>names(pData(x))</code> .
<code>B</code>	An integer specifying the number of replicates used in the chi-square Monte Carlo test (passed on to <code>chisq.test</code> ).
<code>p.adjust.method</code>	Method for P-value adjustment, passed on to <code>p.adjust</code> .

## Details

Test association between the provided clusters and each phenotype.

For variables in `vars2test` `continuous` and `vars2test` `ordinal` a Kruskal-Wallis Rank Sum test is used; for `vars2test` `categorical` a chi-square test (with exact p-value if `simulate.p.value` is set to `TRUE`); for `vars2test` `survival` a Cox proportional hazards likelihood-ratio test.

## Author(s)

David Rossell

**Examples**

```

#load data
data(eset)
eset

#construct vars2test
survival <- matrix(c("Relapse", "Months2Relapse"), ncol=2, byrow=TRUE)
colnames(survival) <- c('event', 'time')
#add positive to have more than one category
pData(eset)[1:20, 'lymph.node.status'] <- 'positive'
vars2test <- list(survival=survival, categorical='lymph.node.status')
vars2test

#first half of the samples will be one cluster and the rest the other cluster
cluster <- c(rep('Cluster1', floor(ncol(eset)/2)), rep('Cluster2', ncol(eset)-floor(ncol(eset)/2)))

#test association
ClusterPhenoTest(eset, cluster, vars2test=vars2test)

```

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ExpressionPhenoTest

*Tests univariate association between a list of phenotype variables and*

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**Description**

Tests univariate association between a list of phenotype variables and gene expression.

**Usage**

```

ExpressionPhenoTest(x, vars2test, adjustVars,
  p.adjust.method='BH', continuousCategories=3, mc.cores)

```

**Arguments**

<code>x</code>	ExpressionSet containing expression levels in <code>exprs(x)</code> and phenotype information in <code>pData(x)</code> .
<code>vars2test</code>	list with components 'continuous', 'categorical', 'ordinal' and 'survival' indicating which phenotype variables should be tested. 'continuous', 'categorical' and 'ordinal' must be character vectors, 'survival' a matrix with columns named 'time' and 'event'. The names must match names in <code>names(pData(x))</code> .
<code>adjustVars</code>	variables that will be used as adjustment variables when fitting linear models and/or cox models. These variables have to exist in <code>colnames(pData(x))</code> .
<code>p.adjust.method</code>	method for p-value adjustment, passed on to <code>p.adjust</code> . Valid values are <code>c("holm", "hochberg", "hommel", "bonferroni", "BH", "BY", "fdr", "none")</code> .
<code>continuousCategories</code>	number of categories used for continuous variables.
<code>mc.cores</code>	the number of cores to use, i.e. how many processes will be spawned (at most).

**Details**

The effect of both continuous, categorical and ordinal phenotype variables on gene expression levels are tested via `lmFit`. For ordinal variables a single coefficient is used to test its effect on gene expression (trend test), which is then used to obtain a P-value (means for each category are reported in the output). Gene expression effects on survival are tested via Cox proportional hazards model, as implemented in function `'coxph'`.

**Value**

The output is an `epheno` object, which basically extends an `ExpressionSet` object. The means, fold changes, standardized hazard ratios and pvalues are stored in the `experimentData` slot which is accessible with the `exprs` method. Information about the kind of information of each variable can be found in the `phenoData` slot which is accessible with the `pData` method.

There are several methods that can be used to access the information stored in an `epheno` object. For more information please type one of the following: `getFc(x)`, `getHr(x)`, `getMeans(x)`, `getPvals(x)`, `getSummaryDif(x)`, `logFcHr(x)`, `p.adjust.method(x)`, `phenoClass(x)`, `phenoNames(x)`.

**Author(s)**

David Rossell

**Examples**

```
#load eset
data(eset)
eset

#prepare vars2test
survival <- matrix(c("Relapse","Months2Relapse"),ncol=2,byrow=TRUE)
colnames(survival) <- c('event','time')
vars2test <- list(survival=survival)

#run ExpressionPhenoTest
epheno <- ExpressionPhenoTest(eset,vars2test,p.adjust.method='none')
epheno
```

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barplotSignatures-methods

*Methods for Function barplotSignatures in Package 'phenoTest'*

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**Description**

Methods for function `barplotSignatures` in Package `'phenoTest'`. For more information read the function's manual.

**Methods**

`signature(x = "epheno", signatures = "character")` Method for an `epheno` object and one signature stored in an object of class `character`.

`signature(x = "epheno", signatures = "GeneSetCollection")` Method for an `epheno` object and several signatures stored in an object of class `GeneSetCollection`.

`signature(x = "epheno", signatures = "GeneSet")` Method for an epheno object and one signature stored in an object of class `GeneSet`.

`signature(x = "epheno", signatures = "list")` Method for an epheno object and several signatures stored in an object of class `list`.

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`barplotSignifSignatures-methods`

*Methods for Function `barplotSignifSignatures` in Package 'phenoTest'*

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### Description

Methods for function `barplotSignifSignatures` in Package 'phenoTest'. For more information read the function's manual.

### Methods

`signature(x = "epheno", signatures = "character")` Method for an epheno object and one signature stored in an object of class `character`.

`signature(x = "epheno", signatures = "list")` Method for an epheno object and several signatures stored in an object of class `list`.

`signature(x = "epheno", signatures = "GeneSet")` Method for an epheno object and one signature stored in an object of class `GeneSet`.

`signature(x = "epheno", signatures = "GeneSetCollection")` Method for an epheno object and several signatures stored in an object of class `GeneSetCollection`.

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`epheno-class`

*Class "epheno"*

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### Description

Object obtained with the `ExpressionPhenoTest` function. Contains FC, HR and pvals from testing expression values of each gene against phenotypic variables.

### Objects from the Class

Objects can be created by calls of the form `new("epheno", assayData, phenoData, featureData, exprs, ...)`.

### Slots

`p.adjust.method`: Object of class `"character"` containing the multiple testing adjustment method used (if one was used).

`assayData`: Object of class `"AssayData"` that is inherited from the `ExpressionSet` object used to create the epheno object.

`phenoData`: Object of class `"AnnotatedDataFrame"` that contains information about the variables stored in the `experimentData` slot such as their class (continuous, categorical, etc) or type (mean, `summaryDif`, `pval`, etc).

**featureData:** Object of class "AnnotatedDataFrame" that is inherited from the ExpressionSet object used to create the epheno object.

**experimentData:** Object of class "MIAME" that is inherited from the ExpressionSet object used to create the epheno object.

**annotation:** Object of class "character" that is inherited from the ExpressionSet object used to create the epheno object.

**protocolData:** Object of class "AnnotatedDataFrame" that is inherited from the ExpressionSet object used to create the epheno object.

**.\_classVersion\_:** Object of class "Versions" that is inherited from the ExpressionSet object used to create the epheno object.

### Extends

Class "[ExpressionSet](#)", directly. Class "[eSet](#)", by class "ExpressionSet", distance 2. Class "[VersionedBiobase](#)", by class "ExpressionSet", distance 3. Class "[Versioned](#)", by class "ExpressionSet", distance 4.

### Methods

[ signature(x = "epheno", i = "ANY", j = "ANY"): inherited from the ExpressionSet class.

**dim** signature(x = "epheno"): inherited from the ExpressionSet class.

**export2CSV** signature(x = "epheno"): ...

**getFc** signature(x = "epheno"): getter for the fold changes.

**getHr** signature(x = "epheno"): getter for the hazard ratios.

**getMeans** signature(x = "epheno"): getter for the means.

**getPvals** signature(x = "epheno"): getter for the pvalues.

**getSummaryDif** signature(x = "epheno"): getter that returns hazard ratios, fold changes and pvalues.

**gseaSignatures** signature(x = "epheno", signatures = "list"): Used to compute GSEA. Please read the gseaSignatures manual.

**logFcHr** signature(x = "epheno"): getter for the log of fold changes and hazard ratios.

**p.adjust.method** signature(x = "epheno"): getter for the p value adjustment method that has been used.

**phenoClass** signature(x = "epheno"): Returns the class off all variables.

**phenoNames** signature(x = "epheno"): Returns the names of the tested phenotypes.

**show** signature(object = "epheno"): Shows a brief overview of the object.

### Author(s)

Evarist Planet

### Examples

```
showClass("epheno")
```

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 epheno

 epheno object.
 

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## Description

Object obtained with ExpressionPhenoTest function.

## Usage

```
data(epheno)
```

## Format

The format is: Formal class 'epheno' [package "phenoTest"] with 8 slots ..@ p.adjust.method : chr "none" ..@ assayData :<environment: 0x1050d5a78> ..@ phenoData :Formal class 'Annotated-DataFrame' [package "Biobase"] with 4 slots .. ..@ varMetadata :'data.frame': 5 obs. of 1 variable: .. .. ..\$ labelDescription: chr [1:5] NA NA NA NA ... .. ..@ data :'data.frame': 12 obs. of 5 variables: .. .. ..\$ phenoName : Factor w/ 3 levels "lymph.node.status",...: 3 3 3 3 3 1 1 1 2 3 ... .. .. ..\$ phenoClass: Factor w/ 3 levels "categorical",...: 2 2 2 2 2 1 1 1 3 2 ... .. .. ..\$ phenoType : Factor w/ 3 levels "mean","pval",...: 1 1 1 3 3 1 1 3 3 2 ... .. .. ..\$ meanLabel : Factor w/ 5 levels "[45.2,49.2)",...: 1 2 3 NA NA 4 5 NA NA NA ... .. .. ..\$ survTime : Factor w/ 1 level "Months2Relapse": NA NA NA NA NA NA NA NA NA 1 NA ... .. ..@ dimLabels : chr [1:2] "sampleNames" "sampleColumns" .. ..@ \_\_classVersion\_\_:Formal class 'Versions' [package "Biobase"] with 1 slots .. .. ..@ .Data:List of 1 .. .. .. ..\$ : int [1:3] 1 1 0 ..@ featureData :Formal class 'AnnotatedDataFrame' [package "Biobase"] with 4 slots .. .. ..@ varMetadata :'data.frame': 0 obs. of 1 variable: .. .. ..\$ labelDescription: chr(0) .. .. ..@ data :'data.frame': 1000 obs. of 0 variables .. .. ..@ dimLabels : chr [1:2] "featureNames" "featureColumns" .. ..@ \_\_classVersion\_\_:Formal class 'Versions' [package "Biobase"] with 1 slots .. .. ..@ .Data:List of 1 .. .. .. ..\$ : int [1:3] 1 1 0 ..@ experimentData :Formal class 'MIAME' [package "Biobase"] with 13 slots .. .. ..@ name : chr "" .. .. ..@ lab : chr "" .. .. ..@ contact : chr "" .. .. ..@ title : chr "" .. .. ..@ abstract : chr "" .. .. ..@ url : chr "" .. .. ..@ pubMedIds : chr "" .. .. ..@ samples : list() .. .. ..@ hybridizations : list() .. .. ..@ normControls : list() .. .. ..@ preprocessing : list() .. .. ..@ other : list() .. .. ..@ \_\_classVersion\_\_:Formal class 'Versions' [package "Biobase"] with 1 slots .. .. ..@ .Data:List of 1 .. .. .. ..\$ : int [1:3] 1 1 0 ..@ annotation : chr "hgu133a" ..@ protocolData :Formal class 'AnnotatedDataFrame' [package "Biobase"] with 4 slots .. .. ..@ varMetadata :'data.frame': 0 obs. of 1 variable: .. .. ..\$ labelDescription: chr(0) .. .. ..@ data :'data.frame': 12 obs. of 0 variables .. .. ..@ dimLabels : chr [1:2] "sampleNames" "sampleColumns" .. .. ..@ \_\_classVersion\_\_:Formal class 'Versions' [package "Biobase"] with 1 slots .. .. ..@ .Data:List of 1 .. .. .. ..\$ : int [1:3] 1 1 0 ..@ \_\_classVersion\_\_:Formal class 'Versions' [package "Biobase"] with 1 slots .. .. ..@ .Data:List of 4 .. .. .. ..\$ : int [1:3] 2 12 0 .. .. .. ..\$ : int [1:3] 2 10 0 .. .. .. ..\$ : int [1:3] 1 3 0 .. .. .. ..\$ : int [1:3] 1 0 0

## Examples

```
data(epheno)
## maybe str(epheno) ; plot(epheno) ...
```

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epheno2html                      *Create html files and plots from an epheno object.*

---

## Description

Creates html files and plots using an epheno object, which stores the association between a list of variables and gene expression.

## Usage

```
epheno2html(x, epheno, outputdir, prefix = "", genelimit = 50, categories = 3, w
```

## Arguments

<code>x</code>	An object of class <code>ExpressionSet</code> (used to generate the epheno object) containing expression levels in <code>exprs(x)</code> , phenotype information in <code>pData(x)</code> and annotation in <code>annotation(x)</code> .
<code>epheno</code>	an object produced by <code>ExpressionPhenoTest</code> . this object will contain univariate association between a list of phenotype variables and gene expression as well as p-values.
<code>outputdir</code>	where to place files.
<code>prefix</code>	will be used to add a text to the beginning of the files that will be created.
<code>genelimit</code>	maximum number of genes on the list.
<code>categories</code>	Number of categories used for continuous variables. It has to be the same as the one used for <code>ExpressionPhenoTest</code> .
<code>withPlots</code>	when <code>FALSE</code> no plots will be produced. Makes the process faster.
<code>id.entrezid</code>	this is for the particular case in which <code>x</code> 's <code>featureNames</code> is not storing probeset ids but entrezids. The <code>ExpressionSet</code> will be storing information at the gene level and the row identifiers will be entrezids.
<code>organism</code>	this information will be used to query the BioMart database in order to get feature information. It will be used only if <code>id.entrezid</code> is <code>TRUE</code> .
<code>homol.symbol</code>	the homology table can be provided. This is useful if we want to run the same/similar code several times. This one will store the homology between entrezid and gene symbol. It will be used only if <code>id.entrezid</code> is <code>TRUE</code> . An example on how to create the homology table is shown in the examples section.
<code>homol.genename</code>	the same as <code>homol.symbol</code> but in this case the homology table stores the homology between entrezid and gene name.
<code>mc.cores</code>	number of cores that will be used to run the process.

## Author(s)

Evarist Planet

## Examples

```
#Example on building homology tables for human.
#mart <- useMart("ensembl", "hsapiens_gene_ensembl")
#homol.symbol <- getLDS(attributes = c("entrezgene"),
#   mart = mart, attributesL = c("external_gene_id"),
#   martL = mart, filters = "entrezgene", values = entrezid)
#mart <- useMart("ensembl", "hsapiens_gene_ensembl")
#homol.genename <- getLDS(attributes = c("entrezgene"),
#   mart = mart, attributesL = c("description"), martL = mart,
#   filters = "entrezgene", values = entrezid)
```

---

getters for the epheno object

*Getters for the epheno object:*

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## Description

`getFc` gets the fold changes. `getHr` gets the hazard ratios. `getMeans` gets the means. `getPvals` gets the p values. `getSummaryDif` gets fold changes and hazard ratios. `lofFcHr` gets the fold changes and hazard ratios after log scaling. `p.adjust.method` gets the p value adjustment method that was used when creating the object. `phenoClass` returns a data.frame telling the class (ordinal, continuous, categorical or survival) of each phenotype. `phenoNames` gets the phenotype names.

## Usage

```
getFc(x)
getHr(x)
getMeans(x)
getPvals(x)
getSummaryDif(x)
logFcHr(x)
p.adjust.method(x)
phenoClass(x)
phenoNames(x)
```

## Arguments

x                    epheno object

## Author(s)

Evarist Planet

eset

*Example data.*

**Description**

Example data of class ExpressionSet.

**Usage**

data(eset)

**Format**

The format is: Formal class 'ExpressionSet' [package "Biobase"] with 7 slots ..@ assayData :<environment: 0x1050d9390> ..@ phenoData :Formal class 'AnnotatedDataFrame' [package "Biobase"] with 4 slots .. ..@ varMetadata :'data.frame': 7 obs. of 1 variable: .. .. .. ..\$ labelDescription: chr [1:7] NA NA NA NA ... .. .. ..@ data :'data.frame': 286 obs. of 7 variables: .. .. .. ..\$ PID : int [1:286] 3 5 6 7 8 9 11 14 15 17 ... .. .. .. ..\$ GEOaccession : Factor w/ 286 levels "GSM36777","GSM36778",...: 17 20 21 22 24 25 58 59 60 61 ... .. .. .. ..\$ lymph.node.status: chr [1:286] "negative" "negative" "negative" "negative" ... .. .. .. ..\$ Months2Relapse : int [1:286] 101 118 9 106 37 125 109 14 99 137 ... .. .. .. ..\$ Relapse : int [1:286] 0 0 1 0 1 0 1 0 0 1 0 0 ... .. .. .. ..\$ ER.Status : num [1:286] 0 1 0 0 0 1 1 0 1 1 ... .. .. .. ..\$ BrainRelapse : int [1:286] 0 0 0 0 0 0 0 0 0 0 ... .. .. ..@ dimLabels : chr [1:2] "sampleNames" "sampleColumns" .. .. ..@ ..\_classVersion\_:Formal class 'Versions' [package "Biobase"] with 1 slots .. .. .. ..@ .Data:List of 1 .. .. .. .. ..\$ : int [1:3] 1 1 0 ..@ featureData :Formal class 'AnnotatedDataFrame' [package "Biobase"] with 4 slots .. .. ..@ varMetadata :'data.frame': 16 obs. of 3 variables: .. .. .. ..\$ Column : chr [1:16] "ID" "GB\_ACC" "SPOT\_ID" "Species Scientific Name" ... .. .. .. ..\$ Description : Factor w/ 15 levels "", "A gene symbol, when one is available (from UniGene)",...: 3 5 15 13 12 1 11 1 10 14 ... .. .. ..\$ labelDescription: chr [1:16] NA NA NA NA ... .. .. ..@ data :'data.frame': 1000 obs. of 16 variables: .. .. .. ..\$ ID : Factor w/ 22284 levels "1007\_s\_at","1053\_at",...: 1 2 3 4 5 6 7 8 9 10 ... .. .. .. ..\$ GB\_ACC : Factor w/ 21129 levels "AF052179","AF061832",...: 93 30 95 97 25 24 96 99 28 20 ... .. .. .. ..\$ SPOT\_ID : chr [1:1000] NA NA NA NA ... .. .. .. ..\$ Species.Scientific.Name : Factor w/ 2 levels "Homo sapiens",...: 1 1 1 1 1 1 1 1 1 1 ... .. .. .. ..\$ Annotation.Date : Factor w/ 2 levels "Jul 11, 2007",...: 1 1 1 1 1 1 1 1 1 1 ... .. .. .. ..\$ Sequence.Type : Factor w/ 4 levels "Consensus sequence",...: 2 2 2 2 2 2 2 2 2 2 ... .. .. .. ..\$ Sequence.Source : Factor w/ 3 levels "Affymetrix Proprietary Database",...: 1 2 1 2 1 2 1 1 2 1 ... .. .. .. ..\$ Target.Description : Factor w/ 21363 levels "Consensus includes gb:AI656011 /FEA=EST /DB\_XREF=gi:4739990 /DB\_XREF=est:tt42e08.x1 /CLONE=IMAGE:2243462 /UG=Hs.116875 KIAA0156" | \_\_truncated\_\_,...: 16 13 18 20 8 7 19 22 11 4 ... .. .. .. ..\$ Representative.Public.ID : Factor w/ 21197 levels "AF052179","AF061832",...: 93 30 95 97 25 24 96 99 28 20 ... .. .. .. ..\$ Gene.Title : Factor w/ 14208 levels "ADP-ribosylation factor 1",...: 35 66 46 60 44 97 96 64 26 33 ... .. .. .. ..\$ Gene.Symbol : Factor w/ 13293 levels "ABCF1","ARF1",...: 20 59 40 53 33 96 94 58 15 18 ... .. .. .. ..\$ ENTREZ\_GENE\_ID : chr [1:1000] "780" "5982" "3310" "7849" ... .. .. .. ..\$ RefSeq.Transcript.ID : Factor w/ 13074 levels "NM\_000409","NM\_000661" /// NM\_001024921",...: 37 45 41 52 1 50 49 82 47 4 ... .. .. .. ..\$ Gene.Ontology.Biological.Process: Factor w/ 7245 levels "", "0000074 // regulation of progression through cell cycle // traceable author statement /// 0006139 // nucleobase, nucleoside, nu" | \_\_truncated\_\_,...: 61 22 78 32 79 60 14 63 72 20 ... .. .. .. ..\$ Gene.Ontology.Cellular.Component: Factor w/ 4148 levels "", "0000502 // proteasome complex (sensu Eukaryota) // traceable author statement /// 0005634 // nucleus // inferred from electroni" | \_\_truncated\_\_,...: 72 45 1 44 1 1 42 71 6 68 ... .. .. .. ..\$ Gene.Ontology.Molecular.Function: Factor w/ 7314 levels "", "0000049 // tRNA binding // non-traceable author statement /// 0000166 //



**Arguments**

x	object to be exported. Currently methods for objects of class epheno (produced with ExpressionPhenoTest function) are implemented.
file	Name of the file where the results are to be saved
row.names	Passed on to write.csv
...	Other arguments to be passed on to write.csv

---

```
get gseaSignatures' elements
```

*Substract element's of a gseaSignaturesSign or gseaSignaturesVar object*

---

**Description**

getEs returns ES (enrichment scores) getEsNes returns simulated ES (needed to compute pvals) and getFcHr returns the fold changes or hazard ratios used to compute the ES and simulated ES.

**Usage**

```
getEs(x)
getEsSim(x)
getFcHr(x)
```

**Arguments**

x	an gseaSignaturesSign or gseaSignaturesVar object. Those objects are obtained using the gseaSignatures function.
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**Author(s)**

Evarist Planet

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```
getVars2test-methods
```

*Methods for Function getVars2test in Package 'phenoTest'*

---

**Description**

Methods for function getVars2test in Package 'phenoTest'. For more information read the funcion's manual.

**Methods**

signature(x = "epheno") Method for an object of class epheno.

---

```
getVars2test
```

*Get phenotypic variables that were tested.*

---

### Description

Returns an object containing the names of the variables that were tested when the epheno object was created. Will return an object of class list. Variables of the same type (categorical, survival, etc) will be in the same slot of the list. The slot names are the types of the variables.

### Author(s)

Evarist Planet

### Examples

```
data(epheno)
getVars2test(epheno)
```

---

```
gseaSignatures-class
```

*Class "gseaSignatures" ES and EsSim container.*

---

### Description

This object contains de ES (enrichment scores) and simulated ES that will be used in the GSEA (Gene Set Enrichment Analysis) process.

### Objects from the Class

Objects can be created by calls of the form `new("gseaSignatures", ...)`.

### Slots

`.Data`: Object of class "list".

`es`: Object of class "numeric" Contains the observed enrichment scores. The ones that were computed from the data without permuting anything.

`es.sim`: Object of class "numeric" Contains the enrichment score that were obtained after permutations.

`signature`: Object of class "numeric" The subset of genes we are interested in.

### Extends

Class "list", from data part. Class "vector", by class "list", distance 2. Class "AssayData", by class "list", distance 2.

### Methods

No methods defined with class "gseaSignatures" in the signature.

**Author(s)**

Evarist Planet

**Examples**

```
showClass("gseaSignatures")
```

---

gseaSignatures-methods

*Methods for Function gseaSignatures in Package 'phenoTest'*

---

**Description**

Methods for function `gseaSignatures` in Package 'phenoTest'. For more information read the function's manual.

**Methods**

`signature(x = "ANY", signatures = "character")` Method for signature of class `character`.

`signature(x = "ANY", signatures = "GeneSet")` Method for signature of class `character`.

`signature(x = "epheno", signatures = "list")` Method for an `epheno` object and several signatures stored in an object of class `list`.

`signature(x = "matrix", signatures = "GeneSetCollection")` Method for an `matrix` object and several signatures stored in an object of class `GeneSetCollection`.

`signature(x = "epheno", signatures = "GeneSetCollection")` Method for an `epheno` object and several signatures stored in an object of class `GeneSetCollection`.

`signature(x = "numeric", signatures = "GeneSetCollection")` Method for an `numeric` object and several signatures stored in an object of class `GeneSetCollection`.

`signature(x = "matrix", signatures = "list")` Method for an `matrix` object and several signatures stored in an object of class `list`.

`signature(x = "numeric", signatures = "list")` Method for an `numeric` object and several signatures stored in an object of class `list`.

---

<code>gseaSignatures</code>	<i>Compute ES (enrichment scores) and es.sim (simulated enrichment scores)</i>
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**Description**

This function computes the first step in the process of obtaining a GSEA-like plot. It computes the enrichment scores and simulated enrichment scores for each variable and signature. The output will usually be used as input for the `gseaSignificance` function. The `getEs`, `getEsSim`, `getFc`, `getHr` and `getFcHr` methods can be used to access each subobject. For more information please visit the man pages of each method.

**Usage**

```
gseaSignatures(x, signatures, logScale=TRUE, absVals=FALSE, averageRepeats=FALSE, B=1
```

**Arguments**

<code>x</code>	ePhenoTest, numeric or matrix object containing hazard ratios or fold changes.
<code>signatures</code>	character or list object containing the names of the genes that belong to each signature.
<code>logScale</code>	if values should be log scaled.
<code>absVals</code>	if TRUE fold changes and hazard ratios that are negative will be turned into positive before starting the process. This is useful when genes can go in both directions.
<code>averageRepeats</code>	if <code>x</code> is of class numeric and has repeated names (several measures for some individual names) we can average the measures of the same names.
<code>B</code>	number of simulations to perform.
<code>mc.cores</code>	number of processors to use.
<code>test</code>	the test that will be used. 'perm' stands for the permutation based method, 'wilcox' stands for the wilcoxon test (this is the fastest one) and 'tperm' stands for permutation t test.

**Details**

ES score was calculated for each signature and variable (see references). If parameter `test` is 'perm' the signature was permuted and the ES score was recalculated (this happened `B` times for each variable, 1000 by default). If `test` is 'wilcox' a wilcoxon test in which we test the fact that the average value of the genes that do belong to our signature is different from the average value of the genes that do not belong to our signature will be performed. If `test` is 'tperm' a permutation t-test will be used. Take into account that the final plot will be different when a 'wilcox' is used.

**Author(s)**

Evarist Planet

**References**

Aravind Subramanian, (October 25, 2005) *Gene Set Enrichment Analysis*. [www.pnas.org/cgi/doi/10.1073/pnas.0506580102](http://www.pnas.org/cgi/doi/10.1073/pnas.0506580102)

**Examples**

```
#load epheno object
data(epheno)
epheno

#we construct two signatures
sign1 <- sample(featureNames(epheno))[1:20]
sign2 <- sample(featureNames(epheno))[50:75]
mySignature <- list(sign1,sign2)
names(mySignature) <- c('My first signature','My preferred signature')
```

```
#run gsea functions
#my.gseaSignatures <- gseaSignatures(x=epheno,signatures=mySignature,B=100,mc.cores=1)
#my.gseaSignificance <- gseaSignificance(my.gseaSignatures)
#my.summary <- summary(my.gseaSignificance)
#my.summary
#plot(my.gseaSignatures,my.gseaSignificance)
```

---

```
gseaSignaturesSign-class
      Class "gseaSignaturesSign"
```

---

### Description

This class is an ES (enrichment score) and ES.sim (simulated enrichment score) container that will be used in the GSEA (Gene Set Enrichment Analysis) process. There is one container for every gene signature.

### Objects from the Class

Objects can be created by calls of the form `new("gseaSignaturesSign", ...)`.

### Slots

**.Data:** Object of class "list".

**gseaSignatures:** Object of class "gseaSignatures" This is the object that will contain the ES and ES.sim.

**fc.hr:** Object of class "character" fold change or hazard ratio used to compute the enrichment scores.

**s:** Object of class "logical" The subset of genes we are interested in.

**test:** Object of class "character" The statistical test that will be used.

### Extends

Class "list", from data part. Class "vector", by class "list", distance 2. Class "AssayData", by class "list", distance 2.

### Methods

**getEs** signature(x = "gseaSignaturesSign"): Returns the enrichment scores.

**getEsSim** signature(x = "gseaSignaturesSign"): Returns the simulated enrichment scores (the ones obtained after permutations).

**getFcHr** signature(x = "gseaSignaturesSign"): Returns the fold change and/or the hazard ratio that were used to compute the enrichment scores.

**gseaSignificance** signature(x = "gseaSignaturesSign"): This is the next step in the process of performing GSEA. This function will test if the gene sets are enriched.

### Author(s)

Evarist Planet

**Examples**

```
showClass("gseaSignaturesSign")
```

---

```
gseaSignaturesVar-class
  Class "gseaSignaturesVar"
```

---

**Description**

This class is an ES (enrichment score) and ES.sim (simulated enrichment score) container that will be used in the GSEA (Gene Set Enrichment Analysis) process. There is one container for every phenotype. Every one of this containers (of class `gseaSignaturesSign`) is a container itself and has the enrichment scores of all signatures. `GseaSignaturesVar` contains one element per phenotype (phenotypic variable). Every one of this elements is of class `gseaSignaturesSign` and contains one element per signature.

**Objects from the Class**

Objects can be created by calls of the form `new("gseaSignaturesVar", ...)`.

**Slots**

`.Data`: Object of class "list".

`gseaSignatures`: Object of class "gseaSignaturesSign" This object contains the enrichment scores and other elements that will be used in the GSEA process.

**Extends**

Class "`list`", from data part. Class "`vector`", by class "list", distance 2. Class "`AssayData`", by class "list", distance 2.

**Methods**

**getEs** signature (x = "gseaSignaturesVar"): Returns the enrichment scores.

**getEsSim** signature (x = "gseaSignaturesVar"): Returns the simulated enrichment scores (the ones obtained after permutations).

**getFcHr** signature (x = "gseaSignaturesVar"): Returns the fold change and/or the hazard ratio that were used to compute the enrichment scores.

**gseaSignificance** signature (x = "gseaSignaturesVar"): This is the next step in the process of performing GSEA. This function will test if the gene sets are enriched.

**Author(s)**

Evarist Planet

**Examples**

```
showClass("gseaSignaturesVar")
```

---

`gseaSignificance` *ES' (enrichment scores) significance.*

---

## Description

This function performs the second step in the process of obtaining a GSEA-like plot. It computes the NES (normalized enrichment score), p values and fdr (false discovery rate) for all variables and signatures. A `gseaSignaturesSign` or `gseaSignaturesVar` object will be needed as input (these objects can be obtained with the `gseaSignatures` function). For an overview of the output use the `summary` method. The next step after using the `gseaSignificance` function would be using the `plot` method.

## Usage

```
gseaSignificance(x, p.adjust.method='none', pval.comp.method='original', pval.smooth
```

## Arguments

`x` `gseaSignaturesSign` or `gseaSignaturesVar` object obtained with the `gseaSignatures` method. This object contains the enrichment scores, the simulated enrichment scores and the fold changes or hazard ratios.

`p.adjust.method` p adjustment method to be used. Common options are 'BH', 'BY', 'bonferroni' or 'none'. All available options and their explanations can be found on the `p.adjust` function manual.

`pval.comp.method` the p value computation method. Has to be one of 'signed' or 'original'. The default one is 'original'. See details for more information.

`pval.smooth.tail` if we want to estimate the tail of the ditribution where the pvalues will be generated.

## Details

The simulated enrichment scores and the calculated one are used to find the p value.

P value calculation depends on the parameter `pval.comp.method`. The default value is 'original'. In 'original' we are simply computing the proportion of absolute simulated ES which are larger than the observed absolute ES. In 'signed' we are computing the proportion of simulated ES which are larger than the observed ES (in case of having positive enrichment score) and the proportion of simulated ES which are smaller than the observed ES (in case of having negative enrichment score).

## Author(s)

Evarist Planet

## References

Aravind Subramanian, (October 25, 2005) *Gene Set Enrichment Analysis*. [www.pnas.org/cgi/doi/10.1073/pnas.0506580102](http://www.pnas.org/cgi/doi/10.1073/pnas.0506580102)

C.A. Tsai and J.J. Chen. *Kernel estimation for adjusted p-values in multiple testing*. *Computational Statistics & Data Analysis* [http://econpapers.repec.org/article/eeecsdana/v\\_3a51\\_3ay\\_3a2007\\_3ai\\_3a8\\_3ap\\_3a3885-3897.htm](http://econpapers.repec.org/article/eeecsdana/v_3a51_3ay_3a2007_3ai_3a8_3ap_3a3885-3897.htm)

## Examples

```
#for examples see the help file of gseaSigntaures: ?gseaSignatures
```

---

```
gseaSignificanceSign-class
      Class "gseaSignificanceSign"
```

---

## Description

This object contains the results of the test of enrichment that was performed on each gene set. There is one container for every gene signature.

## Objects from the Class

Objects can be created by calls of the form `new("gseaSignificanceSign", ...)`.

## Slots

`.Data`: Object of class "list".

`gseaSignificance`: Object of class "matrix" Contains the statistics. Use the summary method to access this information.

`p.adjust.method`: Object of class "character" The p-value adjustment method that was used.

## Extends

Class "list", from data part. Class "vector", by class "list", distance 2. Class "AssayData", by class "list", distance 2.

## Methods

No methods defined with class "gseaSignificanceSign" in the signature.

## Author(s)

Evarist Planet

## Examples

```
showClass("gseaSignificanceSign")
```

---

```
gseaSignificanceVar-class  
  Class "gseaSignificanceVar"
```

---

### Description

This object contains the results of the test of enrichment that was performed on each gene set and phenotype. There is one container for every phenotype. Every one of this containers (of class `gseaSignificanceSign`) is a container itself and has the results of the tests for all signatures. `GseaSignificanceVar` contains one element per phenotype (phenotypic variable). Every one of this elements is of class `gseaSignificanceSign` and contains one element per signature.

### Objects from the Class

Objects can be created by calls of the form `new("gseaSignificanceVar", ...)`.

### Slots

`.Data`: Object of class "list".

`gseaSignificance`: Object of class "gseaSignificanceSign" This object contains the results of the tests.

### Extends

Class "`list`", from data part. Class "`vector`", by class "list", distance 2. Class "`AssayData`", by class "list", distance 2.

### Methods

No methods defined with class "gseaSignificanceVar" in the signature.

### Author(s)

Evarist Planet

### Examples

```
showClass("gseaSignificanceVar")
```

---

```
heatmapPhenoTest-methods  
  Methods for Function heatmapPhenoTest in Package 'phenoTest'
```

---

### Description

Methods for function `heatmapPhenoTest` in Package 'phenoTest'. For more information read the function's manual.

## Methods

`signature(x = "ExpressionSet", signatures = "character")` Method for an ExpressionSet object and one signature stored in an object of class character.

`signature(x = "ExpressionSet", signatures = "list")` Method for an ExpressionSet object and several signatures stored in an object of class list.

`signature(x = "ExpressionSet", signatures = "missing")` Method for an ExpressionSet object and no signatures.

`signature(x = "ExpressionSet", signatures = "GeneSet")` Method for an ExpressionSet object and one signature stored in an object of class GeneSet.

`signature(x = "ExpressionSet", signatures = "GeneSetCollection")` Method for an ExpressionSet object and several signatures stored in an object of class GeneSetCollection.

---

`heatmapPhenoTest` *Produce heatmap from phenotype data.*

---

## Description

Show the associations between clusters that each sample belongs to and each phenotype in a heatmap and/or a Kaplan-Meier plot.

## Usage

```
heatmapPhenoTest(x, signatures, vars2test, probes2genes = FALSE,
  filterVar, filteralpha = 0.05, distCol = "pearson", nClust = 2, distRow = "cor",
  p.adjust.method = "none", simulate.p.value = FALSE, B = 10^5, linkage = "average",
  equalize = FALSE, center = TRUE, col, survCol, heat.kaplan="both", ...)
```

## Arguments

<code>x</code>	ExpressionSet with phenotype information stored in <code>pData(x)</code> .
<code>signatures</code>	Either character vector or list of character vectors with gene sets to be used to draw heatmaps (gene names should match those in <code>featureNames(x)</code> ). A separate heatmap will be produced for each element in the list.
<code>vars2test</code>	list with components 'continuous', 'categorical', 'ordinal' and 'survival' indicating which phenotype variables should be tested. 'continuous', 'categorical' and 'ordinal' must be character vectors, 'survival' a matrix with columns named 'time' and 'event'. The names must match names in <code>names(pData(x))</code> .
<code>probes2genes</code>	If set to <code>TRUE</code> a single probe is selected for each gene. <code>nsFilter</code> is used to select the probe with highest inter-quartile range.
<code>filterVar</code>	If specified, only genes with significant differences in the variable <code>filterVar</code> will be displayed in the heatmap. Note that this option will not affect the sample clustering, as this is obtained using both significant and non-significant genes.
<code>filteralpha</code>	Significance level for the filtering based on <code>filterVar</code> .
<code>distCol</code>	Distance metric used to cluster columns (e.g. patients/samples). Can take any value accepted by <code>dist</code> . Pearson and Spearman correlations are also allowed. Write 'spearman' or 'pearson' to use them.

nClust	Number of desired clusters.
distRow	Distance metric used to cluster rows (e.g. genes). Can take any value accepted by <code>distancematrix</code> .
p.adjust.method	Method for P-value adjustment, passed on to <code>p.adjust</code> .
simulate.p.value	If set to FALSE the chi-square test p-values are computed using asymptotics, otherwise a simulation is used (see <code>chisq.test</code> for details).
B	An integer specifying the number of replicates used in the chi-square Monte Carlo test (passed on to <code>chisq.test</code> ).
linkage	Linkage used for clustering. Must be either 'complete', 'average' or 'minimum'.
equalize	Should color codes be equalized between genes, i.e. all genes present the same range of colors. Passed on to <code>heatmap_plus</code> .
center	centering is done by subtracting the column means (omitting NAs).
col	Color scheme to be used for heatmap. Defaults to a green/red scheme designed to look nice for microarray data.
survCol	Colors for the Kaplan-Meier survival curves.
heat.kaplan	can be "heat" if we want to plot a heatmap, "kaplan" if we want to plot a kaplan-meier or "both" if we want both of them.
...	Other arguments for the survival plot, e.g. <code>lty</code> etc.

### Details

Makes two clusters of samples based on the expression levels of the genes from the given signature and plots a heatmap and/or a Kaplan-Meier showing the association between belonging to one cluster or the other and each phenotype.

For variables in `vars2test` continuous and `vars2test` ordinal a Kruskal-Wallis Rank Sum test is used; for `vars2test` categorical a chi-square test (with exact p-value if `simulate.p.value` is set to TRUE); for `vars2test` survival a Cox proportional hazards likelihood-ratio test.

### Author(s)

David Rossell

### Examples

```
#load data
data(eset)
eset

#construct vars2test
survival <- matrix(c("Relapse", "Months2Relapse"), ncol=2, byrow=TRUE)
colnames(survival) <- c('event', 'time')
vars2test <- list(survival=survival)
vars2test

#construct a signature
sign <- sample(featureNames(eset)) [1:20]

#make plot
heatmapPhenoTest(eset, sign, vars2test=vars2test, heat.kaplan='heat')
heatmapPhenoTest(eset, sign, vars2test=vars2test, heat.kaplan='kaplan')
```

---

htmlpage	<i>htmlpage</i>
----------	-----------------

---

**Description**

This is a copy of the `htmlpage` function from `annotate` package. The copy was made to allow `htmlpage` to produce links to local files.

**Details**

See `?annotate::htmlpage` for more information.

---

pAdjust-methods	<i>Methods for Function pAdjust in Package 'phenoTest'</i>
-----------------	--

---

**Description**

Methods for function `pAdjust` in Package 'phenoTest'. This function adjusts the p-values of an `ePheno` object. For more information read the function's manual.

**Methods**

`signature(x = "ePheno")` Adjusts the p-values of an `ePheno` object.

**Author(s)**

Evarist Planet

---

pAdjust	<i>Adjust p values of an ePheno object.</i>
---------	---

---

**Description**

Adjusts the p values of an `ePheno` object. The `p.adjust` function will be used. For more information read the `p.adjust` function's help.

**Usage**

```
pAdjust(x, method = "BH")
```

**Arguments**

<code>x</code>	an <code>ePheno</code> object.
<code>method</code>	the correction method that will be used. See the <code>p.adjust</code> help for more info about the methods.

**Author(s)**

Evarist Planet

**Examples**

```
#load epheno object
data(epheno)
epheno

#Adjust pvalue
p.adjust.method(epheno)
epheno <- pAdjust(epheno, method='BH')
p.adjust.method(epheno)
```

---

phenoTest-package *Test correlation between gene expression and phenotype.*

---

**Description**

Test correlation between gene expression and phenotype.

**Details**

Package:	phenoTest
Type:	Package
Version:	1.0
Date:	2010-04-28
License:	What license is it under?
LazyLoad:	yes

**Author(s)**

Evarist Planet Maintainer: Evarist Planet <evarist.planet@irbbarcelona.org>

---

plot.gseaSignatures  
*GSEA-like Plot.*

---

**Description**

Builds a GSEA plot using a gseaSignature object (one of gseaSignaturesSign or gseaSignaturesVar obtained with the gseaSignatures function) and a gseaSignificance object (one of gseaSignificanceSign or gseaSignificanceVar obtained with the gseaSignificance function).

**Usage**

```
plot.gseaSignaturesSign(x, gseaSignificance, es.ylim, nes.ylim, es.nes="both", ...)
plot.gseaSignaturesVar(x, gseaSignificance, es.ylim, nes.ylim, es.nes="both", ...)
```

**Arguments**

`x` object of class `gseaSignaturesSign` or `gseaSignatures Var`.  
`gseaSignificance` object of class `gseaSignificanceSign` or `gseaSignificanceVar`.  
`es.ylim` ylim values for the ES plot.  
`nes.ylim` ylim values for the NES plot.  
`es.nes` can be "es" if we want to plot enrichment score, "nes" if we want to plot normalised enrichment scores or "both" if we want to plot them both.  
`...` Arguments to be passed to `plot`.

**Author(s)**

Evarist Planet

**References**

Aravind Subramanian, (October 25, 2005) *Gene Set Enrichment Analysis*. [www.pnas.org/cgi/doi/10.1073/pnas.0506580102](http://www.pnas.org/cgi/doi/10.1073/pnas.0506580102)

**Examples**

```
#for examples see the help file of gseaSignatures: ?gseaSignatures
```

---

barplotSignatures *Summary plots for gene signature vs phenotype association*

---

**Description**

Summarizes the univariate relationships between genes in one or more signatures and several phenotype variables, as summarized in `ePheno` objects (which can be created with the `ExpressionPhenoTest` function).

By default `barplotSignifSignatures` performs a binomial test (`binom.test` from package `stats`) for each signature to see if the number of up up regulated and down regulated genes is different enough to be statistically different. When a reference gene set is provided we test if the proportions of up and down regulated genes of each gene set is different from the proportions in the reference gene set. This has been done with a chi-square test. When a reference gene set is provided and parameter `testUpDown` is `TRUE` (by default its `FALSE`) the number of genes corresponding to up and down regulated are compared with those of the reference gene set separately.

**Usage**

```
barplotSignatures(x, signatures, referenceSignature, alpha=.05,
p.adjust.method='none', ylab, cex.text=1, ...)
barplotSignifSignatures(x, signatures, referenceSignature, testUpDown=FALSE,
simulate.p.value = FALSE, B = 10^4, p.adjust.method='none', alpha=.05,
ylab, ylim=ylim, cex.text=1, ...)
```

**Arguments**

<code>x</code>	epheno object, as returned by <code>ExpressionPhenoTest</code> .
<code>signatures</code>	List with each element corresponding to a signature. The gene names in each signature must match those in <code>epheno</code> .
<code>referenceSignature</code>	If specified, the average fold change in each signature is compared to the average fold change in the signature <code>referenceSignature</code> .
<code>testUpDown</code>	If set to <code>TRUE</code> , bars corresponding to up and down-regulated genes are compared with those of <code>referenceSignature</code> separately. This argument is ignored if <code>referenceSignature</code> is not specified.
<code>cex.text</code>	Character expansion for the text indicating the P-values. Ignored if <code>referenceSignature</code> is missing.
<code>alpha</code>	Confidence levels for barplot error bars.
<code>p.adjust.method</code>	P-value adjustment method, passed on to <code>p.adjust</code> .
<code>simulate.p.value</code>	A logical indicating whether chi-square p-values should be computed by Monte Carlo simulation (passed on to <code>chisq.test</code> ).
<code>B</code>	Integer specifying the number of replicates in the Monte Carlo simulation (passed on to <code>chisq.test</code> ).
<code>ylab</code>	y-axis labels
<code>ylim</code>	y-axis limits
<code>...</code>	Other arguments to be passed on to <code>boxplot</code> .

**Value**

When a single signature is provided as input, a single plot assessing the association of that signature with all phenotype variables is created. If several signatures are provided, one separate plot is created for each phenotype variable.

**Author(s)**

Evarist Planet

**Examples**

```
#create epheno
data(epheno)

#construct two signatures
sign1 <- sample(featureNames(epheno)) [1:20]
sign2 <- sample(featureNames(epheno)) [1:15]
mySignature <- list(sign1, sign2)
names(mySignature) <- c('My first signature', 'My preferred signature')

#plot
barplotSignifSignatures(epheno[, 'Relapse'], mySignature, alpha=0.05)
```

---

 show-methods

*Methods for Function show in Package 'methods'.*


---

## Description

Methods for function show in Package 'methods'.

## Methods

signature(object = "AnnotatedDataFrame") Will show an object of class AnnotatedDataFrame.

signature(object = "ANY") Will show an object of class ANY.

signature(object = "classRepresentation") Will show an object of class classRepresentation.

signature(object = "container") Will show an object of class container.

signature(object = "epheno") Will show an object of class epheno.

signature(object = "eSet") Will show an object of class eSet.

signature(object = "genericFunction") Will show an object of class genericFunction.

signature(object = "gseaSignaturesSign") Will show an object of class gseaSignaturesSign.

signature(object = "gseaSignaturesVar") Will show an object of class gseaSignaturesVar.

signature(object = "gseaSignificanceSign") Will show an object of class gseaSignificanceSign.

signature(object = "gseaSignificanceVar") Will show an object of class gseaSignificanceVar.

signature(object = "LargeDataObject") Will show an object of class LargeDataObject.

signature(object = "MethodDefinition") Will show an object of class MethodDefinition.

signature(object = "MethodSelectionReport") Will show an object of class MethodSelectionReport.

signature(object = "MethodWithNext") Will show an object of class MethodWithNext.

signature(object = "MIAME") Will show an object of class MIAME.

signature(object = "namedList") Will show an object of class namedList.

signature(object = "ObjectsWithPackage") Will show an object of class ObjectsWithPackage.

signature(object = "oldClass") Will show an object of class oldClass.

signature(object = "ScalarCharacter") Will show an object of class ScalarCharacter.

signature(object = "ScalarObject") Will show an object of class ScalarObject.

signature(object = "signature") Will show an object of class signature.

signature(object = "TestResults") Will show an object of class TestResults.

signature(object = "traceable") Will show an object of class traceable.

signature(object = "Versioned") Will show an object of class Versioned.

signature(object = "Versions") Will show an object of class Versions.

signature(object = "VersionsNull") Will show an object of class VersionsNull.

---

smoothCoxph                      *Plots the Cox proportional hazard smoothed by gene expression level.*

---

### Description

Builds a plot showing how hazard behaves over different levels of expression of a given gene. Confidence intervals are also provided.

### Usage

```
smoothCoxph(time, event, x, xlim, ylim, ...)
```

### Arguments

time	variable where time to survival is stored.
event	variable where survival event is stored.
x	numeric containing the expression levels of a given gene.
xlim	xlim for the plot.
ylim	ylim for the plot.
...	other arguments that will be passed to plot.

### Author(s)

David Rossell.

### Examples

```
#load eset
data(eset)

#make plot
smoothCoxph(pData(eset)$Months2Relapse, pData(eset)$Relapse, exprs(eset)[25,])
```

---

summary.gseaSignificance  
*Obtain a data.frame with the pvalues and fdr for all signatures and*

---

### Description

Builds a data.frame object that can easily be written to a csv file containing the ES, NES, pval.ES, pval.NES and FDR.

### Usage

```
summary.gseaSignificanceSign(object, ...)
summary.gseaSignificanceVar(object, ...)
```

**Arguments**

object            object of class gseaSignificanceSign or gseaSignificanceVar.  
...               Arguments to be passed to summary.

**Author(s)**

Evarist Planet

**References**

Aravind Subramanian, (October 25, 2005) *Gene Set Enrichment Analysis*. [www.pnas.org/  
cgi/doi/10.1073/pnas.0506580102](http://www.pnas.org/cgi/doi/10.1073/pnas.0506580102)

**Examples**

```
#for examples see the help file of gseaSigntaures: ?gseaSignatures
```

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